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GENETICS

126

Cistron analysis of Lac loci using Lac$_2$-F$_{13}$.

29/VII 1959

REF: W3112 F$_{13}$.

Plate	1	2	3	4	5	6	7	8	9	10
Lac$^-$			× 3112 F$_{13}$ (Lac$^-$)	× 3112 (Lac$^-$)	× 3747 (Lac$^+$ π-λ_2)	blank				
DNPG	3112	2	—	—	+	—				
unstable	3127	4	+	+	+	+ unstable				
	2247	7	—	—	— ?	—			non-transducible Lac.	
+ m unstable	3230	11a	+	+	+	+ many unstable				
+ m	3089	11b	+	—	+ ?	+				
+	4112	52	+	—	+	+				
− f	4128	61	+	—	+	—				
− m	4147	87	+	—	+	—				
− m	4148	88	+	—	+	—				
− m	4149	89	+	—	+	—				
− m	4150	90	+	—	+	—				
− m	4151	91	+	—	+	—				
− m	4153	93	+	—	+	—				
	2243	Lac$_2$	+	+	+	+ unstable				
	2245	5	+	+	+	+				
	2247	Lac$_7$ − also Gal−	−	−	−	+4				

Lac$_2$ Hfr.

Lac$_2$ 3164 nr.

Lac$_{61}$ 4021

Test segregants.

Reaction is very slow.

This black spot gives many lac$^-$ on Blac after streaking.

non-transducible Lac

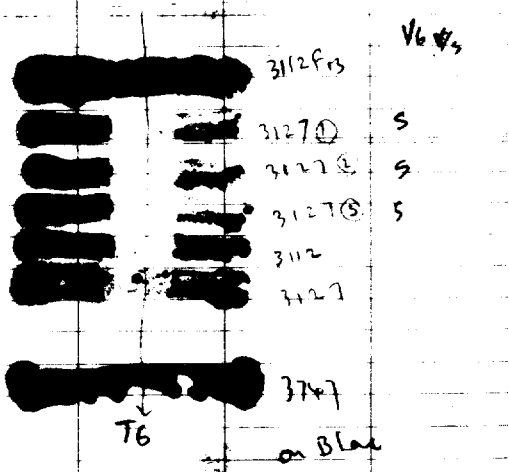
λ_2^s non-transducible Gal

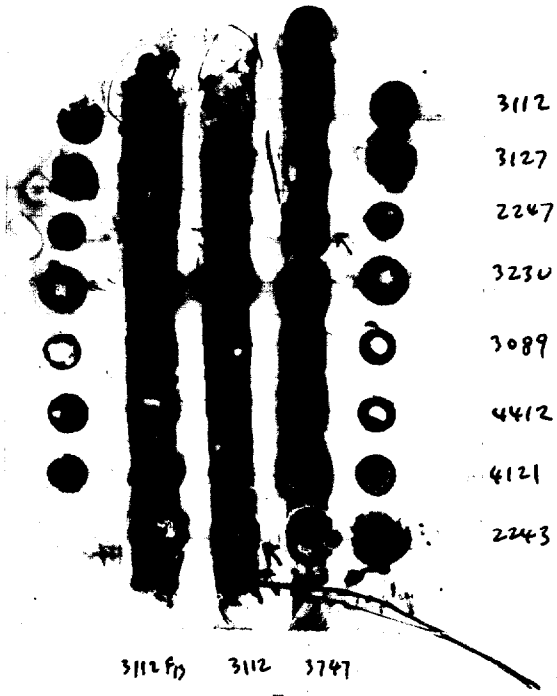
Method: Spot method : on MLac. × various Lac$^-$ mutants. (λ^+ Lac$_\lambda^-$)

culturel age : overnight culture + Pen. 5 mt. at 37°c.

W3112 F$^+$ Lac$_2$ χ^+ ——, Cure.

Result:





	Lac
3112	2
3127	4
2247	7
3230	1a
3089	1b
4412	52
4121	61
2243	3

3112 F⁻ 3112 3747

F⁻ Lac F⁻
M⁻

Segregating between two different Lac⁻.

4147	
4148	
4149	
4150	
4153	
2295	
2247	7

3112 F⁻ 3112 3747

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127 b

Stability of Lac^+ of the treated and untreated heterozygous diploid with AO.

28/VII ; 1959 REF:

	1 AO treated Lac+	2 Lac+ stability T6	3 T6	4 Lac+ Untreated T6	5	6 Lac+ T6	7	8	9	10
1			51 U m	1 U m	51	U m				
2	U	m control	52 U m	2 S r	52	U m				
3	U	m 2596	53 U m	3 U m	53	U m				
4	U	m	54 U m	4 U m	54	U m				
5	U	m	55 S S r	5 U m	55	U m				
6	F⁻ S S	s .	56 U m	6 U m	56	U m				
7	U	m	57 S S m	7 U m	57	U m				
8	U	m	58 S S S	8 U m	58	U m				
9	U	m	59 S S r	9 U m	59	U m				
10	F⁻ S S	s .	F⁻ 60 S S	10 U m	60	U m				
11	U	m	61 U m	11 U m	61	U m				
12	U	m	62 U m	12 U m	62	U m				
13	U	m	63 U m	13 U m	63	U m				
14	U	m	64 U m	14 U m	64	U m				
15	U	m	65 U m	15 U m	65	U m				
16	U	m	66 U m	16 U r	66	S S r				
17	F⁻ U	m	67 U m	17 S control r 3747	67	U m				
18	U	m control	68 U m	18 U m	68	U m				
19	U	m 2596	F⁻ 69 S S S	19 U m	69					
20	U	m	70 U m	20 U m	70					
			F⁻ 71 S S r							
21	U	m		21 U m						
22	U	m	Lac+ (stable or unstable)	22 S S r		Lac+ (stable, or unstable)				
23	U	m	S : 21	23 U m		S : 64				
24	F⁻ S	s .	U : 48	24 U m		U : 63				
25	F⁻ S S	s .		25 U m						
26	S S	r		26 U m						
27	U	m		27 U m						
28	U	m	T6-resistance	28 U m		T6 resistance (Lac+ V6R)	%			
29	F⁻ S S	s	%	29 U m		V6R : 5/67	7.46			
30	S S	r	11.6 V6R : 8/69	30 U m		V6S : 0/67	0			
31	U	m	18.8 V6S : 13/69	31 U m		V6mix : 62/67	92.7			
32	U	m		32 S r						
33	F15 S S	r	88.2 V6mix : 58/69	33 U m						
34	U	r		34 U m						
35	F⁻ S S	r		35 U m						
36	U	m		36 U m						
37	U	m		37 U m						
38	U	m		38 U m						
39	U	m		39 U m						
40	U	m		40 U m						
41	U	m		41 U m						
42	U	m		42 U m						
43	U	m		43 U m						
44	U	m		44 U m						
45	F⁻ S S	s .		45 U m						
46	F⁻ S S	s .		46 U m						
47	F⁻ S S	s .		47 U m						
48	U	m		48 U m						
49	S	s		49 U m						
50	S S	s .		50 U m						

(colonie)
Sf. • Lac^+ found it is noted as unstable. { U: Unstable; S: stable.

Method ①: pick Lac^+ from streaked plate, inculated into broth (1 ml), and incubated them for overnight.

② Streak on Blac, and see is there Lac^+ only or Lac^+ and Lac^- colonie on it.

Lac^+

1. AO-treatment: AO 30 γ/ml.
   - NSB - pH. 7.6.
     One drop size one small loop of ca 10^7 cells/ml / 5 ml NSB
     40
   - The treated culture was streaked on Blac after 24hr incubation at 37°C.

m : mixture of V_6^R and V_6^S, segregated from V_6 R/s.

S : V_6^S

r : V_6^R

U : Unstable for Lac^+ accounted for Lac diploidy.

S : Stable for Lac^+

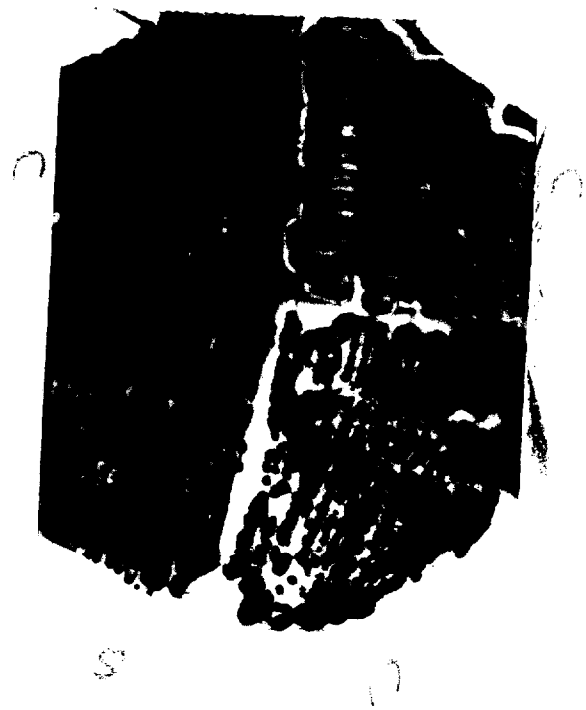
Conclusion treated with AO

1. Almost of the F^- Lac-stable, V_6^R or V_6^S, and Lac^- stable.
   11/88 : F^- Lac^{+st} V_6^R or V_6^S.
   #17 1/12 : F^- Lac^{mix} V_6^m.

2. Lac^{st} is T_6^R or T_6^S, except $T_6^{r/s}$.

This means, AO removes full segment of FLacV at a time.

U : unstable
S : stable



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127a

Elimination of Lac - F13 segment with AO-treatment

23/IV ; 1959

REF: Cf. P 118

Experimental conditions.

Inoculum size : ca. 10^8 cells /ml. AO : 30γ /ml. Med: NSB for AO-T.
Time : overnight at 37°C. Strain treated W2594 F13 (M S^R V_S^R Lac_1^+ V_G^R/S.)

Method : ① Suspend a single colony into 1.0 ml of H_2O, and ~~dilute~~ to use the 10^{-6} ml.
Add 0.1 ml to 5 ml NSB with $AO_{30\gamma}$ and without AO.
② Seed ca 10^8 /ml on to BLac. 5×10^{-7} ml of untreated control.

Result I. Elimination of Lac^+ segment from Lac^+/Lac^- heterozygotes.

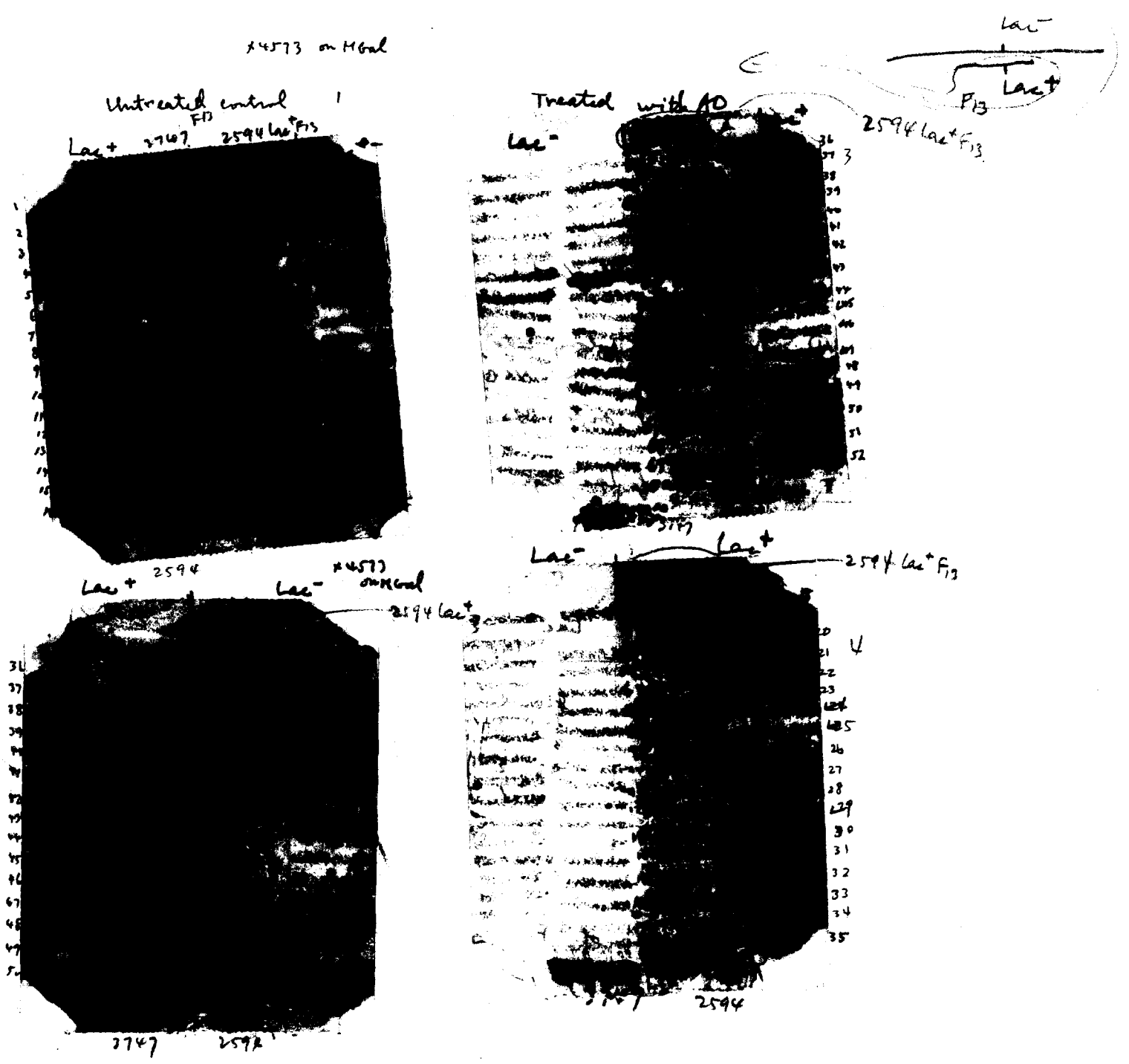
	AO-treated (30γ/ml)						Untreated control			
	I				II		C			
Plate #	Lac^+	Lac^-	Lacv	Σ (%)	Lac^+	Lac^-	Lacv	Σ (%)	Lac^+	Lac^-	Lacv	Σ (%)
1	310	131	4		439	42	2		103	311	2	
2	275	100	2		383	34	3		86	246	2	
3	284	120	3		398	35	2		109	298	3	
4	274	95	2		453	43	2		119	297	3	
5	283	138	2		422	33	1		84	322	2	
Σ												
%												

Results II. Elimination of F13 from Lac^+/Lac^- heterozygotes.

	AO-treated (from #II)				Untreated control			
	Lac^+		Lac^-		Lac^+		Lac^-	
	F_{13} Hi	F_{13} Lo	F^-	F_{13}	F^-	F_{13} Hi	F^-	F_{13} Hi	F_{13} Lo	F^-
	59	0	13	0	64	67	0	22	50	0
Σ										
%										

(Tested by bal-transfer. × 4573)

(see back page)



4421 (Lac61 F⁻)

3112 F_{13} [redacted]

3112 F⁻

on Mac

24 hrs.

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128a

Multiplication of Gal- F_8 segment.

W4520
W6F8 × W4573.

30/4 1959 REF: p. 127

Overnight culture
0.2 ml / 10 ml plm.

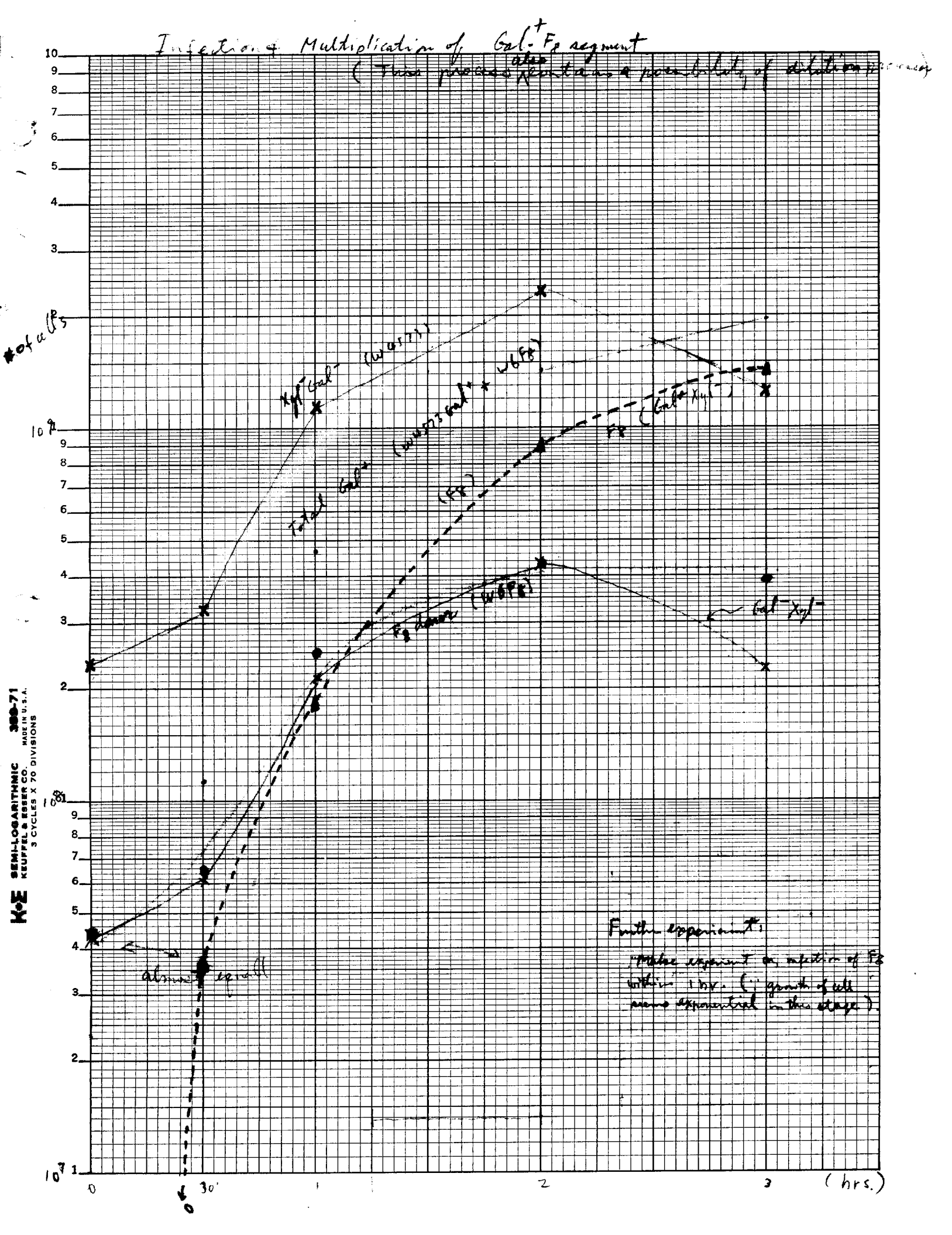
Incubate it at 37°C
on rotator.
for 2 hrs.

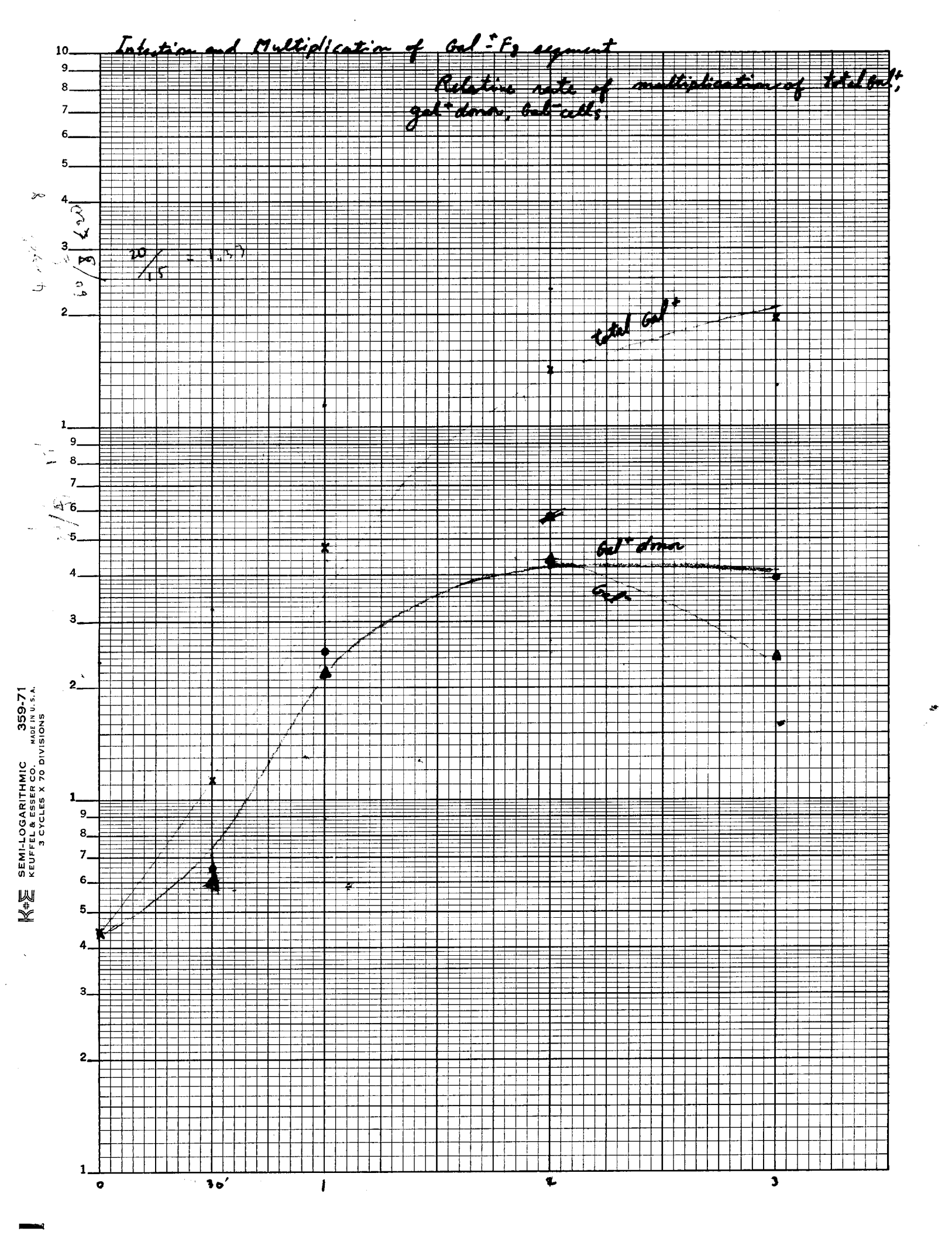
Mix { 1 ml W6F8
+
10 ml W4573.

		AM				PM									
		12:00		12:30		1:00		2:00		3:00					
Inoculum size (ml.)		10^{-6} ml		10^{-6} ml		10^{-6} ml		4×10^{-7}		2×10^{-7}		10^{-7}		10^{-7}	
Time	Vol	0		30'		1 hr		2 hr.		3 hr		4 hr		5 hr	
		+	−	+	−	+	−	+	−	+	−	+	−		
on Bact. Gal	1	8	54	27	92(1)	84	224(7)	101	167(10)	61	62(6)				
	2	6	36	68	88(1)	90	238(6)	108	157(9)	69	88(5)				
	3	10	49	19	101(6)	78	211(12)	113	224(10)	68	76(6)				
	4	4	48	29	72(2)	84	258(4)	99	196(11)	103	74(8)				
	5	16	46	12	59(4)	104	229(3)	105	181(4)	60	59(3)				
	Σ	+V 44 (44)	− 233 277	+V 112 (101,11)	− 322 334	+V 472 (440,32)	− 1160 1632	+V 570 (528,44)	− 925 1820 1495 ×2.5	+V 389 (361,28)	− 359 1985 748 ×2.5				
		0		0		0		0	2310	0	1290				
on Bact. growth of W6F8	Xyl	+	−	+	−	+	−	+	−	+	−				
		8	54	18	108	53		35		8					
		6	36	13	88	51		39		14					
		10	49	11		43		35		16					
		4	48	18		43		31		14					
		16	46	5		60		29	×2.5	26	×5				
% of Lac+	Σ	44	233	65		250		169	422	78	390				
		15.9		19.5		15.3		12.1		10.4					
multiplication of Lac+	Xyl	+		+		+		+		+					
		0		9		29		66		53					
		0		1		29		69		55					
		0		8		35		78		52					
		0		11		41		68		89					
		0		7		44		76		34					
% of Lac+	Σ	0	233	36		188		357	892	283	1414				
		0		10.8		11.5		23.8		37.8					

D : actually counted number

57

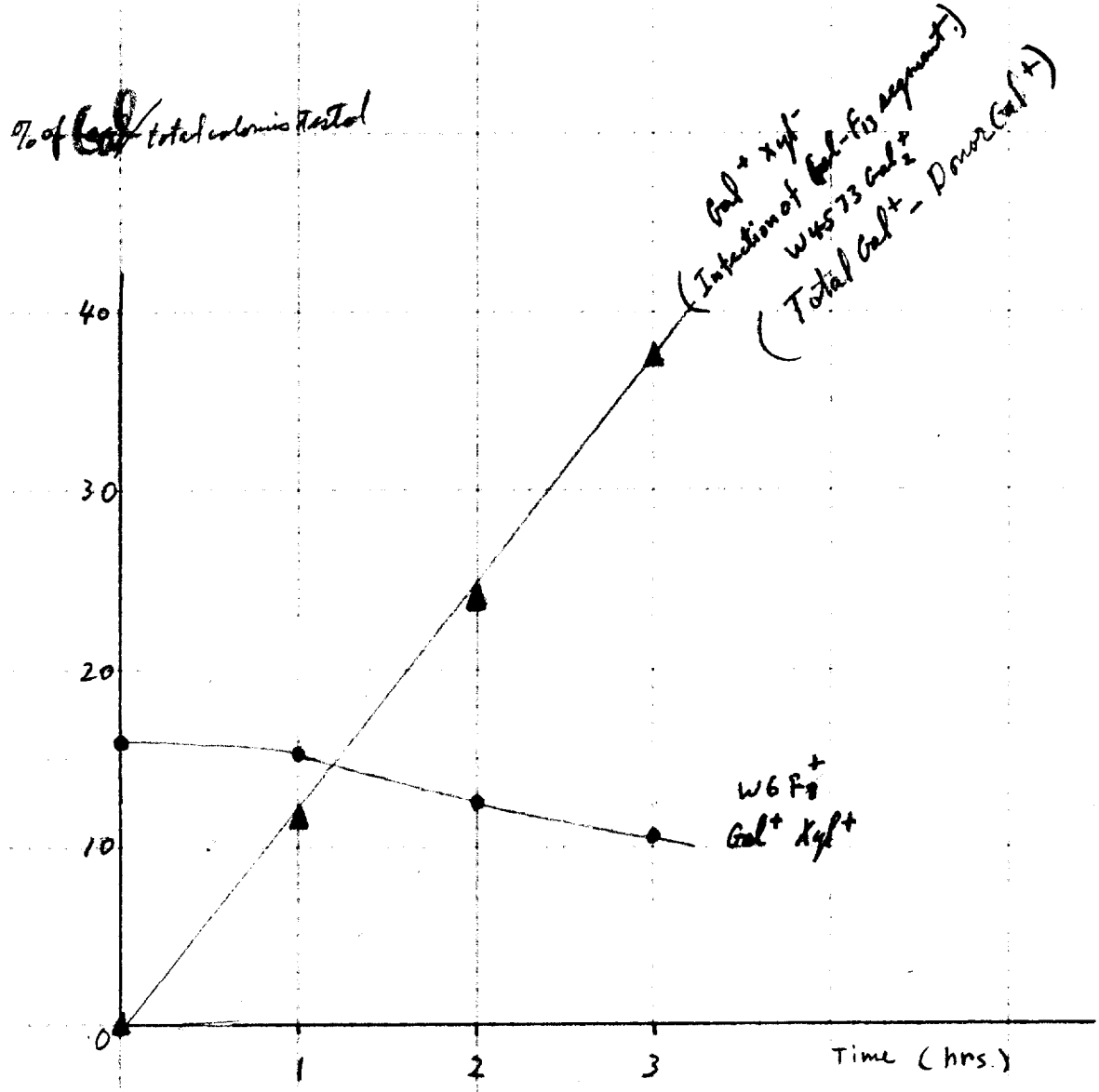

Infection & Multiplication of Gal⁺ Fg segment
(Time process after Kontact a non-lab. of dilution)
# of cells
hrs.
Gal⁻ Xyl⁺
Gal⁺ Xyl⁻
Fg
Fg Gal⁻
Fg Gal⁺
Fg Gal⁻ Xyl⁺
Gal⁻ Xyl⁺
almost equal
Further experiment:
Make experiment on infection of Fg within 1 hr. (: growth of cell more exponential in this stage).
SEMI-LOGARITHMIC 359-71
KEUFFEL & ESSER CO. MADE IN U.S.A.
3 CYCLES X 70 DIVISIONS

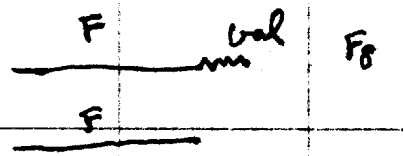

Intestine and Multiplication of Gal⁻ F₂ segment
Relative rate of multiplication of total Gal⁺, Gal⁺ donor, Gal⁻ cells;
total Gal⁺
Gal⁺ donor
Gal⁻
SEMI-LOGARITHMIC 359-71
KEUFFEL & ESSER CO. MADE IN U.S.A.
3 CYCLES X 70 DIVISIONS
0
30'
1
2
3

Rate of multiplication of Gal-F_8 segment. 128L

$W6\,F_8$ — × W4573

37°C, on rotator,
, in penassay broth,
, exponentially growing cultures




F
Wal
F_B
F

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Multiplication of F8 Gal loci.

129

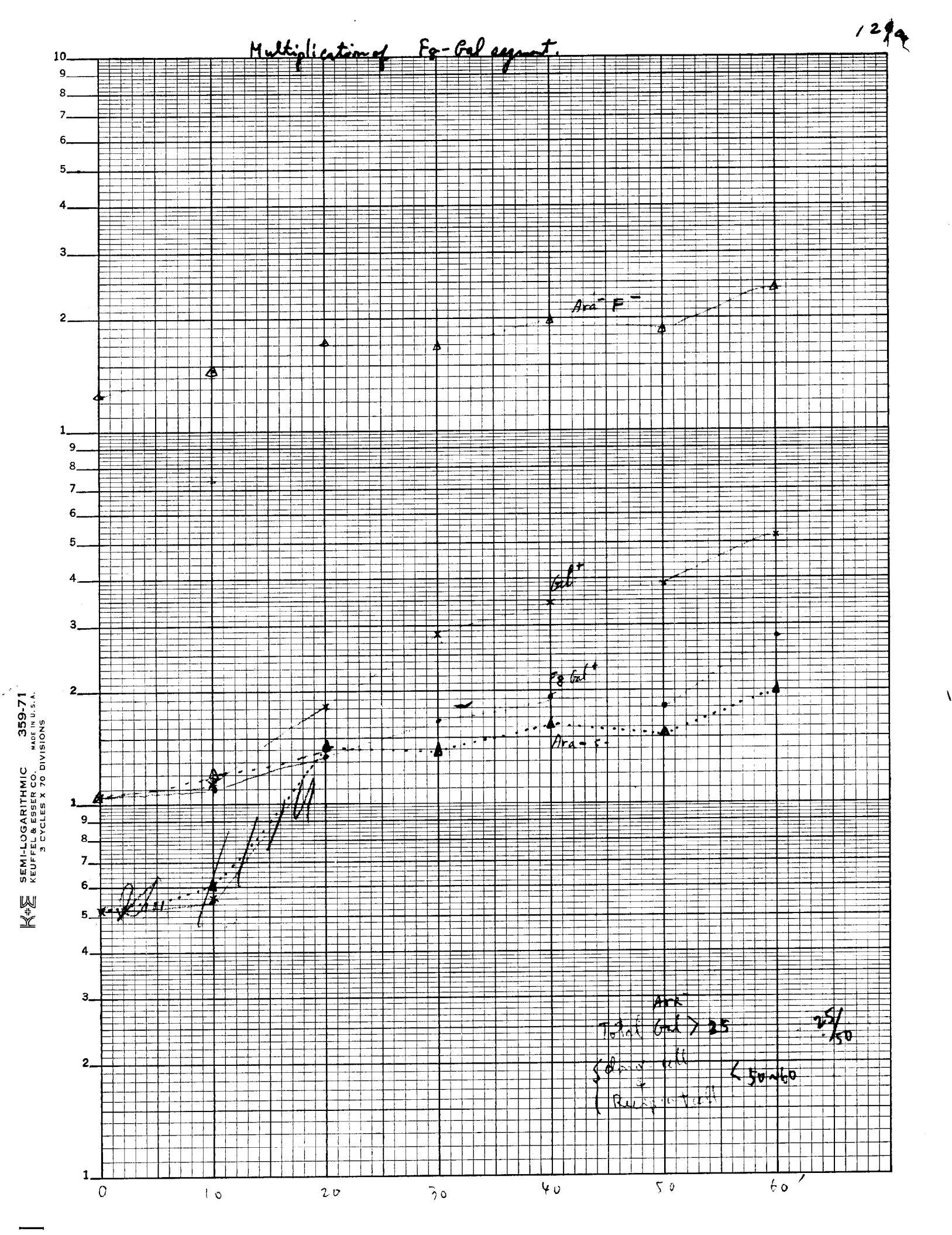
14/VIII 1959 REF:

Principle: W6 F8 → × 4573. Gal_2 Xyl_2 Mtl Ara_2 Mal, S^R.

Method: 0.2 ml overnight culture / 10 ml Pen. → 3 hr at 37°C on rotator 12:00 ~ 2:55. → 5 ml ♀ / 10 ml. + 0.5 ml ♂

time (min)		0		10		20		30		40		50		60	
		2:55		3:05		3:15		3:25		3:35		3:45		3:55	
Dilution		2×10^{-6} ml		1×10^{-6}		2×10^{-6}		..		"		2×10^{-6} ml		1×10^{-6}	
spread		$10^{-2} \times (10^{-2} \times 10^{-1} : 2)$ ml													
		+	−	+	−	+	−	+	−	+	−	+	−	+	−
B Gal	1	39	441	22	201	62	570	82	526	96	615	135	657	74	367
	2	31	408	15	285	65	578	114	598	132	704	147	572	88	389
	3	35	396	19	255	54	577	91	573	115	675	106	637	99	445
	Σ	105	1245	56	741	181	1725	287	1695	343	1994	388	1866	261	1201
														522	2402 (×2)
		+		+		+		+		+		+		+	
B Ara.	1	39		22		48		54		52		56		41	
	2	31		14		47		64		71		69		51	
	3	35		19		40		51		66		55		48	
	Σ	105		55		135		169		190		180		140	
				110										280	(×2)
Net infection.	Δ	0		1		46		118		153		208		242	

Multiplication of Eg-Col segment.



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3 CYCLES X 70 DIVISIONS

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Stock No, 1~27

130

Segregation of Lac⁻ from Heterogenote for Lac segment.

30/VII ; 1959

REF:

(Lac 61)

W4121 × W3112 F_{13}^{+} (Lac 61 / Lac 2) (Took from spot test (see P. 126))

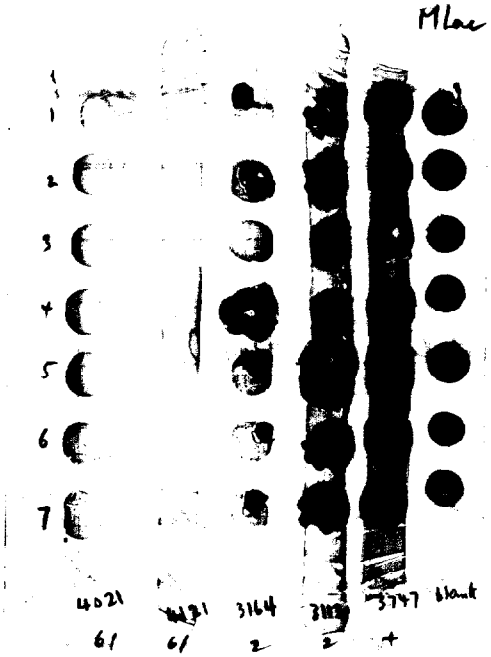
Method: Streak

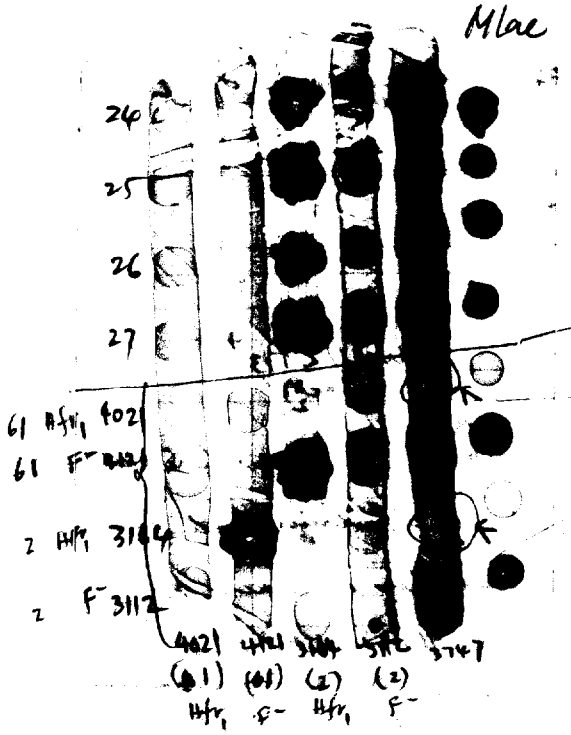
1. purify it on BLac. (5 plates)
2. Pick Lac⁺ colonies and suspend them into Lac⁻ Hfr. tubes.
3. Restreak them on BLac, and pick Lac⁻ segregant (pick one Lac⁻ from each Lac⁺ streaks).
4. streak the isolated Lac⁻ on BLac. (ca. 48 colonies are isolated. Some of them maybe Lac⁺. 29 were Lac⁻.)
5. Test Lac⁻ by cross streaking on MLac. (and sex-compatibility)

	V_6	Lac⁻ mutation end. ex	F	× 3112 (Lac 2) F⁻	× 3164 (Lac 2) = Hfr	× W4121 (Lac 61) F⁻	× W4021 (Lac 61) Hfr	× 3747 (Lac⁺)
1	S	61	♂	+11	+1	●	–	+
2	S	61	♂	+21	+3	–	–	+
3	S	61	♂	+14	–	–	–	+
4	S	61	♀	–	+32	–	–	+
5	S	61 61	♂	+	+2	–	–	+
6	S	61	♂	+21	+1	–	–	+
7	S	61	♂	+14	+1	–	–	+
8	S	61	♂	+16	–	–	–	+
9	S	61	♂	+18	+1	–	–	+
10	S	61	♂	+	+15	–	–	+
11	r/s	61	♂	+	+4	–	–	+
12	S	61	♂	+14	+3	–	–	+
13	S	61 61	♀	–	+26	–	–	+
14	S	61	♂	+18	–	–	–	+
15	S	61 61	♀	–	+24	–	–	+
16	S	61 61	♀	–	+18	–	–	+
17	S	61	♂	+2	–	–	–	+
18	S	61	♂	+13	+1	–	–	+
19	S	61	♂	+13	+3	–	–	+
20	S	61 61	♂	+	+2	–	–	+
21	S	61	♂	+14	–	–	–	+
22	S	61	♂	+10	+1	–	–	+
23	S	61	♀	–	+29	–	–	+
24	S	61 61	♂	–	+23	–	–	+
25	S	61 61	♂	–	+43	–	–	+
26	S	61 61	♂	–	+23	–	–	+
27	S	61 61	♂	–	+34	–	–	+
Hfr 1	4021	Lac 61		(–)	–	–	+	
	4121	Lac 61	61	–	+	–	+	
Hfr 1	3164	Lac 2	2	–	–	+	–	
	3112	Lac 2	2	–	+	–	–	

Conclusion: 27/27 : Lac 61 (100%)

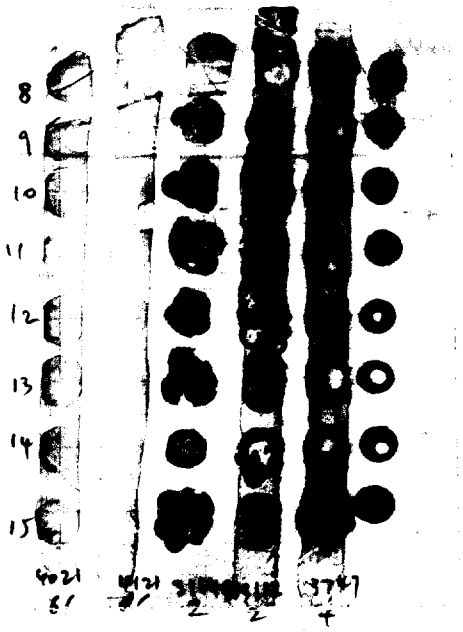
Non-transductants of Hfr 1 and Hfr 3





control.

?2 Hfr, x F' stock!

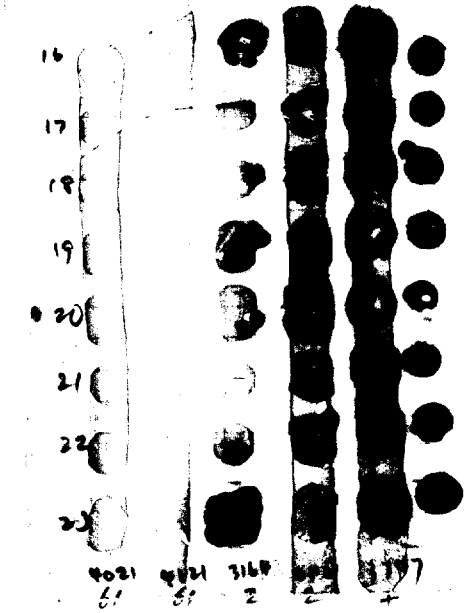


4021 looks wrong! It may be a mixture (lac⁺).

" W4021 x lac_2 F-

is ok.

Tests for Vi-loci.



Retest # 11.

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Stock No 28 ~ 40

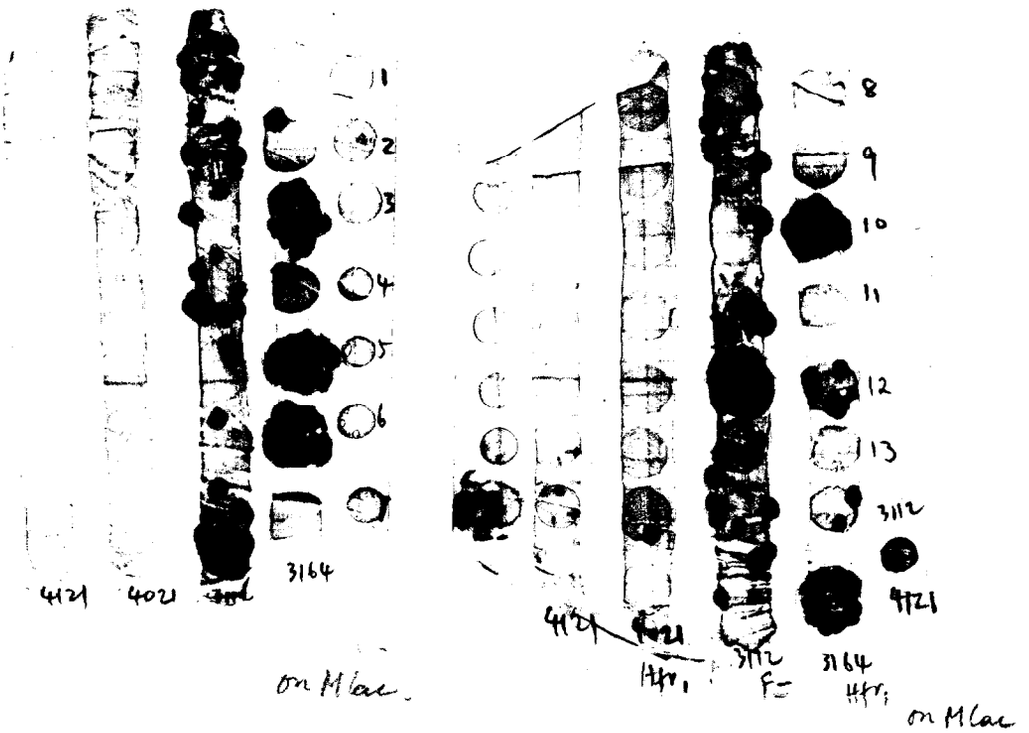
131

Segregants Analysis of Lac_2^{ex} / Lac_{61} F_{13}

7/xxx ; 1959

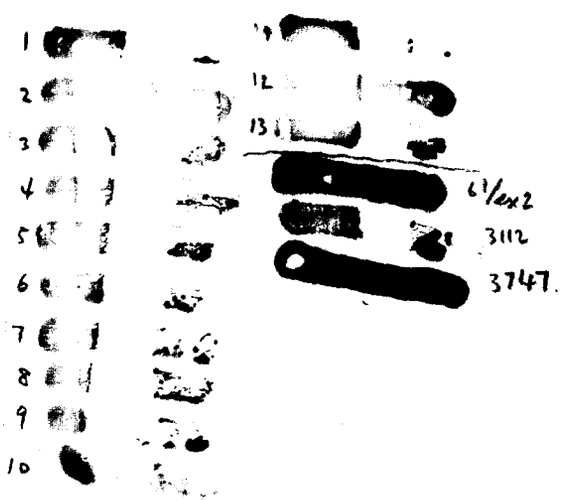
REF:

	1	2	3	4	5	6	7	8	9	10
			W4121 × W3112 F_{13}							
			Lac_{61}	Lac_2						
		Lac	F	× 3112	× 3164	× 4121	× 4021			
	V_L	and$_{61}$ ex$_2$		(Lac_2)	(Lac_2)	Lac_{61}	Lac_{61}			
				F⁻	Hfr$_1$	F⁻	Hfr$_1$			
1										
2										
3										
4										
5										
6										
7										
8										
9	1	S 61	♂	+20	−	−	−			
0	2	r 61	♂	+60	+1	−	−			
	3	S 61	♀	+2	+33	−	−			
1	4	S 61	♂	+9	+1	−	−			
2	5	S 61	♂	+2	+20	−	−			
3	6	S 61	♂	+1	+32	−	−			
4	7	S 61	♂	+23	−	−	−			
5	8	S 61	♂	+11	−	−	−			
6	9	S 61	♂ ♀	+5	−	−	−			
7	10	S 61	♀	+2	+28	−	−			
8	11	S 61	♂	+8	−	−	−			
9 →	12	r/s 61	♂	+61	+9	−	−			
0	13	S 61	♂	+4	−	−	−			
1	[redacted]			[redacted]	[redacted]			See back page.		
2	Lac_{61}/Lac_{ex_2}	r/s								
3	3112	S								
4	3747	R								
5										
6										
7		conclusion:		13/13 : Lac_{61} (100%)						
8										
9										
0										
1										
2										
3										
4										25
5										13
6										27
7										65
8										
9										
0										
1										
2										
3										
4										
5										
6										
7										
8										
9										
0										



Test of V_6 loci

Retest # 12.
This strain looks F_{13}^{+} $Lac_{61}\frac{V_6^S}{Lac_{61}V_6^R}$.



on B lac

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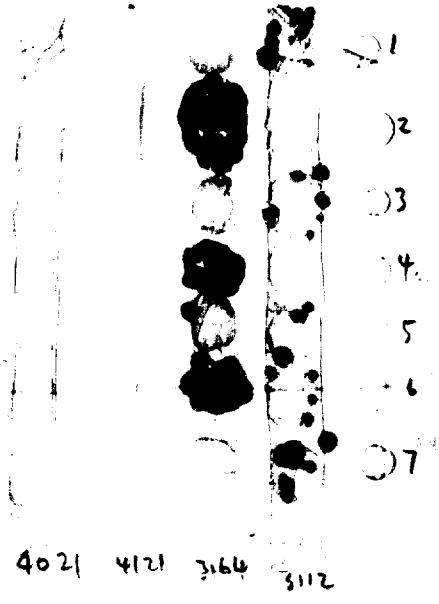
Segregant analysis of Lac61/ex 2. F13.

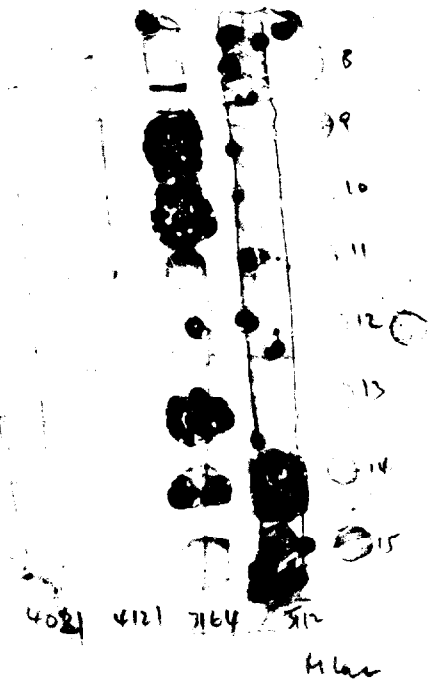
132

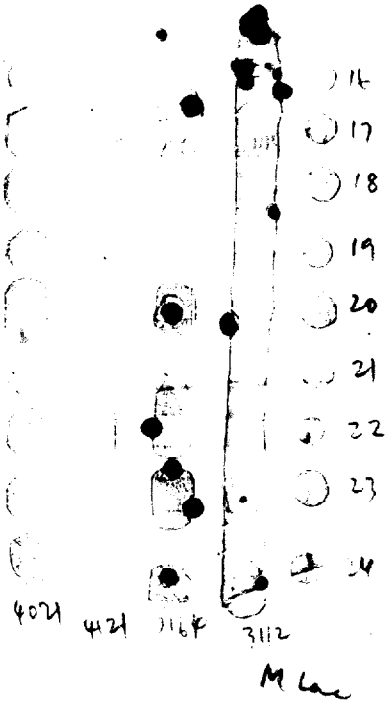
13/VIII :19 59

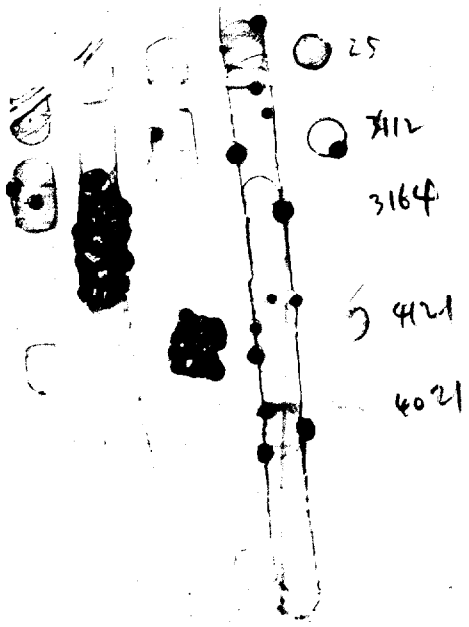
REF:

	1	2	3	4	5	6	7	8	9	10
						× 3112	× 3164	× 4121	× 4021	
		V6	Lac end61 ex2		F	Lac+ F−	Lac+ Hfr1	Lac61 F−	Lac61 Hfr1	
1	1	S			♂	+15	−	−	−	
2	2	S			♀	−	+60	−	−	
3	3	S			♂	+7	−	−	−	
4	4	S			♀	−	+30	−	−	
5	5	S			♀	+2	+2	−	−	
6	6	S			♀?	±2	+36	−	−	
7	7	S			♂	+15	−	−	−	
8	8	S			♂	+6	−	−	−	
9	9	S			♀	±1	+44	−	−	
10	10	S			♀	±	+68	−	−	
11	11	S			♂?	+4	−	−	−	
12	12	S			♂	+7	+1	−	−	
13	13	S			♀	±1	+25	−	−	
14	14	S			♂	±	+10	−	−	
15	15	S			♂	+33	−	−	−	
16	16	S			♂	+15	±1	−	−	
17	17	S			?	−	±1	−	−	
	18	S			?	−	−	−	−	
19	19	S			?	−	−	−	−	
20	20	S			?	±1	±1	−	−	
21	21	S			?	±1	−	−	−	
22	22	S			?	±	±1	−	−	
23	23	S			?	±1	+2	−	−	
24	24	S			?	±1	±1	−	−	
25	25	S			♂?	+2	−	−	−	
					♀					
	61/ex2	r/s								
	3112	S								
	3747	r.								
										??
	3112				F−	±2	±1	−	−	
	3164				F−	−	−	+2	+	
	4121				F−	−	+	−	−	
	4021					+3	−	−	−	

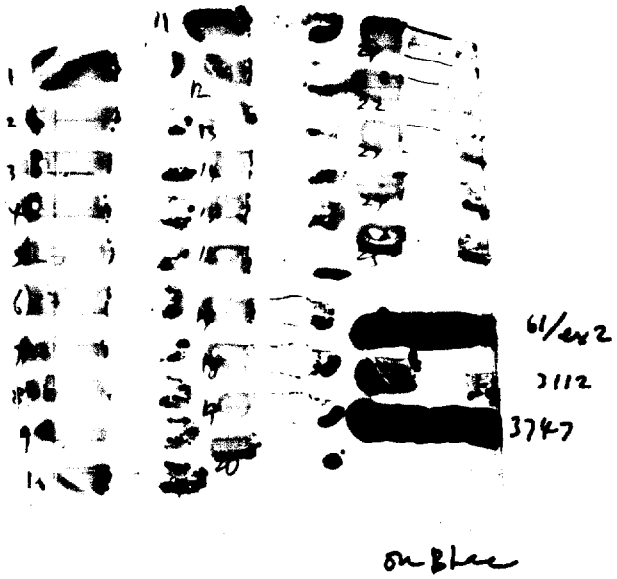








Test of V_6 loci



Retest # 14

This maybe V_6^3 Lac 61/ex 61 V_6^5.

Unsuccessful : use M Lac Sm in next time, (Lac^{61}/Lac_2^- does grow on MLac)

Comparison of the [redacted] frequencies of the transduction of exogenote and endogenote from heterozygous diploid (Lac^{-}_{61}/Lac^{-}_{2}).

4/4/4 ; 1959 REF: cf. 167, 166

Principle

F_{13}. Lac^-_{61} / Lac^-_{2} ex (3112 F_{13} + 4121)

→ × F^- Lac^-_{61} W4121

→ × F^- Lac^-_2 W3112

cultural age : 4 hrs. on rotator.

strain : all these strains used as donor are purified on Blac and Pick single colony and suspended in [illegible] (ca 10^7) 0.1ml is inoculated into [illegible] and kept there on rotator for 4hrs.

Method : 1) Mix Lac^{61-}/Lac^-_2 ex and Lac^-_{61} or Lac^-_2 use excess F^- , Ratio. F_{13} 1 (0.1ml) : F^- 10 (1.0ml)

2) Incubate ~~them~~ for 1/2 hrs. ~~[redacted]~~ at 37°C. 3:25 pm

3.) Seed them on MLac agar after optimal dilutions.

use : 10^{-2}ml, 10^{-4}ml, 10^{-5}ml per plate for each crosses.

4) Count all numbers of each experiment.

Results:

Serival count

	① F_{13} 61^- ½ ex → Lac_2^- W3112		① F_{13} 61^- ½ ex → 4121 Lac^-_{61}		② F_{13} 61^- ½ ex → [redacted] Lac^-_2 W3112		F_{13} 61^- ½ ex 2 → × 4121 61^-		3. F_{13}^+ W3112 → × 2^-	F_{13}^+ W4121 → × 61^-
Serival count	Lac^-	Lac^+	Lac^-	Lac^+	Lac^-	Lac^+	Lac^-	Lac^+	Lac^- Lac^+	Lac^- Lac^+
10^{-6}ml/plate	1057	89	1084	81	944	51	868	85		

Fertility on ml/plate

10^{-1} [redacted]

10^{-3} [redacted]

[redacted] too much, if cannot count them. (Use 1/100 of these experiments)

[redacted] 1084 81 [redacted] 944 51 [redacted] 868 85

[redacted] /ml

ml/plate

* Qualitative experiment.

3112 F_{13} × 4121

3747 1 2

×3112 (2) [redacted]

on MLac

3112 F_{13} × W421.

3747 1 2

4121 (61)

on MLac

Successful.

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Transduction of Lac⁻ segment from heterozygote, $Lac^{61}_{+}/_{ex}Lac^{-}_{2}$.

134.

11/VIII 1959 REF: cf. 145, 146

Principle:

Lac 61 /ex Lac 2 S_S → Lac 61 F⁻ S^R W 4121 S^R (W4629)

W4121 (Lac 61 /ex Lac 2) → Lac 2 F⁻ S^R W 3112

compare the fertility in both crosses.

Check, Sm-sensitivity of these strains. on B Lac Sm.

Result. on B Lac Sm

Qualitative test.

Preliminally test by spotting method — 4121 3112 3112 F13 4121 (Lac 61 /ex Lac 2)

Spot 4121 (61/ex 2) on W 4121 S^R and W 3112 streaked on B Lac Sm.

Result: see back page.

	3112 (Lac_2^-)	4121 S^R (Lac_{61}^-)	blank
4121 61/ex 2	+	++	−
3112 2/ex 2	+ reversion	++	+ reversion.
3747 +/ex +	++	++	−

at 37°C

12/VIII

Quantitative test.

· cultural age: 0.2 ml : overnight culture / 5 ml/ phase — Shake on rotator 10:30 a.m. 3:30.

· Ratio of mix: 1 ml | 3112 + 4121 Lac 61/ex 2 0.1 ml

1 ml | 4121 S^R (W4629) + 4121 Lac 61/ex 2 0.1 ml.

· Time of incubation: ~~1 hr 2 hr 3 hr 4 hr 5 hr~~ 90.

· survival count: inoculum size 10^{-6} ml. of original culture. / plate

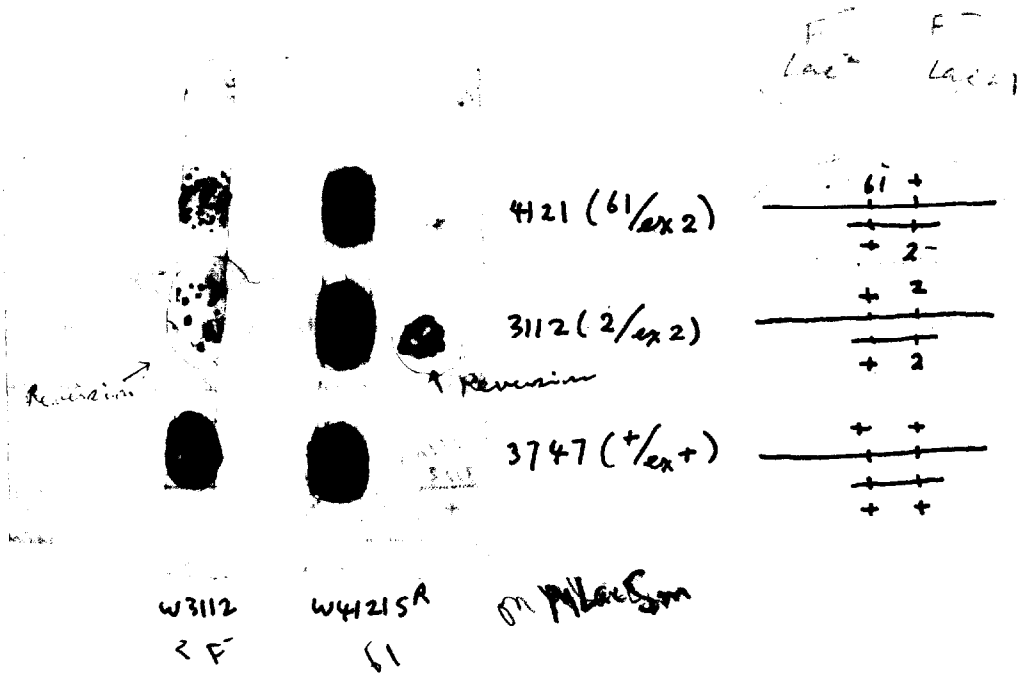
4121 61/2 × 3112 Lac⁻ 751 : Lac⁺ 10 — 4121 61/2 × 4121 S^R Lac⁻ 0 : Lac⁺ 13

· Inoculum size for counting recombinations. 10^{-3} ml. / plate on M Lac Sm.

Result:

		# of Recombinants. Plate 1	Plate 2	# of Colonies 10^{-3} ml / plate.	% of Rec.
3747 +/+	× 3112	too much ca 10^3	10^3	+/0 0 − 24 ; + 2 − 38	
3747	× 4629	ca 10^3	10^3	+/0 22 193 ; + 38 − 212	
4121 61/2	× 3112	0	0	+/0 0 − 65 ; + 1 − 68	C ≥ 10^{-3}
				+/0 0 − 112 ; + 4 − 269	
4121 61/2	× 4629	2	5	+/0 0 − 66 ; + 1 − 53	C ca 10^{-3}
				+/0 2 − 136 ; + 4 − 100	

Conclusion: Exogenote is transferred with very high frequency, but not very high for endogenote. This implies all F13 Lac HI clone are diploid in that (Lac) segment. And also possible to speculate that linkage between F13 and endogenote, and exchange of end-ess fragment are relatively low.

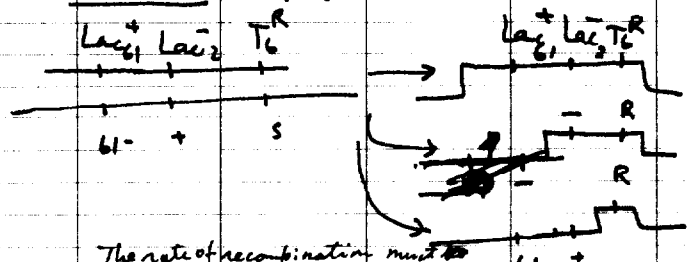

F⁻ lac⁻
F⁻ Lac⁺
4121 (61/ex 2)
3112 (2/ex 2)
Reversion
3747 (+/ex +)
W3112 ? F⁻
W4121 SR 61

Linkage of $T6^R$ and Lac^-_2 in exogenote of ($Lac_2\ T6^R \times Lac_{61}\ T6^S$) $3112F_{13} \times 4421$

6/VII 1957

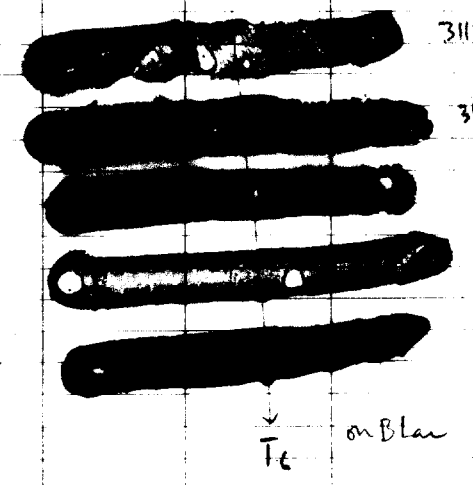
REF:

Principle. Segregant (non-recombinant) is Lac^-



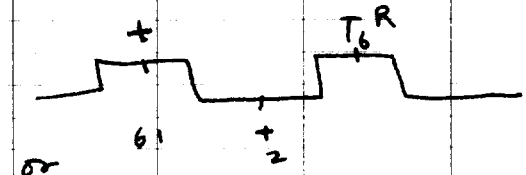
The rate of recombination must be express ratio of length of $V_6 - Lac_2 - Lac_{61}$.

If Lac is closely linked with V_6, Lac^- can be found at the lysed part of $Lac_{61}\ V6^S$ / $Lac_2\ T6^R$ by T_6.



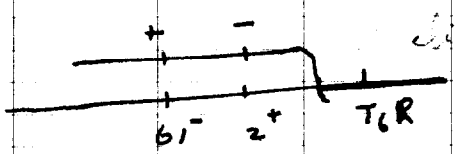
Result:
actually the lysed part is also Lac^+.

explanation: There are enough number of segregants which arised by recombination between two different Lac^- mutations. Their fermentation of lactose may cover the presence of Lac^- colonies.



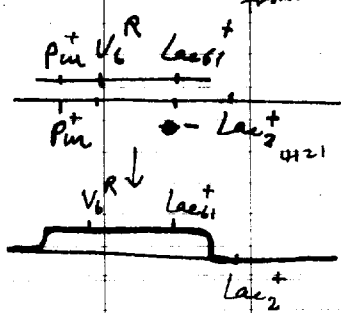
T_6^R Lac^+.

or



elimination of $T6^S$ segment

or $Lac_2\ F_{13}$ is hemizygote for Lac_2.



Comparisons on the stability of maleness of F+ and F' to AO treatment.

8/IV ; 1959 REF:

Strain W3086 F_8 , W3744/1 , W6.
F_8 F' F+.

Mixture apposit combination
W6 F_8
3086 F+.

- Cultural age : Overnight culture in Plenary broth.
- Inoculumsize : Ca. 10^8 cells/ml. $10^{-2} \times 10^{-2} \times 10^{-1}$ (0.1ml)/5ml
- AO-treatment : AO-medium : 30γ/ml ; NSB pH. 7.6.

Result :

A). Survival count on B Mal. 10^{-6} ml/plate. (before treatment.)

Exp II compatibility

A). Mal+ W6 F_8	Mal- 3086 F+ F_8 F-
234	89
327	152
227	107

Exp II

Total W6 F_8 tested	F-	Total 3086 F+ tested	F-
44		43	
%			

B). After treatment. (After treatment)

Exp ① 3086 F_8 + W6

Treated with AO. Replicated on MGal (x 4573.)

W3086 F_8 Σ	F_8	F-	W6 F+ Σ	F+	F-
35	4	31	35	5	30
21	2	19	24	1	23
Σ 56	6	50	59	6	53
% 89.3 F-			89.9 % F-		

Conclusion : Looks no difference in accessibility of F. to acridine method.

Exp ② W6 F_8 + 3086 F+.

Untreated control

W6 F_8 Σ	F_8	F-	W3086 F+ F+	F-
32	32	0	32	0

X4573 on MGal.

Treated experiment (with AO: 30γ/ml)

W6 F_8 Σ	F_8	F-	W3086 F+ F+	F-
	0	59	0	42
%	0	100	0	100

X 4573 on MGal.

Untreated control

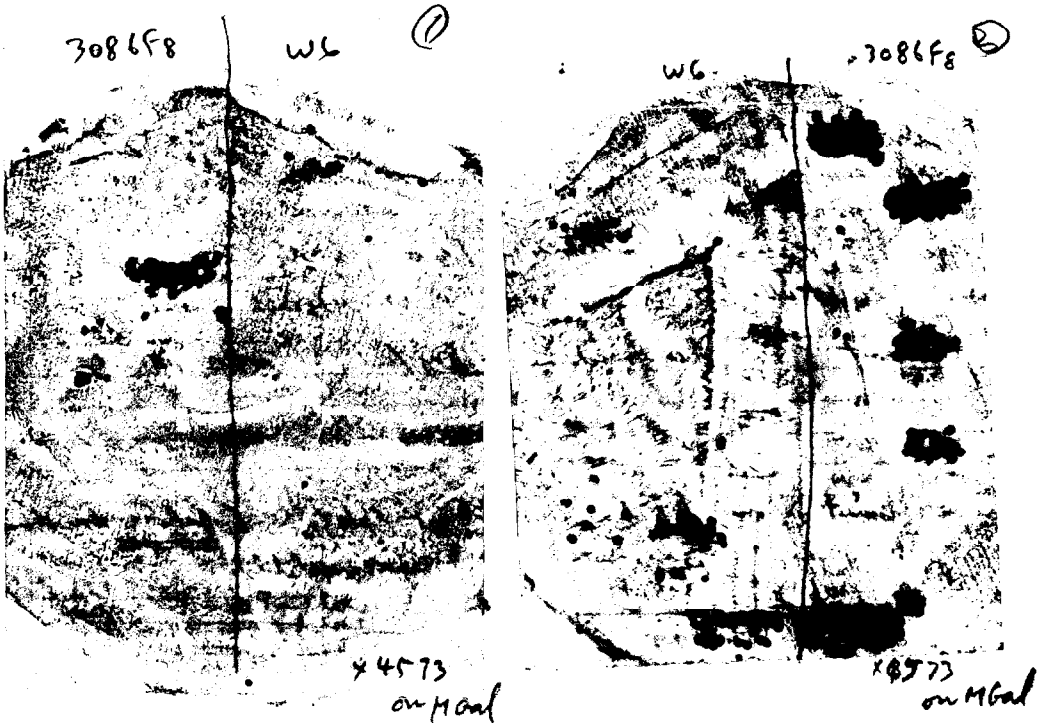
3086 F_8 Σ	F_8	F-	W6 F+ F+	F-
55	55	0	55	0
%	100	0	55 100	0

Exp (I).

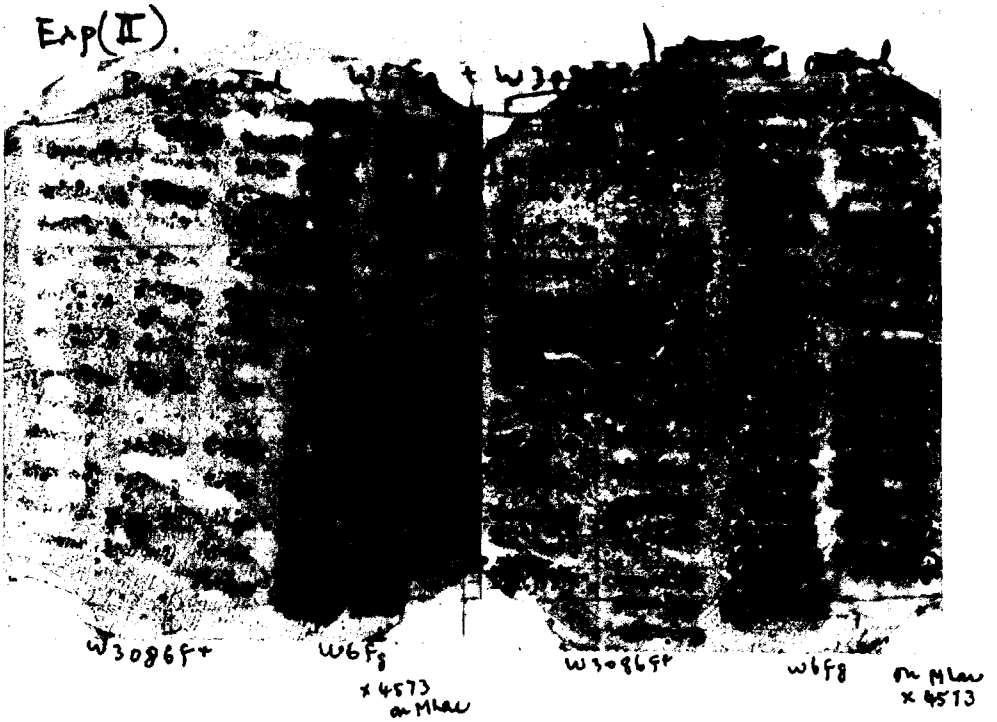
Inoculum ca. 10^2 cells/ml

AO 30 g/ml

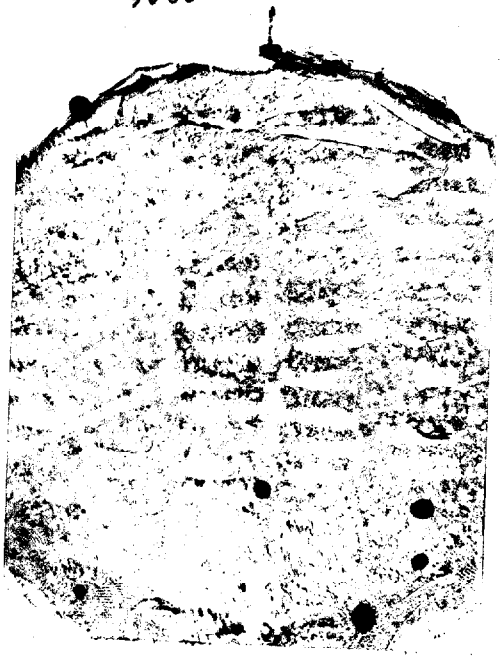
AO 3086Fg + W6 6 min light



Exp (II).

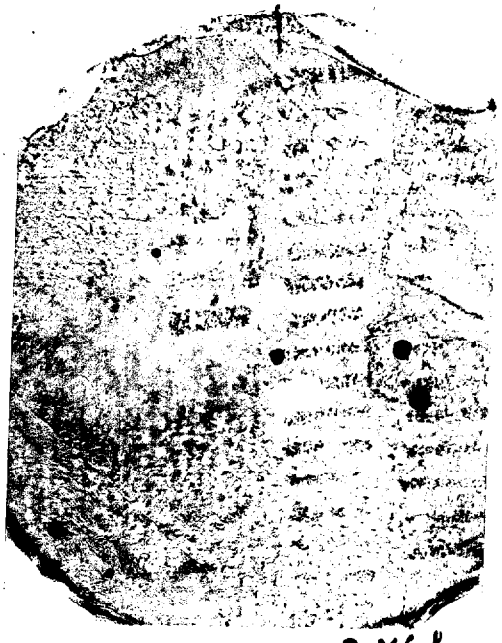


Ao treated
3086 F+ W6F8.



x 4573
on MGel

Ao treated.
W3086F+ W6F8



on D MGel
x 4573

Size of Gal - F_{8D} segment.

12/VIII ; 1959

REF:

Strain used : ♀ Pur⁻ Try⁻ Gal_2^- Xyl 2 Mtl Mal, Ara_2 S^R F^-.

♂ F_8 M^- (W6 F_8)

Cultural age : 0.2 ml : overnight culture / 5 ml Penassay broth.

↓ 2 hr on rotator at 37°C.

10:45 AM ~ 12:45 AM. ca 10^8 cells/ml

Ratio of mix : 1 ♂ F_8 : 0.1 ml F^- → Incubate it overnight at 37°C

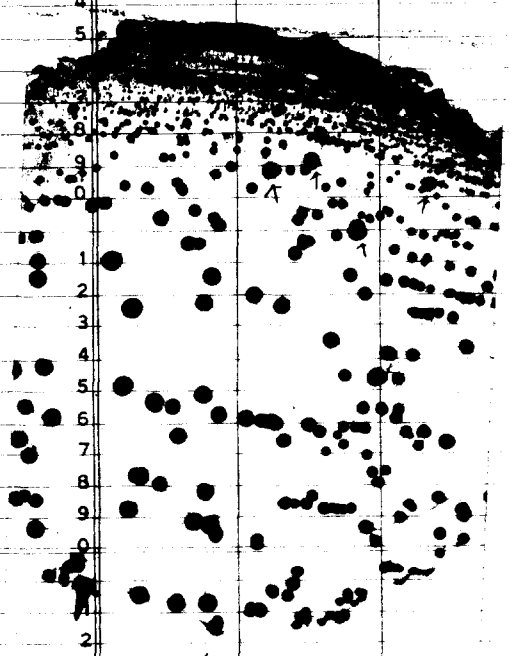
Principle :

W6 F_8 → × F^- Pur⁻ Try⁻ Gal_2^- S^R

Select on B Gal Sm.

Pick Gal^+ and test the other markers.

Result. Very few colonies become Gal^+. It may come from pur⁻ try⁻ of the recipient strain.



On B Gal Sm.

5 days

Gal^- becomes black

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Unsuccessful

Infection of F_3 and Remove of F_1 from it.

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Principle : $F_3 - F_1 = Rfd_3$

20/VIII 1959

REF: [W4296 shows strong agutrophy [with M] see back page 10

Principle : W4616 (Lac⁻ F_3) → W4295 $F^- L^- S^R$

[→ W4296 $F^- T^- S^R$]

↑ Too much agutrophy.

W4616 → W4295.

F_3	Total Colonies tested
20	350
15	379

- Cultural age : 2 hrs. 0.2ml / 5ml.
- Excess F_3^+ : 10 : 1. F^-
- Ratio : (2ml 0.2ml)
- Purified on B Lac Sm.
- Replicated on M Lac Seeded W3086.

Result: The first step.

3133 F_3 → 4295

on M Lac

~~4577~~

W3086

Retest of newly isolated F_3^+ by spot test. X 3086 on M Lac.

on M Lac

X 3086

Try it again

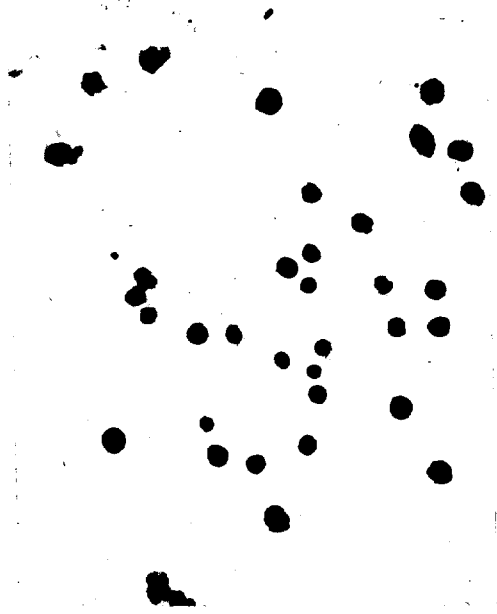
Syntrophy of M and T.

3137 F_3 → W4296

on Mlac

W808

Master plate



on Blac

Unsuccessful.

MICROBIAL GENETICS

Isolation of Lac61⁻ F_{13} by ~~Simple~~ infection of F_{13}

3747 ——— × 4121 S^R

21/VIII 1959 M V_6 F_{13} (Lac61 F^- S^R) REF: cf P120 & P120a P.55

Purpose: Still it is very hard to isolate F_{13} Lac^-_x strain from segregants. This experiment is carried out to find out more easy method.

Principle: 1. F_{13} may split off from Lac loci and infect to Lac^-. (This possibility is very low)

2. Lac^+ F_{13} consist heterozygous diploid state in first step, then recombination occurs, and endogenote Lac^- becomes exogenote. This Lac^- F_{13} becomes infective, and multiplies rapidly.

In any way, if it does occure, it may be possible to isolate Lac^- F_{13} using this method.

Result:

1. Rate of infection of Lac^- F_{13} into 4121 S^R.

Lac^-	91	17.2 %
Lac^+	438	82.8 %

2. Rate of F_{13} · Lac61.

Lac^- isolated	F^-	F_{13}
108	108	0

~~method — The~~ see bellow.

Conclusion: F_{13} itself does not infect into F^- in this experiment. This ratio may be much less than 1 %.

Method:

1. Make 2hrs culture (shaked on rotator in plenary at 37°c.)
2. Mix 10 : 1 : 10
   F_{13} : F^- : fresh broth.
   2 ml, 0.2 ml, 2 ml of fresh broth.
3. Incubate it for overnight.
4. Purify it on Olac 3m.
5. Pick Lac^- colony on Olac 3m.
6. Replicate on MLac, on which Lac^- F^- are seeded.

5/12.

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Isolation of Lac61/ex. Lac61. F_{13}.

24/VIII, 1959.

REF: Lac_x must be Sm^S

Principle:

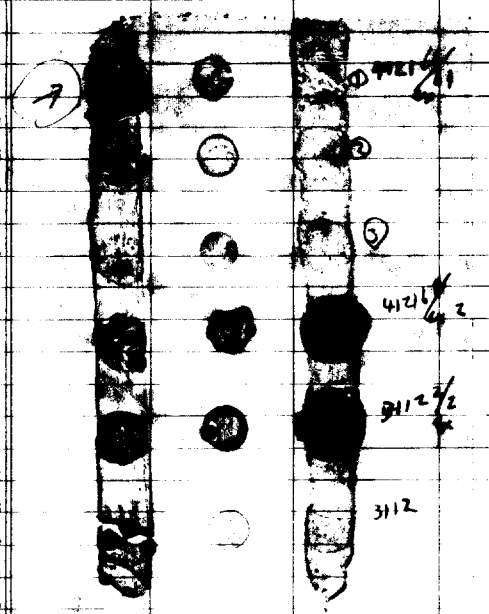
1st step: $Lac_2/_{ex}Lac_2\ F_{13} \rightarrow Lac_{61}\ F^- \rightarrow Lac_{61}/_{ex}Lac_2\ F_{13}$

2nd step: $Lac_{61}/ex\ Lac_2\ F_{13} \longrightarrow Lac_2/ex\ Lac_{61}\ F_{13}$ on MLac^S 3112.
(exchanging Promes of exogenote and endogenote.) (Detect it by replica plating method) see back page.

3rd step: $Lac_2/ex\ Lac_{61}\ F_{13} \rightarrow Lac_{61} \rightarrow Lac_{61}/_{ex}Lac_{61}.F_{13}$

Principle is the indirect selection of a clone which contains many cells of $Lac_2/ex\ Lac_{61}$ arised by exchange of Lac locus from $Lac_{61}/_{ex}Lac_2$.

Retest



Replicated from Blac.

1st selection.

Pick.

on MLac
× 3112.

4121 61/4x2

3112 2/2

3787 1/1

4121 3/4x61

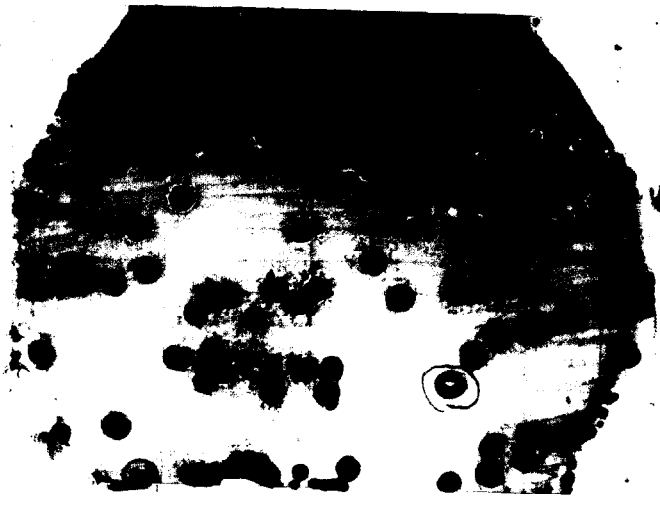
4829
Lac 61.
SR

3112
Lac 2
SA

← Reisolated on MLac

Replicify

2nd selection. on Blac

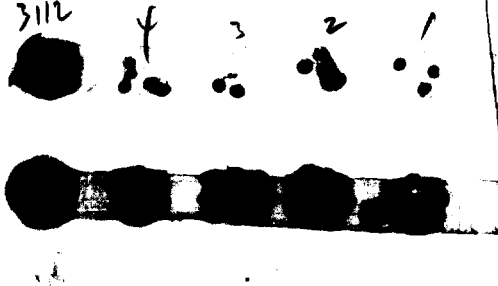


on MLac

×3112.

4121

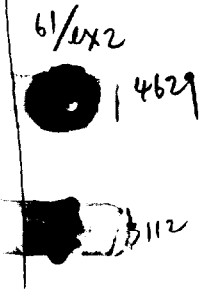
3112 4 3 2 1



4121
61/4x2

4829

3112



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6/40 ; 1959

REF:

	1	2	3	4	5	6	7	8	9	10
1		Isolation of		HI Gal	from W3747.					
2										
3										
4										
5										
6										
7										
8										
9										
0										
1										
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MGal
×2979.

on Blac

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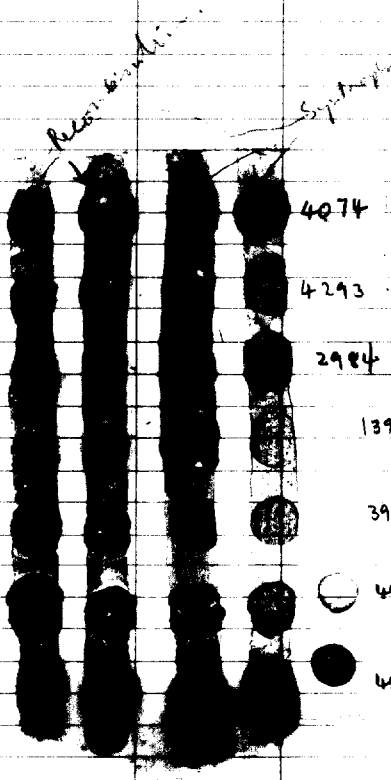
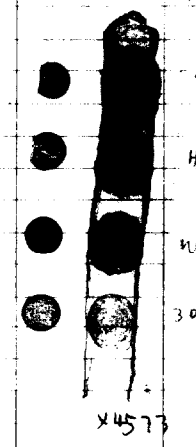
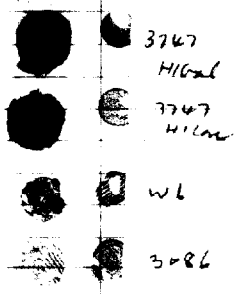
Comparison of the fertility of W3747, HI Gal and HI Lac.
I II.

1959. REF:

Purpose : W3747 shows two kinds of colonies, one is high fertile to







X4573 on MXyl

HI Lac 3747 HIGal 3747 W6 W3086-

on MLac

Conclusion

HI gal TLB1 A1 Try

HI Lac Pur

Lac Pur

gal TLB1 A1 Try

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Order of integration of chromosome via ~~cross~~ $F_2 \times F^-$

26/VIII ; 1959 REF:

W4540

Strains : W6 F_2 , W4573,
H^- F_2 $Ara_2 Lac_{85} Gal_2 S^R Mal_1 Xyl_2 Mtl_1$

Blending : Gage : 60 , 1 min.

overnight culture 0.1 ml/5 ml pen →

Cultural age : 1:30 ~ 4:00 2½ hrs. at 37°C on rotator

W6 F_2 : 2 ml
W4573 : 0.2 ml

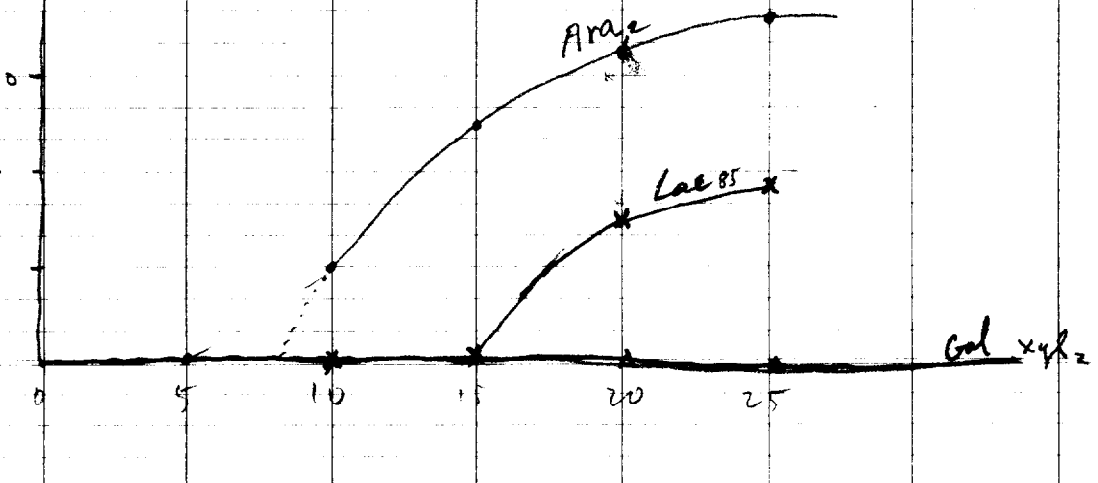
# of Recombinants/plate

Time	0	5	10	15	20	25
	Ara_2 Lac Gal_2 Xyl_2	A, L, G, X	A L G X	A L G X	A L G X	A L G X
plate# 1	0 0 0 0	0 0 0 0	1 0 0 0	9 0 0 0	45 1 0 0	51 3 0 0
10^{-3} ml 2	0 0 0 0	0 0 0 0	0 0 0 0	12 0 0 0	33 1 0 0	57 5 0 0
Σ	0 0 0 0	0 0 0 0	1	21 0	78 2	108 8

Survival count	Lac											
10^{-6} ml	−	+	−	+	−	+	−	+	−	+	−	+
	4	54	6	64	8	51	3	73	6	71	7	102
	●3	29	5	67	6	76	7	63	9	93	9	137

fertility % ca. 0.1 % ca. 1 % ca. 0.5 % ca. 1 %

%



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Comparisons of the fertility of exogenic and endogenic segment.
(Quantitative test.)

16/IV ; 1959 REF:

	1	2	3	4	5	6	7	8	9	10
		Cultural age:		Overnight culture in Penassay broth.						
1										
2		Cross	:	10:30 AM. ~ 12:30 AM			at 37°C.			
3				· ratio of mix			of 0.1 ml : 8 ml.			
4				· mixed well.						
5										
6										
7					fertility		survival count		10^{-6} ml.	
8					10^{-2} ml	10^{-3} ml	Lac+	Lac-		
9		W 4121	61/ex 2	x 4629		11	4	457		
0							4	382		
			Pre incubation.							
1										
2			Post incubation.							
3										
4										
5		W 4121	61/ex 2	x 3112.		[redacted]				
6						1148	3	364		
7			Pre				4	547		
8										
9			Post.							
0										
1					10^{-3}	5×10^{-4}	93	713		
2		W 3747	x	4629		985	106	754		
3										
4			Pre							
5										
6			Post.							
7										
8										
9		W 3747	x	7112		1065				
0										
			Pre							
1										
2										
3			Post.							
4										
5										
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7										
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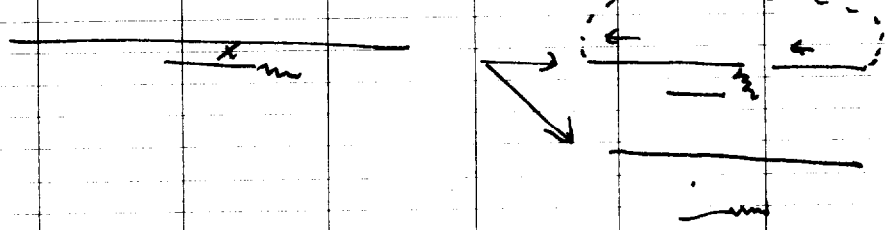
Comparisons of the fertility of exogenetic and endogenetic segment.

1959 REF:

	1	2	3	4	5	6	7	8	9	10
		Cultural age : Overnight cultures.								
1										
2										
3					(F⁻ Lac⁻ 615ˢ)	fertility 10^{-3} ml	fertility 10^{-2} ml	Survival (10^6 ml) Lac⁺	Survival (10^6 ml) Lac⁻	Rec. / Lac⁺
4									+ −	
5		W4121 61/ex 2	→	× W4629		79	1214	8	38 | 8 71	
6						180	1097	5	45 | 8 75	
7										Ca. 10^{-2}
8						Ca. 100	Ca. 1000	Ca. 5×10^8	4×10^7	
9										
0										
1										
2		W4121 61/ex 2	→	× W 3112	(F⁻ Lac₂ 3ˢ)	0	12	1	27 | 18 15	
3						0	20	3	32 | 23 155	
4										
5						0	Ca. ~~10~~ 20	Ca. 2×10^8	Ca 3×10^7	< Ca. 10^{-5}
6										
7										
8										
9						10^{-3} ml	10^{-2} ml	+	− | + −	
0								248	35 | 174 14	
1		W3747 +/+	→	W4629		1231	68	~~262~~ → 79	138 20	
2							101	~~120~~ 46		
3								262	41	
4						Ca. 1000	Ca. 100			Ca. 10^{-2}
5								Ca 2×10^8	4×10^7	
6										
7										
8										
9								+	− | + −	
0		W3747 +/ex+	→	× W 3112		126	52	170	46 | 240 70	
1							113	262	79 | 291 68	Ca. 10^{-2}
2						Ca 1000	Ca 100	Ca. 2×10^8 ; 5×10^7		
3		Conclusion:								
4		Fertility of								
5		exogenetic ≅ endogenetic × 100.								
6										
7										
8										
9		This may be explained by crossing over between F' and host-								
0		chromosome at about 1%.								



Transformation of Gal in 4-12.

20/X. 1959 REF:

Experimental condition:

1 Conc. of Duponol (Na-lauryl sulphate) : 1%.

2 Time of Treatment : 3 hrs at 37°C. in H_2O.
1:45 ~ 4:45.

3 Bacterial culture : overnight; ca 10^8 ~ 10^9 cells/ml. washed once with H_2O.
~~the~~ primary grown. suspended into H_2O.
32 ml's of W6FG culture was used. (4 tubes of 8 ml per.)
↓
concentrated into 2 ml's.

1 ml ↙ ↘ 1 ml

treated with duponol. final 1%. untreated

20% duponol Soln 1 ml + Bact. Suspension 1 ml

H_2O 1 ml + Bact. susp. 1 ml

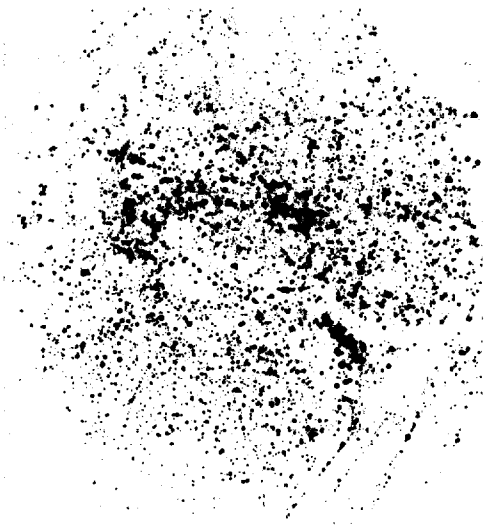
4. Method of transformation.

DNA (dilution of Original culture)	10^{-1}	10^{-2}	10^{-3}	
# of $_3$ Recombinants.	~~too much~~	521	43	
surviving cells in 0.1 ml	—	—	2,5 ↑	

Bacteria	10^{-1}	10^{-2}	10^{-3}	10^{-6}
# of Recombinants				41
surviving cells in 0.1 ml	—	—	—	435 585 ↑

Next step. Filter this DNA sample.

DNA 10^{-1} dil



DNA 10^{-2} dil

DNA 10^{-3} dil.

Intact.

10^{-2} dil.

10^{-5} dil.

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Transformation of F8 Gal. Negative result.
(filtered material).

23/IX ; 1959 REF:

	1	2	3	4	5	6	7	8	9	10

Isolation of Sample.

1. W6 F8. M⁻ F8 ; W4573 $Lac_{95}Gal_2$ .....
4 tubes, 8 ml. → suspend in 4 ml of H_2O → add 4 ml of dupanol 20 %.
2. ~~Treatment~~ treatment ~~with~~ dupanol : Na-lauryl-sulfate.
(final conc. in dist. W. 10 %.)
37°C 4 hr.
3. Filtered through millipore filter.
4. Mix : dupanolated sample.
0.2 ml of W4573 + 0.2 ml of broth + 1 ml of DNA.
: old culture (2 days)
and incubate it for 30'.
5. Seed it on M Gal.

Result :

1) survival count : 0.2 ml of the extract was seeded onto EMBlac.
No colonies are observed.

2) Recombinating activity.

Extract :	1 ml	(1/10 diluted) 0.1 ml	(1/100 diluted) 0.01 ml
# of Recombinations.	0	0	0

Conclusion :
Mating activity of W6 F8 is in insoluble part.

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Action of propamidine isethionate (P.I.) to F and F'(E)

4:4'-diamidino-diphenoxypropane-di(β-hydroxyethane sulphonate) 5 gr.

May & Baker Ltd. Dagenham England.

16/IX ; 1959

Glucose gave me on 11/Sept. '59.

Stock solution : 100 mg / 1 ml soluble in water.
(Weigh 100 mg of P.I and dissolved into sterile H_2O in vial.)

1. Test survival & bacteriostatic concentrations of P.I.

Conc. of P.I. (cc. of Stock (1 ml) + 0.95 ml)	.05 mg/ml 0.05	1 mg 0.05	100 γ (with 0.05)	x 1/100 10 γ 0.05	x 1/1000 1 γ. 0.05	Control 0 γ
W6 F_8	−	−	−	+	+	+
W6.	−	−	−	+	+	+

make ~~stock~~ sterility-test of P.I. soln. 100 γ 4. 10 γ.
− −

" Media . 5 ml Penassay broth, pH. 7.0.

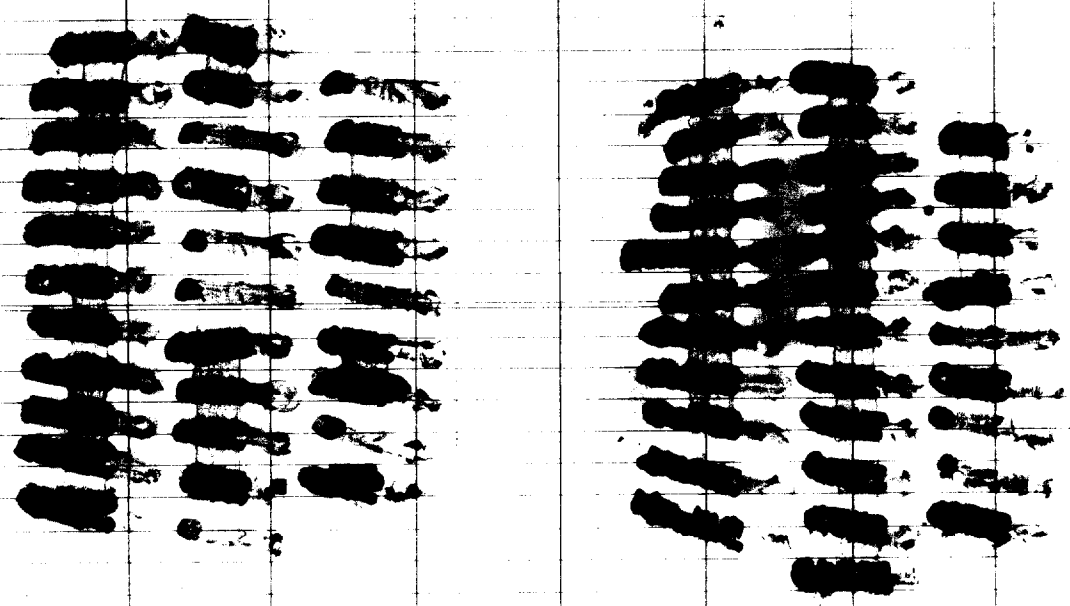
2. Streak these treated cells on Blue agar,
100 γ. P.I.

3. Test their compatibility by cross-brushing method.

Result: P.I.-treated (10⁸/ml Rec.) Untreated control.

F⁻	F⁺	% F⁻	F⁻	F⁺	% F⁻
7	24		3	29	

W6 P.I. W6 Cont.



MLac MA573

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Action of Propamidine isethionate. on F. II.

150

24/4 ; 1959

REF:

1	2	3	4	5	6	7	8	9	10
	Med. Penassay.	inoculum size: ca. 10^5 cells/ml.			Strain: W6.			(×1/100) Conc. of P.I. Soln used: 1 mg/ml.	
Conc. of P.I.		0 γ / 0 ml	10 γ / 0.05 ml		20 γ / 0.1 ml		50 γ / 0.25 ml	(5 ml plm.)	
Ratio of conversions F⁻/total tested		0/30	0/28		0/28		4/29		
(%).		0	0		0		14.0		
effect		−	−		−		+		

purpose: Is PI active for removal of F? (P.I. can remove chloroplast of Euglena.) In former experiment, 10 γ/ml of PI looks active. Increase the concentration of P.I and see the increase of that activity.

conclusion. P.I. seems effective at 50 γ. in penassay, inoculum size, 10^5 cell/ml.

cont.

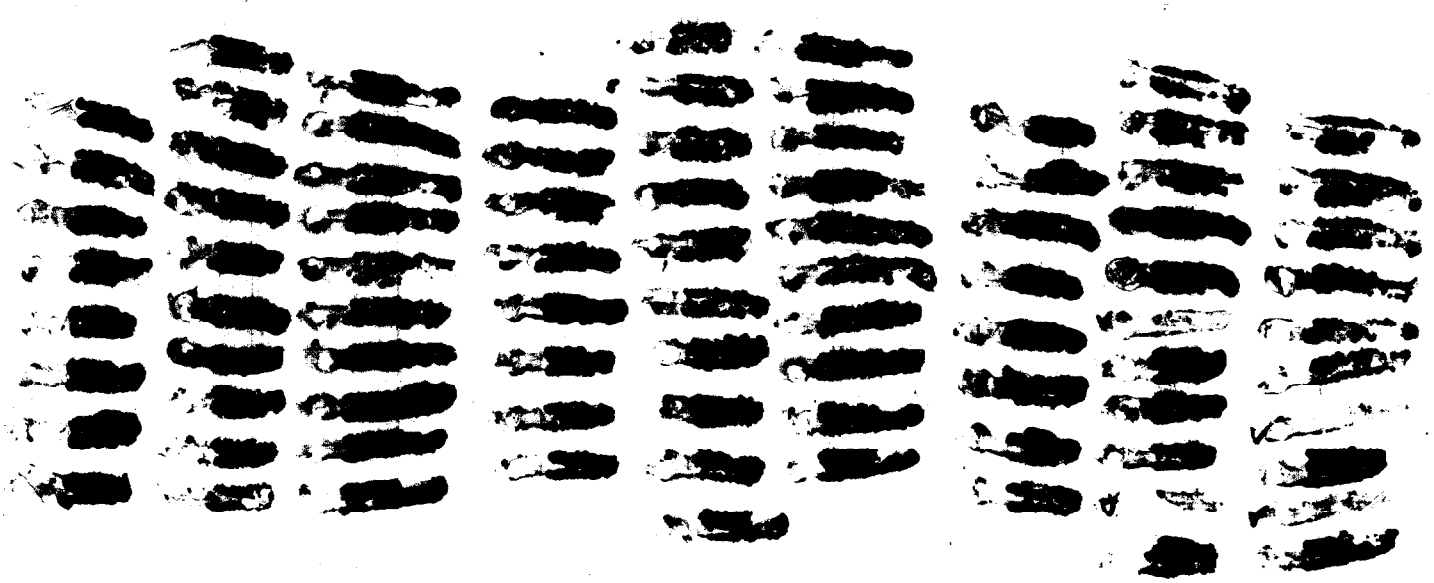
W6 P.I. 0σ.

MLac
X4573

P.I 10σ

P.I. 20σ

P.I 50σ



MLac X4573