Tuesday 12/23/52

- 902 hp^S - transduction of old culture - Repeat using younger cell.

	EMB gal	additions	no. per 3 days	
	1	no add	14	
2.4×10^{10}	2	0.1 750 λ - 1	52	38
	3	0.1 902 λ - 1	11	—
3.8×10^9	4	0.1 892 λ - 1	51	37
$c. 10^{11}$	5	0.1 811 λ - 4	43	29

$\frac{3.8 \times 10^2}{2.4 \times 10^{10}} = \frac{1}{6.3 \times 10^7} = \frac{1}{6.3 \times 10^6}$

values low because of cell age? Repeat.

- 578

EMB gal	additions	no per 2 days	
1	no add	72	high?
2.	0.1 1485 λ	131	
3	0.1 K-12 λ 12/20	$\frac{1}{10} = 120 = 1920$	

- ✓ λ titer of this appears about as expected since titer is about 1/10 that of K-12 λ 12/30

(2/2) 1485 λ = 1.1×10^9 59/1.11

K-12 (12/29) λ = 2.3×10^{10} 1848/23

$59 \times \frac{23}{1.1} = 1233$

"nat." λ = 1233

thigg λ = 1848

booked

Thursday 12/26/52

811 prototrophic gals - - To test the hyp that gal+ h of 811 not transducing 811, 578 are reverting to an allele with not sufficient selective advantage in EMB gal to allow detection of transduction.

EMB gal ↓	additions	no per 2 days	
1.	none	23	
2.	(811 λ) - 4	18	
3.	(811 gal+ -1) - 1	15	This gal+ doesn't work in EMB
4.	(811 gal+ -2) - 1	22	" " " " " "
5.	(811 gal+ 5) - 1 untreated (CONTROL)	214	all work in EMB
6	(811 gal+ 5) - 2	465	" " "
7.	(811 gal+ -8) - 1	309	" " "
8	(K-12 λ) 30-1	164	" " "

No screen - presumably gal+ -1, gal+ -2 are since maintains closely linked to gal+

2050 gals - gal deletion - Repeat - has high background - Make purified

EMB gal	2050 - additions	no per. 2 days	
1	none	296	
2	750 λ - 1	1/4 = 255 = 1020	these counts all low - large nos of small pfg -
3	902 λ - 1	1/4 = 476 = 1904	
4	892 - 1	288	
5	811 - 4	1/4 = 208 = 832	

$>2.9 \times 10^{10}$

4.9×10^{10}

3.8×10^9

$\frac{7.74 \times 10^3}{2.4 \times 10^{10}} = \frac{1}{3.3 \times 10^6}$

$\frac{1.608 \times 10^4}{4.9 \times 10^{10}} = \frac{1}{3.1 \times 10^6}$

Sunday 12/28

874 gal deduction = another Lysogenic cut of 518

EMB gal	Addition	hr pop 18 days	
1	none	17	
2	0.1 750 λ -1	77	believe as 811 with greater
3	0.1 902 λ -1	518	pow ducability by K-12 ?
4	0.1 892 λ -1	167	
5	0.1 811 λ -4	23	
6	0.1 K-12 λ 30-1	1067	

D- 518 & 892 ↙ (-) from single transductions

gal - segregants - → purified thru 3 single col. isolations

test - ~~transduced~~ deficiency to by 811 and 892 examination for cellule by transducability

12/29

- 1402

duction to see if 1821 deficiency is also 1402 dif. - apparently is -

EMB gal	Addition	hr pop 18 days
1.	none	17
2.	0.1 750 λ -1	28
3.	0.1 902 λ- 1	61
4.	0.1 892 λ- 1	31
5.	0.1 811 λ- 4	22
6.	0.1 K-12 30-1	85

purification begun - streak survival single colony isolation / tz

892 - 2 λ not sterile discarded

902 - 2 λ sterile -

892-1 λ - recovered - lost 0.1 ml

$10^8 - 15$

8

23 = 2.3×10^9

Tuesday 12/30/52

Exp D = Repeat examination of segs. from 51 & 892 - Seg purified thru 3 pickings -

Page 2 days

	Gal-bal	Model	811λ-9	892λ-2	Probable gal locus	1985	From Strain
strk all	1	40	142	253	(?)	Segs	Recheckers - unstable
892	2	53	35	69	4	"	Rechecks
811	3	22	40	87	4	"	not t/811λ
all	4	8 (few small)	58 (all small)	173 all small	(?)	"	Rechecks - unstable
	5	37 >5 (obj segs)	32	71	4	"	
811	6	32 >5	40 >5	74 >5	4	"	not t/811λ
	7	28 >small	40 >small	99 >5	4	"	
	8	40	36	90	4	"	
	9	29 >small	37 >small	88 >5	4	"	
892	10	93 all small	46 all small	79 all small	4	"	Rechecks
	11	35	34	71	4	"	
892	12	52 >5	59 >5	81	4	"	Rechecks
892	13	47 >small	31 small	82	4	"	Rechecks
	14	last - but both are low in pop.		73		"	
	15	21 small	121	275	(?)	"	Recks - unstable
	16	30 (custom)	33	68	4	"	
892	17	39 >small	35 >small	60 >5	4	"	Rechecks
	18	35 >small	44	75 >5	4	"	
	19	27 (all small)	95 (all small)	256 (all small)	(?)	"	Rechecks - unstable
	20	27 all small	101 all small	324 all small	(?)	"	" "
	21	5	40	146	(?)	"	" "
892	22	45	35	61	4	"	Rechecks 892
892	23	58	36	87	4	"	" "

5/23 transduced by both 811λ 892λ

5/23 probably new type!

Checks

Pop #	D1 t 811			D1 t 892			D2 t 892			D3 t 811			D4 t 811			D4 t 892			D6 t 811			D10 t 892			D12 t 892		
	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
1.	mix	mix	mix	mix	mix	mix	mix	+	mix	+	+	+	mix	mix	mix	mix	mix	mix	mix	+	+	mix	+	+	mix	+	+
2.	"	"	+	"	"	+	"	+	+	+	+	+	"	"	"	"	"	"	"	+	+	"	+	+	"	mix	mix
3.	"	"	mix	"	"	+	"	mix	mix	+	+	+	"	"	"	"	"	"	"	+	+	+	+	+	"	+	+
4.	"	"	mix	"	"	+	"	+	+	mix	+	+	"	+	+	"	"	"	"	+	+	mix	mix	mix	"	mix	mix
5.	"	+	+	"	"	+	+	+	+	+	+	+	"	mix	mix	"	"	"	"	+	+	"	mix	"	"	mix	mix
6.	"	mix	mix	"	"	+	mix	+	+	"	+	+	"	+	+	"	+	+	+	+	+	"	mix	"	"	+	+
7.	"	"	mix	"	"	+	"	+	+	"	+	+	"	mix	mix	"	mix	mix	+	+	+	+	+	+	+	+	+
8.	"	"	+	"	"	+	"	+	+	"	+	+	"	+	+	"	"	"	mix	+	+	mix	mix	mix	+	mix	mix

single colony isolation

	D13 t 892			D15 t 811			D15 t 892			D17 t 892			D19 t 811			D19 t 892			D20 t 811			D20 t 892			D21 t 811		
1.	mix	+	+	mix	+	+	mix	mix	mix	mix	+	+	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix	mix
2.	"	mix	mix	"	+	+	"	"	"	"	+	+	"	"	"	"	+	+	"	"	"	"	"	"	"	"	+
3.	+	mix	+	"	mix	mix	"	"	"	"	+	+	"	"	"	"	mix	mix	"	"	"	"	"	"	"	mix	mix
4.	mix	+	+	"	"	mix	"	+	+	"	mix	mix	"	"	"	"	"	"	"	"	"	"	"	"	"	+	+
5.	"	mix	mix	"	+	+	"	mix	mix	"	+	+	"	"	"	"	"	"	"	"	"	"	"	"	"	+	mix
6.	+	+	+	"	mix	mix	"	"	"	"	mix	mix	"	"	"	"	"	"	"	"	"	"	"	"	"	mix	+
7.	mix	+	+	"	"	mix	"	+	+	"	mix	mix	"	"	"	"	"	"	"	"	"	"	"	"	"	-	+
8.	"	+	+	"	"	mix	"	+	+	"	+	+	"	"	"	"	+	+	"	"	"	"	"	"	"	"	mix

Check on Exp D = 578892 Second run

Pap

	D21E892 1	2	3	D22E892 1	2	3	D23E892 1	2	3
1.	mix	mix	mix	+	+	+	mix	+	+
2.	"	+	+	mix	+	+	+	+	+
3.	"	mix	mix	+	+	+	mix	mix	mix
4.	"	+	-	mix	+	+	"	+	+
5.	"	mix	"	"	mix	mix	"	mix	mix
6.	"	+	"	"	+	+	"	+	mix
7.	"	+	"	"	+	+	"	mix	mix
8.	"	mix	"	+	+	+	+	+	+

1/6/53 Tuesday.

Recheck on D4, D15

1/11/53

D15

EHB gal	Additions	Inc. last 2 days
1.	none	14
2.	750λ-1	$\frac{1}{2}$=151, 302
3.	902λ-1	22
4.	2050λ-1	40
5.	811λ-4	73

1/12/52 >70

D4

EHB gal		
1.	none	12
2.	750λ-1	$\frac{1}{2}$=180 360
3.	902λ-1	12
4	2050λ-1	50
5.	892λ-1	365
6.	811λ-4	90

D4X902	(+)	(-)			
1.	0	88			
2.	2	96			
3.	2	130			
4.	3	109			
	7	923	7/930	1.6%+	

maybe be suspect - 902 culture probably contained (+) cells

second cross D1X902 P8 166 → 4.2%

0.042 / 18.0 / 12.8 / 12

241 / 177 / 422

Tuesday 12/30

2175 = 902 lp_2^S - Repeat duction ē aerated cultures

		EMB gal	Addaketi	Pap. 2 days			
		1	mm	S	(gum plate)		
2.4×10^{10}	2		750 A-1	71	57	66	$\frac{1.6\times10^3}{2.4\times10^{10}} = \frac{1}{2.6\times10^6}$
	3		902 A-1	14	"		
3.8×10^9	4		892 A-1	102	51	97	$\frac{9.70\times10^3}{3.8\times10^9} = \frac{1}{2.6\times10^5}$
2×10^{10}	5		811 A	51	43	46	$\frac{9.6\times10^3}{2\times10^{10}} = e.\frac{1}{2.2\times10^6}$
	B				pieces?		

1402 Examination of parent strain for possible presence of lp_2^R
6 isolated colonies tested against 1-2 - all sensitive

= 2175

902 lp_2^S ABOVE - Stability

	902 + 750			902/902 A			902 + 892			902 + 811		
	1	2	3	1	2	3	1	2	3	1	2	3
1.	mix	+	+	mix	+	+	mix	+	+	mix	mix	mix]
2.	"	mix	mix]	"	+	+	"	mix	+	"	mix	"
3.	"	mix	mix]	"	+	+	"	+	+	"	mix	+
4.	"	+	+	"	+	+	"	+	+	"	+	+
5.	"	mix	mix	"	+	+	"	+	+	"	mix	mix]
6.	"	+	+	"	+	+	"	+	+	"	+	+
7.	"	+	+	"	+	+	"	mix	+	"	+	+
8.	"	+	+	"	+	+	"	mix	mix]	"	+	+

[stock: make]

STOCK OF 1655 X 902 → gal_2 - lp_2^S
made - Entered in Stock book = 2175

Sat. 1/3/53

-2050 gal 3- Reexamination of galactose using a single colony isolate from streak to lower background-

	Gal 3 gal	Addition	No. per 8 days	
	1.	none	32	
>2.9×10^10	2.	750 d -1	104+	many small
4.9×10^10	3.	902 λ -1	1/8 = 132 = 1056	" "
	4.	892 λ -2	130	" "
	5.	811 λ -4	1/4 = 55 = 210	" "

$\frac{7.3\times10^{2}}{2.9\times10^{10}} = \frac{1}{3.3\times10^{7}}$

$\frac{1.0\times10^{3}}{2.9\times10^{10}} = \frac{1}{4.9\times10^{6}}$ ← large no? See examination below

- Sunday 1/4/53

2050 above - stability examination

Pipiliate #	2050 + 750			2050 + 902			2050 / 892 λ			2050 + 811		
	1	2	3	1	2	3	1	2	3	1	2	3
1	mixed	+	+	mixed	+	+	mixed	+	+	+	+	+
2	"	+	+	"	+	+	+	mixed	mixed	mixed	+	+
3	"	+	+	"	+	+	mixed	"	mixed	"	+	+
4	"	+	+	"	+	+	+	+	+	"	+	+
5	"	+	+	"	+	+	+	+	+	+	+	+
6	"	+	+	"	+	+	+	+	+	mixed	+	+
7	"	+	+	"	+	+	+	mixed	mixed	"	+	mixed?
8	pure	mixture		"	+	+	+	+	+	"	+	" (?)

1/7/53
Wednesday (Held over from 1/6/52)

Eq E = 2050 + 81 - gal - segments

Segment #	Wood.	81 λ-4	2050 λ-1	1985	Probable age in years	
1	* 61	⅛ = 87 456	⅛ = 102 204	Eyes	(?)	
2	* 75	⅛ = 76 608	* * 111	"	3 (?)	
3	15	⅛ = 33 264	14	"	3	
4	c. 30	c. 200	c. 10	"	3	
5	c. 20	c. 200	c. 20	"	3	
6	17	c. 300	18	"	3	
7	12	⅛ = 39 136	22	"	3	
8	69	⅛ = 83 116	48	"	3	
9	* 116	⅛ = 197 588	* 182 *	"	3 (?)	on 1/12/52 this plate 2050 λ = c. 360
10	c. 10	c 200	c. 20	"	3	
11	* 20	½ = 78 156	20	"	3	
12	* 28	¼ = 96 384	89	"	(?)	
13	18	½ = 41 82	11	"	3	
14	½ = 147 294	¼ = 154 (many small) 616	¼ = 148 (many small) 592	"	(?)	
15	50 > 31 >	c. 400	c. 200	"	(?)	
16	c. 10	c. 80	c. 10	"	3	
17	c. 100	c. 150	c. 10	"	3?	
18	c. 200	c. 100	c. 10	"	?	
19	17	½ = 80 640	⅛ = 62 496	"	?	11 (3)
20.	c. 10	c. 150	c. 20	"	3	9 (?)

Disregard → 2050

* = [illegible]

Nearly all plates have large numbers of small papillae making counting and interpretation difficult. - In general results suggest the same conformity suggested by the 5184 results.

892

Sunday 1/11/53

- B-7 X 902 - Repeat cross B7 = 578t902 → (-) rxn duplicated by and 1 but 902 1

		(+)	(-)		
EMS	1.	3	222		
gal	2.	7	276		
	3.	9	315		
	4.	7	239		
		26	1052	26/1078	= 2.5% ←

These results suspect since 902 cultures probably contaminated (+)

- B-8X902 B8 same as B7

EMS	1.	3	105		
gal	2.	4	94		
	3.	3	98		
	4.	4	88		
		14	385	14/399	3.5% ←

811 X892

		(+)	(-)		
EMS	1.	0	132		
gal	2.	0	156		
	3.	0	123		
	4.	0	184		
		0	595	0/595	

gal4 X gal3

811 X 902

		(+)	(-)		
EMS	1.	1	143		
gal	2.	0	230		
	3.	0	129		
		1	602	1/603 =	0.16%

gal4 X gal2

811 cultures probably contaminated upon rereading results.

2050 · Gal uptake

EMB-gal		Day 2 days	
1.	no add	22	
2.	K-12 (304)	32	what happened?
3.	1486tK-12-1	227	
4.	D1 (see pg 153	solid smear of pop 5000?	D1 = 578t892 → (-) by characteristics by gal as 902, unknown by crossing data?
5.	578t892-1	solid smear of pop 5000?	D1 ≠ 578t892 578t892 results as previous.

8/20/53 Tuesday after my -

	no ppt 2 days 518	874
518 ab° 874		
{2 ml cell susp. ⟶ no add assay 0.1 ml	12	19
{1 ml K-12 (39-1) ⟶ adsorb 5 min		
{centrifuged susp'd 2.0 ml ⟶ assay 0.1 ml = K12 ductai	c.87	593
↓ {leaves 1.8 ml add 0.9 ml 81-4 λ - adsorb 5 min		
↓ {centrifuge + susp'd 1.7 ml ⟶ assay 0.1 = 1st 81 effect	c.59	233
↓ {add 0.8 ml to 1.7 ml remain - adsorb 5 min {centrifuge - susp'd in 1.7 ml ⟶ assay 0.1 - 2nd 81 effect	c.44	169

these plates rod-growth does not look like 518. sp. ppt small after 2 days

Exposure to virus λ seems to reduce pop. - Better technique

518 vs 518t892

10x cell dil	
1.0 ml	0
1.0 :	1-10 1 ml
1.0 :	1-100 1 ml
1.0 :	1-1000 1 ml

all diluted 1/2 → adsorb 5 min → centrifuge, resusp in original vol → assay 0.1 ml

0.1 ml

this plate shows almost confluent lysis

Titer these 518t892 811t892 preps used as ppt oddity observed is due to low titer - rechecked on 2050 - Is this phage lytic? - different from λ, λ-2 ?

SW927 Rugh

1st lysates show 1 plaque / 950 SW in del 7 106 - titer c. 109

Wg 16
Wg 19 } Auxotrophs catalogued
date as pg 120-130.

1/23/6

Lysate titers

811-4
10^8 - 113, 67 = $90 \times 10^8 = 9 \times 10^9$

902-1
10^8 - 93, 90 = $91 \times 10^8 = 9.1 \times 10^9$

902-2
10^8 - 24, 19 $22 \times 10^8 = 2.2 \times 10^9$

902-3
10^8 - 100, 48 7.4×10^9

2050
10^8 - 314, 263 2.9×10^{10}

518E892-1
10^7 - 195, 166 1.8×10^9

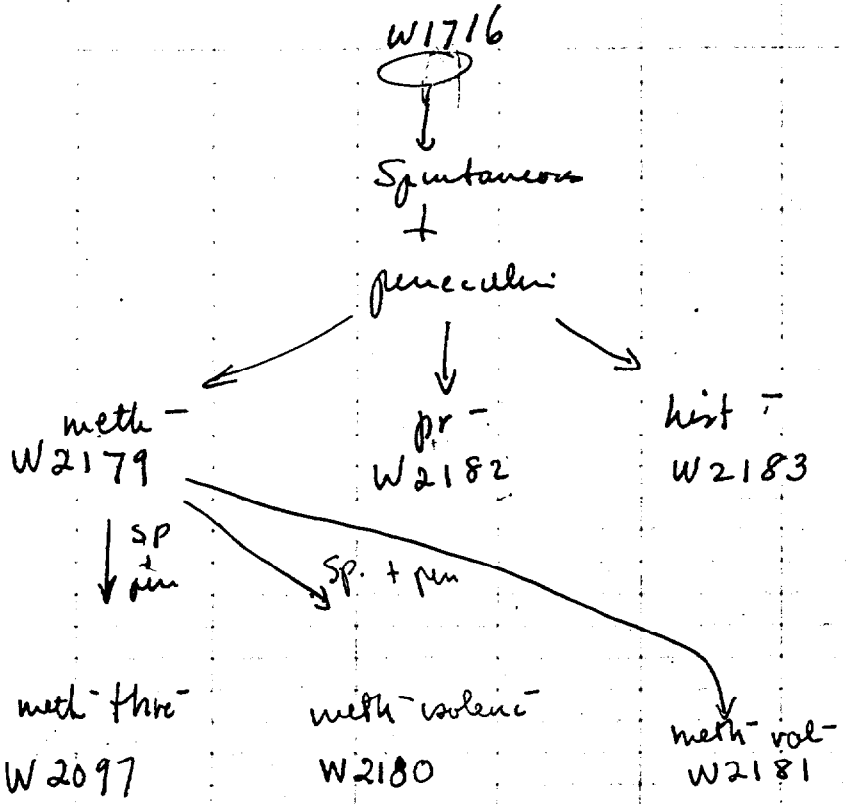
$\frac{2 \times 10^5}{2 \times 10^9} = \frac{1}{10^4}$

01
10^8 105, >87 9.6×10^9

K-12 - (12/20)
10^6 = 244, 213 2.3×10^{10}

1485
10^7 = 129, 96 1.1×10^9

Wg 16


W1716
Spontaneous
+
penicillin
meth⁻
W2179
pr⁻
W2182
his⁻
W2183
SP
+
pen
sp. + pen
meth⁻ thre⁻
W2097
meth⁻ isoleu⁻
W2180
meth⁻ val⁻
W2181

1/26/53 Monday -

- Recovering -

518tB92 λ - examination for lytic phage present.

streaked against 1485 = lysis
902 = no lysis } suggests lytic of λ - not λ-2

DI lysate - Examination

		no. pap	
1.	518 alone	c. 30	
2.	518 + DI lys	solid smear - 10^4 pap - suggestive of lysis	
3.	902 alone	smeared - 10^4 pap - lysate added by accident?	902 contaminated ☐ (+)
4.	902 + DI lys	smeared - " " "	
5.	2050 alone	c. 10	
6.	2050 + DI lys	solid smear - 10^4 pap	

~~7. 518tK12~~

518 for the purpose of examining a larger no. 518tK12 gal-segs.

1.	control alone	c. 30
2.	0.1 K-12 (12/21)	c. 500 (½ days incubation)

Blue contaminant on EMB

EMB loc - blue colonies - indicating ach. mal+ no use of lac?
lac Sm " " " " " " " " " abs. 5th
mal - " " " " " - no use of mal?
ygl - light blue colonies - utilization of ygl?

518 No prod. as function of cell no's.
Adsorption expt.

518 cell samp. → 0.1 × 10 × 100 × 100 × 125 = 6³ 69 ?

$66 \times 10^8 \times 1.28 = 84.5 \times 10^8 = 8.5 \times 10^9$

Tube	Cells added	Saline	K-H/Gal	Centrifuge/decant	No. reps	No. cells induced
1.0 →	1	1.0 ml	0.2 ml		6	4.2×10^9
	2	5 (1-2)		rapid	2	2.1×10^9
	3	5 (1-4)			8	1.15×10^9
	4	5 (1-8)		1.0 ml	5	5.75×10^8
	5	5 (1-16)		Assay	5	2.87×10^8
	6	5 (1-32)		0.1 ml	20 (nitrogen max)	1.73×10^8
	7	5 (1-64)			12 bkgrd?	7.1×10^7
	8	5 (1-128)			14	

no bkground assay mode at c 20.

tube = 71 - (bkgrd) × 10 ×

518 × 892

	Exp'gal	Prototrophs (+)	(−)	total
Cross plates	1.	0	4	4
base	2.	2	2	4
cysteine	3.	1	1	2
added	4.	0	1	1
	5.	0	6	6
	6.	1	1	2
	7.	0	1	1
	8.	0	3	3
	9.	0	3	3
	10.	1	3	4
		5	25	30

5+/30 = 0.166 = 16.6%

~~mus~~ B8 lysate
A3 Turn
{ 519 absorpt
{ 1912 absorpt

mus 519 K01 –
2050 mutants

behavi

123
124, 1
125, 126

Make Turs

{ 519 Hp^5 gal_4^- gal_3^+
{ 892 Hp^+ gal_4^+ gal_3^-

1. Semi A lysate – A1, ~~A3~~ ~~4k~~ –
2. Cross (1A36) X A1 (A3) (A11) (A13)
2. ~~Transduce~~ ~~1, 2, 3, 4~~ A1, A3, A11, A13
4. Make lysate K-12, B8 → strk out B8 test sub-line
5. (519 absorpt.
6. /

→ cold mus → K-12 (B8-1) → Centrifuge decant

1.	1.0 ml	(10X enc)	0.1 ml	//
1-2	1.0 ml + 1.0 ml sal		//	//
1-4			//	//
1-8			//	Resusp'd
1-16			//	1.0 ml
1-32			//	Sal
1-64			//	//
1-128			//	//

– plate 0.1 ml

Count X DF // total ml /

1-10 (0.1 + 10
1-100
1-1 → 0.1 ml for count

Expt. A SIBtK-12 → seg (-)

A 3 X 1436 EMS gal

	(+)/total
1.	0/212
2.	0/192
3.	0/194
4.	0/172
5.	0/205
	975

A 11 X 1436

	(+)/total
1.	0/179
2.	0/189
3.	0/273
4.	3/891
5.	0/216
	1048

A 13 X 1436

1.	c. 300 (-)
2.	c 300 (-)
3.	c. 250 (-)
4.	c 200 (-)
5.	0/195
	1250

In all of these suggestion of lysis in some colonies - tendency of some colonies to appear flatter also - pale pink -

SIBtK-12 lysate -

$10^8 = 17,25$

Wednesday 2/4/53

518 (H-), 1412 (Lpz-)

preliminary adsorption

K-12 λ (12/20) used - 1.0 ml added + 20 ml cmc {518 / 1412} to sed. Cells stored in broth overnight.

518 10^6 = c. 600,000 = 6.0×10^8 λ/ml ∫ sup.

1412 10^8 = 3,1 = 2×10^8 λ/ml ∫ sup.

initial λ titer = 2.3×10^{10}

Better check this - looks like typ. R adsorbs

750 λ titer (unfiltered prep)

10^8 = 28,94 = $\frac{122}{L} = 6.1 \times 10^9$ lousy assay replates

A3 λ titer (A = 518 CK-12 → (-) seg A3

10^7 = c. 500,500 = 5×10^9

D1 λ titer (D = 518 CK892 → (-) seg D1

10^7 = 64,0

0.5 ml D1 + Pen ——— sterile —

Thursday 2/5/53

$4p^+ gal_4^- gal_3^+ \times 4p^+ gal_4^+ gal_3^-$

518 X 892 in EMB gal · no S added – an aerated culture of 892

	(+)	(–)	total
1.	0	3/18	3/18
2.	0	19	19
3	1	23	24
4.	0	19	19
5.	0	21	21
	(1)	100	101

$101 \overline{)1.000}\ 0.0099$ (909 / 910) = 0.99 %

see pg 163 there recomb = 16.6%

this (+) confirmed on EMB gal

750 X 892 in EMB gal – no S added

1.	0	19	19
2.	0	19	19
3	1	25	26
4.	0	27	27
5.	18	22	23
	1 (18)	112	114

$114 \overline{)1.000}\ 0.0087$ (912 / 880) ~~= 0.87 %~~ 0/119 %

these not confirmed on EMB – both (–)

518 transductions.

	EMB gal	Additions	Pop. after 2 days
	1.	none	40
	2.	518 t 892-1 und.	solid smear
	3.	518 t 892-1 1-10	$>10^4$
	4.	" -1 1-100	1/2 = c. 180 · 2080
Exp.	5.	DI und.	Solid smear
	6.	" 1-10	" "
	7.	" 1-100	1/2 = c. 200 : 6400
			← evidence of lysis – plaques around margin
518 t 892 (–) exp	8	A1 und	9 (lysis?)
used for →	9.	K-12 (W/00) und.	1/16 = c. 120 = 1920 no evidence of lysis – calibrator

$2.0 \times 10^3 \times 10 \times 10^2 = \frac{2.1 \times 10^6 \text{ transductants/ml}}{1.8 \times 10^9 \lambda / \text{ml}} = 1\text{ t} / 8.6 \times 10^2 \lambda$

$6.4 \times 10^3 \times 10 \times 10^2 = \frac{6.4 \times 10^6 \text{ trans/ml}}{9.6 \times 10^9 \lambda} = 1\text{ t} / 1.5 \times 10^3 \lambda$

$\frac{1.9 \times 10^4 \text{ t}}{2.3 \times 10^{10} \lambda} = 1\text{ t} / 1.2 \times 10^6 \lambda$

2/5/53
Thursday

2175 transduction

EMB gal	Addition		No Pap. 2 days	1 day plate
1.	none		13	
2.	518t892-1	und	$>10^4$	
3.	"	1-10	$\frac{1}{16} = 85 = 1360$	
4.	"	1-100	$\frac{1}{2} = 138 = 276$	
5.	D1	und	19	
6.	"	1-10	9	
7.	"	1-100	19	
8.	A3	und	29	

$\frac{1.4\times10^3 \times 10 \times 10}{1.9\times10^9\,\lambda} t = \frac{1.4\times10^5}{1.9\times10^9} = 1\epsilon / 1.3\times10^4\lambda$

$\frac{2.76\times10^3\times10\times10^2}{1.9\times10^9\,\lambda} t = \frac{2.76\times10^5}{1.9\times10^9} = 1\epsilon / 7\times10^3$

D1 + 518t892 ↓ 1 day gal$_2$ - by t test not gal$_2$ by cm

apparently no effect — if D1 is like A4 and the other 1 this ✓ with ability to be transd.

D1 ~~not~~ tr. by 902 λ

indicates D1 is gal^2 1/19/53

2050 transduction

EMB gal	Addition		No. Pap 2 days	
1.	none		22	
2.	518t892-1	und	$>10^4$	
3.	"	1-10	$\frac{1}{32} = 121 = 3872$	
4.	"	1-100	$\frac{1}{16} = 85 = 1360$	
5.	D1	und	solid smear	
6.	"	1-10	" "	
7.	"	1-100	$\frac{1}{16} = 93 = 1488$	

$\frac{3.8\times10^3\times10\times10}{1.9\times10^9\,\lambda} t = \frac{3.8\times10^5}{1.9\times10^9} = 1\epsilon / 5\times10^3$

$\frac{1.4\times10^3\times10\times10^2}{1.9\times10^9} = \frac{1.4\times10^6}{1.9\times10^9} = 1\epsilon / 1.3\times10^3$

$\frac{1.5\times10^3\times10\times10^2}{1.6\times10^9} = \frac{1.5\times10^6}{9.6\times10^9} = 1\epsilon / 6.9\times10^3$

750 transduction

EMB gal	Addition		No. Pap 2 days	
1.	none		1	
2.	518t892-1	und.	solid smear	
3	"	1-10	$>10^4$	
4.	"	1-100	$\frac{1}{32} = c.100 = 3200$	
5.	D1	und	solid smear	
6.	D1	1-10	$>10^4$	
7.	D1	1-100	$\frac{1}{32} = c.200$ 6400	

$\frac{3.2\times10^3\times10\times10^2}{1.9\times10^9} = \frac{3.2\times10^6}{1.9\times10^9} = 1\epsilon / 5.9\times10^3$

$\frac{6.9\times10^3\times10\times10^2}{9.6\times10^9} = \frac{6.9\times10^6}{9.6\times10^9} = 1 / 1.5\times10^3$

Sunday 2/8

Crosses · 518 X 892 — mixed cell-strains — gal$_9$- x gal$_3$-

EMS gal.	(+)	(0)	total
1.	0	1	1
2.	1	12	13
3.	0	5	5
4.	0	5	5
5.	0	6	6
6.	0	3	3
	1	32	33

1(+)/33 = 3.8 % (+) recomb.

p. 165, 0.99 %
p. 163, 16.6 %

Crosses 518 X 902 — gal$_9$- ~~x~~ gal$_2$-

EMS gal			
1.	13	216	229
2.	8	218	226
3.	8	170	178
4.	19	171	190
5.	19	160	179
	67	935	1002

0.0667 = 6.67 %
1002 / 67.00
6012
6880
6012
7680

probably not valid - 902 contains (+) cells -

D1 = 518 ⊂ 892 → (-) segs. = gal$_2$- by transd.

D1 X 902

4/4 strains after 3	EMS gal 1.	11	234	245
	2.	7	170	177
		18	404	422

0.0426 = 4.26 %
422 / 18.00
16 88
1120
844
2760

probably not valid 902 contains (+) cells

confirmed on EMS gal

D1 X 892

1.	1	5	6
2.	0	3	3
	1	8	9

0.111 = 11.1 %
9 / 1.0
9
10
1

Stability examined

confirmed on EMS gal

1st streak - mixed + , =
2nd " - "
3rd " - "

D1 X 1436

1.	0	166	166
2.	0	153	153
3	0	128	128
	0	447	447

0/447 = < 0.22 %

518 X 897 — 165 = Arg+. F

Monday 2/9/53

518 X 892. prototrophs - all dad (-) except #1 gal (+)

	Aux	/1985		λ	Rx	/1985
1.	S	n. lys	14.	+		lys.
2.	S	"	15.	S		n. lys.
3.	S	"	16.	S		n. lys
4.	S	"	17.	+		lys
5.	S	"	18.	+		lys
6.	+	lys	19.	S		n. lys
7.	+	lys	20.	+		lys
8.	S	n. lys	21.	+		lys
9.	+	lys	22.	S		n. lys
10.	+	lys	23.	S		n. lys
11.	+	lys	24.	+		lys
12.	S	n. lys	25.	S		n. lys.
13.	S	n. lys				

← blue on EMB(0) - does this indicate a double (-) ?

14 S Lp^S = 13 Lp^S gal-
11 lys Lp$^+$ = 11 Lp$^+$ gal-

B8 transduction

EMB gal	Addition	No. pop 3 days
1.	none	75
2.	0.1 2050 λ - 1	106
3.	0.1 750 λ - 1	556

D1 transduction

EMB gal	Addition	No. pop 3 days
1.	none	101
2.	0.1 2050 λ - 1	120
3.	0.1 750 λ - 1	707

D4 transduction

EMB gal.	Addition	No. pop 3 days
1.	none	46
2.	0.1 2050 λ - 1	100
3.	0.1 750 λ - 1	536

518X892 pg 166 Leakage already not possible

(+) proto troples picked all grew 40/40 — (-) proto troples picked 8/40 grew — why?

one picked all three

578X892 EXPT F: Examination of (-) proto troples from the cross for their transmission behavior

Proto. #	80 adult	811 λ-9	2050 λ-1	Comment	Stockable Locus	λ-2	Lp Chas. 1967
1	Gal + proto troples —					S	—
2	c. 400 (large small)	c 100 large small	c. 200 large small		?	S	S
3	63 (like 4)	38	68	Lp_2^R ? no	? — 4-	S	S
4	45 (l. 4-)	15	35	Lp_2^R ? no	? — 4-	S	S
5	38 (like 4-)	21	49	Lp_2^R ? no	? — 4-	S	S
6	13 (like 3-)	96	4		3-	S	lys
7	13 (like 3-)	118	15		3-	S	lys
8	29 (like 4-)	11	16	Lp_2^R ? no	? —	S	S
9	28 (rocks like 4-)	29	39	Lp_2^R ?	? —	S	lys —
10	2 (like 3-)	40	2		3	S	lys
11	117 (rocks like 3-)	~~527~~	171		~~3~~ 3	S	lys
12	47 (like 4?)	15	57	Lp_2^R ? no	? — 4-	S	S
13	99 (rocks like 4-)	21 many small	76		? 4-?	S	S
14	3 (like 4-)	11	5	Lp_2^R ?	? —	S	lys —
15	c 300 (large small)	c. 100 Lp S	c. 100	Lp_2^R ? no	? — 4-	S	S
16	53	14	61		? — 4-	S	S
17	0	36	0		3 —	S	lys
18	0	0	0	minute film 2 tiny Lp.	3 see pg. 1706	S	lys
19	57 (like 4)	25	44	Lp_1 ? no	? —	S	S
20	3 (like 3-)	122	2		3	S	lys
21	1 (like 3-)	202	2		?	S	lys
22	30	21	49		? 4-?	S	S
23	c. 400 large small	c.100 (l. 6)	c. 300 (l. 60)		? — 4-?	S	S
24	3 (like 4-)	11	5	Lp_2^R ?	? —	no growth	lys —
25	37 (l. 4-)	29	68		? 4-?	S	S

blue blue — 8/8 stockable after 2 — 8/8 stockable after 2 — blue blue — 3/8 unstable after 2 — 2/8 unstable after 2 — 8/8 stockable after 2

Lp cross contains stable about appear. of plate-lys appears.

8 clear cut 3- and Lp^+

gal- λ cross gal+ has a tendency to lower the no. of outcrosses pos. On this basis nos 2, 3, 4, 5, 8, 12, 13, 15, 16, 19, 22, 23, 25 called be 4-

8 3-
13 possibly 4-
4 unknown

Wednesday 2/11/53

Transductions - Spot Method

Trial 1 - Using K-12 12/20 λ

	no. pap.
518 - 0 add	0
+ K-12 λ	61
811 - 0 add	0
+ K-12 λ	15

Trial 2

518 - 0 add	0
+ K-12	66
811 - 0 add	1
+ K-12	32

B8 = 518t902 ⟶ (-) seg

14/14 spontaneous papillae stable thru 3 purification streaks -

B8t750 unstable

D1 = 518t892 ⟶ (-) seg gave - by tr and does not ~~not~~ gave - by ~~K~~ Gal2 ~~dr~~ possibly gave - by cross K 1436

7/10 16/18 spontaneous papillae

unstable after 3 pickings

D1t750 unstable

D1X892 (+) prototrophs unstable ~~still~~ after 4

D4 = 518t892 ⟶ (-) seg ~~same~~ as D1

13/13 spontaneous papillae stable ~~thru~~ + after 3 pickings

D4t750 unstable

Tuesday 2/17

750XY-10 & obtain gal,- prototroph:
c. 2(-)? /300 prototrophs.

Lysates:

750-2 (2 bottles)
10^8 = 312, 296 = 3.0×10^{10}

811-5
10^8 = 49, 49 = 4.9×10^9

B8 - 578 T 902 (+) seg
10^8 = 699, 785 7.43×10^{10}

Expt. G : DI apart. Std (unstable) (+) segregants to observe their pattern of transduction.
Numbers 1-5

Cr H - 578 tlc 12 (+) seg tested against 2-, 4-

04 T 750
10^7 dil = >1000 titer > 10^{10}

Friday 2/20/53

Expt. G. 01 = 578t 892 → (-) seg. By transducability 6st = 2-
" donor " = 2-
" X 902 ≠ 2 ⇒ the mice 902 cont. (+)

01 spont. gal+ are unstable.

5(-) seg from 01 - from seg segregation.

G-1, 2, 3, 4, 5 - Addition		No. gal 58	days G-2 12	G3 25	G4 75	G5 10
1	none					
2	758A-1	1/4=38 304	1/4=69 552	1/4=69 276	1/4=91 364	1/4=49 392
3	702A-2	59	16	32	32	28
4	2080A-1	108	42	67	62	47
5	811A-5	102 vv	49 ✓	65	55 ✓	53
Prob. locus		2-	2-	2-	? 2-	? 2-

Streak of middle plate
1.
2.
3.

G-2-3 1. 7/8 mix, 1(-)
2.
3. [illegible]

G-2-4 1. 7/8 mixed, 1(-)
2. 4/7 mixed,
3. 7 medicals transduct. by 4-

G-2-9 1. 8/8 mixed
2. 6/8 mixed
3. 6/8 mixed
(some of these may be slows)

G-3-9 1. 8/8 mixed
2. 7/7 mixed
3.
medicals transd. by 4-

G-5-2 1. 5/8 mixed, 1(-)
2. 5/5 (+) ↓ stable
3.

G-5-2 1. 4/8 mixed, x(-)
2. 6/6 (+)
3. ↓ medicates no. 2 - transl.

Friday 2/20/53

Exp. H. = 578 tK-12 → (-) seg from 21 separate transductions

No.	Add			Probably lives	
1.	27	43	1/4=36 144	4-	pure(+) on 1st streak -
2.	29	32	1/4=42 168	4-	
3.	35	29	1/4=27 216	4-	
4.	15	22	1/4=61 244	4-	
5.	32	36	1/4=71 284	4-	
6.	32	42	1/4=73 292	4-	
7.	31	30	1/4=56 224	4-	
8.	28	37	1/4=45 180	4-	pure(+) 1st streak -
9.	(+) cell contamination ?			(?)	
10.	17	27	1/4=54 216	4-	
11.	27	31	1/4=42 168	4-	
12.	(+) cell contam ?			(?)	no add plate 1st (appears seg.)
13.	36	23	1/4=63 252	4-	
14.	30	33	1/4=59 236	4-	
15.	26	31	1/4=55 220	4-	
16.	27	31	1/4=42 168	4-	
17.	(+) cell contam ?		-	(?)	no add plate 1st (appears seg)
18.	32	31	1/4=59=236	4-	
19.	31	39	1/4=61 244	4-	
20.	31	24	1/4=64 256	4-	
21.	40	44	1/4= 52 208	4-	

All lysogenic prime λ+ reactivated

H-9-0 - 8/8 stable after 2 isolations
H-17-0 - 5/5 unstable after 2 "
H-12-0 - 6/6 unstable after 2 "

} probably contamination occur on purification (selecting a (-) colony) and replating - plating out on EMB gal and picking spot + papillae no unstables found after 2 - No (+) sp.th/pop about 30-40/plate

From the transduction data these (-) seg appear to be 4-

18/18

Test some of them for λ + / for allele test

Friday. 2/20/53

518	Addition	No. pap 2 days
1.	None	67
2.	0.4 ± 750 $1-10^3$	24
2.	" $1-10^5$	11
4.	" $1-10^7$	80
5.	" $1-10^9$	46

2062 – isolation of double mutant.

colonies in cmd $1-10^8$ } suitable for replication to DOI + pr.
$1-10^9$

510 x 882 prototroph #18 gal- - alkaline rx on EMB gal-; B(c).

suspicious that it is 3⁻4⁻?

After 3 days	EMB gal	
1	no add	– c. 100 pap. mostly small
2.	750 λ-1	– c. 300-400 pap (larger than no add)
3.	912 λ-2	– c. " " " " "
4.	650 λ-1	– c. 75 pap mostly small
5.	811 λ-5(?)	– c. 300-400 pap larger than no add

enclosed flu. is a 3⁻ but slow in response for some reason

Thursday 2/16/53 — mixed cultures

578 X 892 B, added to X plates

EMB gal	(+)	(−)	tot
1	0	9	9
2	0	12	12
3	1	13	14
4	1	11	12
	2	45	47

Confirmed on EMB gal

$0.042 / 47 \overline{)2.0000}$ (188, 120) = 4.2%

		(+)	(−)
ρ. 166	= 3.3	1	32
ρ. 165	= 0.99	1	100
ρ. 162	= 16.6	5	25
Thin	4.2	2	45
		9	202

211

SOME OF THESE PROTO- √ ON PG 173 FOR 4, L, 2 ρ

$0.0476 / 211 \overline{)9.00}$ (844, 560, 422, 1380)

Error 4.3%

578 X 902

EMB gal	(+)	(−)	tot
1	2	381	383
2	1	278	279
3	0	341	341
4	2	284	286
	5	1284	1289

$0.0038 / 1289 \overline{)5.000}$ (3867, 11330, 10312) = 0.38%

Confirmed in EMB gal - stable after 2 p.c. isol.

811 X 902

EMB gal	(+)	(−)	total
1	0	198	198
2	0	151	151
3	0	184	184
4	0	173	173
	0		706

(Estimated < 0.14%)

$0.0014 / 707 \overline{)1.000}$ (707, 2930)

Friday 2/27/53

2068 pr - A - X 892

EMB gal	(+)	(-)	total
1	3	64	67
2	2	33	35
	5	97	102

$\frac{0.079}{5.00} = 4.9\%$

gal$_1^+$ x gal$_3^-$

NO — indicates 2000 gal$_3^-$ distrib. that gal$_2^-$ what gal's

Weakening on crosses

01 X 902 } repeat crosses to see if initial results (contradictory in that
04 X 902 } (01, 04) were gal$_2^-$ by donor + recip. transd. tests) was caused by + cont in 902

01, 04, 902 pure (-) in EMB gal showing of washed susp.

811 + 902 — to see if 811 is different from 578 as suggested in last cross (apparent linkage gal$_2^-$ gal$_3^-$)

811 pure (-) in EMB gal

2/20/53 Int. — nucleid culture

578 x 892 Prototroph-cross on pg 171.

gal$_1^-$ Lp^s gal$_3^+$

gal$_1^+$ Lp$^+$ gal$_3^-$

	Proto	gal rx	/1	/1-2	/1485	Probable on basis of gal$_3^-$
one of these possibly unstable	1	+	r	s	lys	
	2	+	r	s	lys	
	3	—	s	s	n. l.	4
	4	—	r	s	lys	
	5	—	s	s	n. l.	4
	6	—	r	s	lys	
	7	—	r	s	lys	
	8	—	r	s	lys	
	9	—	s	s	n. l.	4
	10	—	r	s	lys	
	11	—	s	s	n. l.	4
	12	—	r	r	lys	
	13	—	[illegible]	s	[illegible] lys	
	14	—	s	s	n. lys	4
	15	—	r	s	[illegible] lys	
	16	—	r	s	[illegible] lys	
	17	—	r	s	[illegible] lys	
	18	—	r	s	[illegible] lys	
	19	—	r	s	lys	
	21	—	r	s	lys	
	21	—	r	r	lys	
	22	—	r	s	lys	
	23	—	r	r	n. lys	
	24	—	s	s	n. lys	4
	25	—	s	s	n lys	4
			7 s	23 s	8 n.l.	
			18 r	2 r	17 lys	

D1 x 892 - unstable + prototroph.
stable neg (-) on EMB gal after 5 p.c. isolation
homogeneous (-) on {EMB / Loc}

578 x 892 - above (+) possible unstable
s.c. 1. mixed (+),(-)
2. mixed (+),(-)

Sunday 3/1/53

1655 X 1177 on EMS gal to obtain gals - typ^s
25 gal - tested against t 2 - all typ^R

811 X 902 on EMB gal - purity of parent checked on p. 172

	(+)	(−)	total
1.	0	65	65
2.	0	41	41
3.	0	32	32
4.	0	62	62
	0	200	200

total to here 0+/907 = (< 0.11%)

0.0011
907 / 1.000
907
930

D1 X 902 on EMS gal - to recheck on D4 since 902 culture could have been contaminated with (+) cells.

	(+)	(−)	total
1.	0	209	209
2.	0	265	265
3.	0	134	134
4.	0	265	265
	0	873	873

0/873 not in agreement with previous

D4 X 90 as D1

	(+)	(−)	total
1.	0	248	248
2.	0	154	154
3.	0	175	175
4.	0	122	122
		699	699

0/699 = not in agreement with earlier results - probably correct - test D4 for gal - neo

D1 from duchess = mutant ?

Thursday 3/6/53

Crosses with 902 B7, B8, D1, D4, 811 all checked on EMB gal all found (-)

811 X 902

EMB gal	(+)	(-)	total
1.	0	77	77
2.	1?	49	50
3	0	52	52
4	0	105	105
5.	0	75	75
	1?	358	359

1? confirmed on EMB gal

total to here

	(+)	(total)
p. 171	0	706
p. 174	0	200
above	1?	359
		1365

1365 / 1.0000 = 0.00073 = 0.073%
9555
4450

578 5/1289 = 0.38%
6/2654

Recheck on crosses with 902 - 902 pure gal -

D1 X 902

EMS gal	(+)	(-)	total
1.	0	273	273
2.	0	339	339
3.	0	247	247
4.	0	296	296
			1155

0/1155

D4 X 902

1.	0	1/8 = 237	1896
2.	0	1/8 = 249	1992
3	0	1/8 = c. 300	2400
4.	0	1/16 = 109	1744
			7932

0/7932

B7 X 902

1.	0	1/2 = 264	528
2.	0	1/2 = 292	584
3.	0	1/2 = 239	578
4.	0	1/2 = 271	542
			2232

0/2232

B8 X 902

1.	0	1/2 = 203	406
2.	0	1/2 = 176	352
3.	0	1/2 = 191	382
4.	0	1/2 = 227	454
			1594

0/1594