Dear Lillian,

We keep all our media in the form of dry mixes, and have found this very convenient and accurate, especially from the point of view of having untrained personnel make up occasional small batches.

A) The dyes:
- $\text{K}_2\text{HPO}_4$ (anhyd) 200 g.
- Rosin Y 40 g.
- Methylene Blue, HCl 6.5 g.

Use in a final amount of 2.4 g/l.

This can be added either to a complete or a synthetic agar. Our "complete" is:

(per l.):
- Casein Digest (\text{\textquotedbl} \text{\textasciitilde} N-Z-Case\text{\textquotedbl}) 8 g.
- Yeast Extract 1 g.
- NaCl (not essential) 5 g.
- Lactose 10 g.
- Agar 15 g.

The synthetic used for this purpose is:

- Sodium Succinate 5 g.
- $\text{(NH}_4\text{)}_2\text{SO}_4$ 5
- NaCl 10
- MgSO$_4$ 1
- Agar 15.

We have tried a number of other non-acid-producing C sources, and of them asparagine is the only one that can compete with succinate. The synthetic "KMB", or "KMS" as we call it is a little trickier than the complete, but there is usually no difficulty in reading + or - colonies when grown at 37, 36-48 hours.

"either and I will be back the end of next month, and hope to see you then.

Best regards,

Sincerely,