3/6/53 Friday

Exp. J Examination of protoplasts from cross of 578 x 892, Gal-

Protoplasts	Budded	881/1-5	892/1-2		Lp Rx	
1. } Galt		1st smear	2nd ✓			
2. }						
3.	52	18	S. Smear	4-	S	
4.	2	153	S. Smear	?	Lys	J-4-4 8/8 Stable after (2)
5.	24 *	42 *	Solid Smear	4-	✓	
6.	72	347	S. Smear	?	Lys	J-6-4 8/8 stable after (2)
7.	44	227	S. Smear	?	Lys	J-7-4 8/8 stable after (2)
8.	1	6	S. Smear	4-	Lys	J-8-4 6/6 stable after (1)
9.	40	21	S. Smear	4-	S	
10.	41	36	S. Smear	4-	Lys	
11.	37	11	S. Smear	4-	S	
12.	23	21	29	Lp2R	✓	
13.	1	8	S. Smear	4-	Lys	J-13-4 8/8 stable after (2)
14.	39	10	S. Smear	4-	S	
15.	2	61	S. Smear	?	Lys	
16.	77 *	211	S. Smear	?	Lys	J-16-4 8/8 Stable after (2)
17.	33	60	S. Smear	4-	Lys	J-17-4 8/8 Stable after (2)
18.	23	½=208	S. Smear	?	Lys	J-18-4 8/8 stable after (2)
19.	2	11	S. Smear	4-	Lys	
20.	1	7	S. Smear	4-	Lys	J-20-4 8/8 stable after (2)
21.	4	9	14	Lp2R	✓	
22.	1	17	Si Smear	4-	Lys	J-22-4 7/7 Stable after (2)
23.	30 *	35 *	46 *	Lp2R	✓	
24.	87	12	S. Smear	4-	S	
25.	30 *	18	S. Smear	4-	S	

Carried over from smear

* small in bkgrd.

14	4-	6/14 S
6	?	
3	Lp2R	

Mixed cultures

(1765)

3/6/53

Transductions

			N. pfu 3 days
811	EMB 3rd		
	1.	no add	43
	2.	B8	122
	3.	D4t750	42
	4.	A3	47
2050			
	1.	no add	16 (+ some small)
	2.	B8	1/4 = 110
	3.	D4t750	1/2 = 210
	4.	A3	1/4 = 177
750			
	1.	no add	2
	2.	B8	78
	3.	D4t750	2
	4.	A3	18
2175			
	1.	No add	24
	2.	B8	18
	3.	D4t750	12
	4.	A3	64

Tuesday 3/10/53

518 transduction with high activity ~~Bf~~ phage.

0.1 ml of saline resusp 518 cells (overnight culture)
0.9 ml of 01 lysate - 1 - 1 hr at 37 1.0 ml - dil

10^{-2}, 2×10^{1} 0.1 ml → 10 plates.

	unstable (+)	(-)	total
1	72	1708	1780
2.	83	1/4 = 443 (1772)	1855
3.	77	about the same	
4.	83	" " "	
5.	89	" " "	
6.	95	" " "	

remainder about the same

} 24 of these (+) streaked out - all mixed - many mosaic col.

1708 | 72.00 (0.04)
(68 20)
(580)

c 4.5 × 10 cells transduced

6 plates of 518 untreated dil to contain c. 150 colony/plate showed no plus colonies

1 plate of undiluted 518 also showed no (+).

Extended Examination of protoplasts from 518 x 892 mixed culture. Exp J.

J 4 (-)

1. no add	7	
2. 750 λ-2	22	
3. 902 λ-2	1/8 = 66 528	} says it probably gal, - ?
4. 892 λ-2	Solid smear	
5. 811 λ-5	1/8 = 70 = 560	

J 6

1. no add	c. 50	
2. 750 λ-2	1/8 = 73 584	
3. 902 λ-2	1/8 = 87 - 696	} says it a new locus
4. 892 λ-2	Solid smear	
5. 811 λ-5	1/8 = 68 534	

J 7

1. no add	21	
2. 750 λ-2	1/8 = 57 456	
3. 902 λ-2	1/4 = 107 428	} says it a new locus
4. 892 λ-2	Solid smear	
5. 811 λ-5	1/8 = 72 576	

(1776)

3/10/53

1655 (−) gal transduction ~ 892 λ
0.1 ml 1655 saline suspended (aerated overnight) + 0.9 ml 892-2 λ
- incub at 37 C 10 min dil 10^1, 10^2 → 0.1 ml / 10 plates EMB gal

	tot. no. clones	no. mosaic col.	
1.	155	5	3.2 % mosaic
2.	294	8	2.7 %
3.	not about the same		

2 plates made to indicate the amt. of phage added to the gal plates indicate about 10^3-10^4 λ / ml

Lysate sterilities

892-2 bottle 1 — sterile 0.5 ml
01-1 0.2 ml sterile
04-2 bottle 1 sterile 0.5 ml
01-2 ~~bottle 1~~ not sterile 0.5 ml. — Re chloroform

1655 † about 27 mosaic colonies picked and streaked out — no segregg of ± clones evidently ± deletion not easily accomplished

3/17/53 Tuesday.

Transduction: To identify prototrophs from 578 x 892

750	EMB gal	Rep 2 days	
	1. no add	4	
	2. J4	6	
	3. J7	3	
2175			
	1. no add	8	
	2. J4	26	
	3. J7	30	
2050			
	1. no add	1/4 = 56 = 224	
	2. J4	1/4 = 58 = 232	
	3. J7	1/4 = 41 = 164	
	4. 892 λ-2 (1-110)	> 2000	
	5. D2 (presumed 4-) Say 5H+H2	1/4 = 83	332
578			
	1. no add	33	
	2. J4	18	
	3 J7	23	
	4 D2	15	

J4, J7 } appear to transduce gal+ - only what this means I don't know

J4, J7

probably says D2 is gal+ altho duction of 2050 is low (hi bkgd?) also this culture is lysogenic for "normal" λ - not hi-act λ

1485 - Attempt to transduce to hi act λ from 892 - Rep 2

0.2 ml of resusp'd overnight culture c. 10^9 + 0.5 ml 892 λ-2

5 Control plates EMB gal: c. 400 colonies/plate - some intermediate gal+ & possible mosaic colonies - streaked out. On streaking showed + and slow + forms.

5 Expt'l Plates EMB gal: c. 80 colonies/plate - represents about 80% killing?
c. 2# mosaic colonies - streaked out
$\frac{0.04}{400/35}$ = c. 4% ← of the 24 colonies - 1 segregating + on 1st streak

To avoid selection against + (-)

5 Expt'l plates EMB(0): c. 80 colonies/plate -
31 colonies showing evidence of hi

3/21/53 Sat.

Exp K 811 & 892 (+1 segregant analysis)

On EM13 gal

Seg #	Sw add	892 1-2 (1-100 del.)	811 1-5	Probable locus	
1.	31	c. 2000	33	4	
2.	21	c 3000	54	?	3/8 mucid 2 3/8 mucid 3
3.	71	c 2000	68	4	
4.	~~31~~ 32	c 2000	~~33~~ 63	?	3/8 mucid 2 3/8 mucid 3
5.	26	c. 2000	28	4	
6.	26	c 2000	29	4	
7.	17	c. 1000	59	?	4/8 mucid 2 3/8 mucid 3
8.	37	c. 2000	32	4	
9.	36	c 1000	72	?	
10.	27	c. 1500	31	4	
11.	32	c 1500	48	4	
12.	28	c. 1500	33	4	
13.	24	c 1500	22	4	
14.	24	c. 500	54	?	4/8 mucid 2 3/8 mucid 3
15.	12	c. 1000	45	?	5/8 mucid 2 5/8 mucid 3
16.	33	c. 1500	19	4	
17.	47	c. 2000	43	4	
18. (switched)	28	c. 2000	21	4	

892

r on the Stability of spontaneous pap - after 2 all appear pure +

Exp D

Instability of DI spontaneous pap. confirmed (again) 3/8

stability of D4 " " " " 8/8

2050 - from EMC - transduced by 892 1-2 to high frequency

892 - from EMC - ~~low~~ low - - colony) distribution about the same
form) as own 892

- own abs low - ——— small col form + spreading colony form

Sun
Mon: 3/22/53 - 3/23/53

518 Perfect stage I transduction as a function of multiplicity -

2050 Exp -

892 - streaked out EMB line - appears homogeneous lac- mucin - 2 colony types - small round, large spreading. well isolated of each picked, lysates made 892-isol 1 / 892-isolate 2 } don't know which is which! Obtain from culture

2050 Exp is these lysates.
518

cell	lysate →	isolate 1	isolate 2
2050 E	c. 3	892 lysis 150?	lysis ?

c. 200 plaques plus gross lysis - transduction difficult to assess - mucoid? Stock and use extent of gal + recs

518 c. 3. c. 20 no plaques & mucoid? → c. 2 gal

Negative not streaked

dilute with 1-100 dil. c. 10 pfu 1-5 plaques.

no transduction to gal + in 8 different areas

Results suggest cool 1+2 are gal + ?

518 + 2050 - 3 (purified) stocks made of separate transductions not a high incidence of transduction 2050 → × 518

3/24/53
Expt L stored on EMB gal plate

518 - one plate - transduced ε 892 lysogen to see if transinduced are M+ replica (slightly) to D(0) - no growth D(0) - no prototrophs

3/16/53
Thursday

892 - All confused - probably complicated by a switch in media.
Apparently 892 mixed with gal+, gal−? possibly (−) of
a couple of types.

578 Examination of 04 to see if activity of A present.

1. to add	=	12
2. 04 A	=	>10⁹

Summary 518tK-12 See pages 170a

page

1. Examination of a single gal- segregant from ~~[illegible]~~ separate transductions of WS18 gal4- by lysates of K-12

		Receptor test No. of ~~[illegible]~~	No. transd. by gal4- lysates	No. transd. by gal2- lysates	Probable locus by receptor test
170a	A.	22	0	22	22 gal4-

1. (A3)

2 - A1

3 - A4

4 - A3

2. Examination of a number of gal- segregants from a single transduction

		Receptor test Segregant	No. transd. by gal4- lysates	No. transd. by gal2- lysates	Probable locus by receptor test
145	A.	15	0	15	15 gal4-

163 B. Cross test (X1426)

	Seg no.	No. prototrophs	No (+)	Probable locus
1.	A3	975	0	gal4-
	A11	1048	0	gal4-
	A13	c. 1250	0	gal4-

C. Donor Test

	Lysate of Seg	Transductants gal1-	gal2-	gal3-	gal4-	Probable locus
165, 1766	A1	+	+	+	no	gal4-
1766 1.	A3	+	+	+	no	gal4-

Summary of 518t902

page

1. Examination of ~~a series of~~ single gal- segregants from different transductions.

2. Examination of a series of gal- segregants from a single transduction

A. Reception test

No of Seg.	No transd. by gal+ lysate but not by gal-	No transd. by gal- lysate Obtained from gal+	Probable locus
19	2	17	2 gal-, 17 gal+

146

B. Crossing test

1. Seg. gal- by reception test (K902)

Seg. no.	No prototrophs	No. (+)	Probable locus
B7	c. 2200	0	gal-
B8	c. 1600	0	gal-

175

2.

C. Donor Test

Lysate of Seg	gal+	Transducers gal-	gal+	gal+	Probable locus
B7					
B8	+	no	+	+	gal-

176B

76
16
446
76
1216

Sunday 3/29/53

Analysis of 892 mutants

811

892-1 } = 2 (-) gal colonies from stock of 892 - lysate made
892-2 } of these - both are loc - TLR⁻

811	Agr 2 days
1. no add	28
2. 892-1	39
3. 892-2	33

} these lysates appear to have at least 2 plaques in them - 1 resembling 1, other giving plaques on 1+ culture

2175 for the purpose of ~~segregating~~ reisolation of 518 892

1. no add - 6
2. K-12λ 30-1 176 — pick 24 for analysis

750 For the purpose of picking transductants for analysis - 8 segregants.

Monday 3/30/53

1. no add	2		
2. K-12λ 30-1	1/8 = 18	342	
3. 902λ-3	1/4 = 29	116	

518 For the same purpose as 750 above

1. no add	28	
2. 750λ-2	79	
3. 902λ-3	1/16 = 76 = 1216	
4. K-12λ-30-1	1/8 = 65 = 280	

5. filtrate of culture of 1485-01 } 57 ← indicates that if λ from "natural" lysis is not very active - assuming high titers a few plaques resembling 1-2

518

a second mutant to test a lysate of gal + from mixture 892

7 day incub

1. no add	< 20	
2. lysate of 892 gal+ colony 3a purified culture of this approved mutant	7/04	high activity

Monday March 30, 1953

1673X D1 to see if (lt) from this cross have hi-aer lambda-
prods of the cross
D1 prod.-
1677- +, with slow gal present - a plus picked for purif.
and a new

4/2/53 Thursday

— 1485tD1 - the negative transductin - Examination of (-) segregants to determine their nature.

Exp L.

		No pop 2 day	Locus suggested	
L1	no add	4		
	902λ-3	19	2-	
	811λ-6	37		
L2	no add	6		Reason for low
	902λ-3	5	2-	transductn
	811λ-6	21		not known.
L3	no add	10		Data appear
	902λ-3	9	2-	ok. but better
	811λ-6	22		recheck.
L4	no add	5		
	902λ-3	5	2-	
	811λ-6	21		
L5	no add	5		
	902λ-3	8		
	811λ-6	13	2-	
L6	no add	3		
	902λ-3	7	2-	
	811λ-6	21		
L7	no add	3		
	902λ-3	8	2-	
	811λ-6	14		
L8	no add	2		
	902λ-3	3	2-	
	811λ-6	22		

all these found 5 hp and called prototrophic

892 hi powered λ - do hi prove DNAase susceptible?

578	no add	16		
S	892λ untreated (-100)	>10⁴		no
	892λ DNAase treated at 37c 10 min 1-100 x 6/5	>10⁴		

4/2/53

518 gal+- is there a diff. between 578 gal+^{R} and 811 gal+^{R} because 811 was made Lp+ 1st? Data on 811 reversion as gal+ does not decide because strains not transducing gal+ do not give (+) in crosses with Y-10. Examine 578 gal+ to find strains giving (+) with Y-10, then make lysogenic and examine transducing power.

Check in the assay cultures. Y-10 are (+)
578 gal+-1 " "
578 gal+-2 " "

578 gal+-1 × Y-10 - are + prototrophs formed
nos/plate = 65, 48, 75, 114, 67, =

578 gal+-2 × Y-10 - are + prototrophs formed
nos/¼ plate = 75, 125, 95, 87, 108

~~~

892 mixed stock - plated against A^2 - large proportion of cells Lp_2^R
Selection for Lp_2^R pertains to stock.

~~~

2175 EK-12
24 (+) ray obtained - carried thru ② purifications

518 E 750-1 } made from ② purification
518 E 750-2 }
750 E 902-1 } made from ② purification
750 E 902-2 }
750 E 12-12-2 } made from ② purify.
750 E 11-17-1 }

April 5, 1953

892(mix) preliminary examination of occurrence of transducing phage in bursts from single cells of irradiated 892

1. Growing culture of 892 diluted to contain c. 500 cells/0.1 ml
2. irradiated 15 seconds.
3. plated before and after with 750, 1485, and by it ref. to.

1. 892 cells/0.1 ml before irrad. = 585 (gal+ = 101)
2. 892 cells - plaques/0.1 ml on 1485 before = 1.
3. 892 cells - transd/0.1 ml before on 750 = 78 (from 892, 78)

4. 892 cells/0.1 ml after irrad 16.8 (gal+ = 26)
5. 892 cells/0.1 ml plaques on 1485 after irrad. ~~44~~ 247 (total = 415)
6. in 892 cells on 750 cells 35 (gal+ from 892 = 31) / 750 after turbid plaques picked

750 no add = 2

~~~

518 gal+ R #3 X Y-10 controls o.k.

	(+)	(−)
1.	1/4 = 60	240 no. (−)
2.	1/4 = 33	132 "
3.	1/4 = 46	184 "
4.	1/4 = 40	160 "
5.	1/4 = 51	204 "
		930

0(−) / 930 (+)

518 gal+ t #4 X Y-10 controls o.k.

	(+)	(−)
1.	1/4 = 53	1/2 more
2.	1/4 = 57	228 "
3.	1/4 = 50	200 "
4.	1/4 = 58	232 "
5.	1/4 = 55	220 "
		1114

0(−) / 1114 (+)

~~~

1485 ED1 — unstable gal+ ⟶ hi aer. lysates λ

save virus to stable + ⟶ low aer λ

~~~

902 st X 2175 (gal$_2$ - X gal$_2$ -) on low sm - about 200 recombinants per plate.

2251
~~~

Tuesday 4/7/53

- 1821 - vs. hi powered 892 λ

On EMBgal.

1. no add - c10
2. 892λ 1/100 - c. 1000? } appears that hi act λ has less act on 1821. - situation similar w. "normal" λ.

On EMBxgl

1. no add - many small, 3-4 large
2. 892λ - 1/100 - " " c10 "

Repeat xgl using undiluted 892.

1485tD1 - Re-examination of the possibility of (-) ductii.

1. Lysate D1d - 2 pmp.
2. Cross printed on 1485 on EMB10).
3. after 1 day lysed area picked w. 1/100 - streaked out several times on EMB3gal.
4. a single (-) colony observed
   several intermediate gal + col.
   a few segregating (what?) colonies.
   many gal + colonies.
5. some of above colonies picked and checked.
6. Results - streaks -
   a. 2 pure (-) col.
   b. several mixed?
   c. mostly gal + col.

Summary Stock cultures on Plates in Refrigerator

- 1924 t K-12 Unsuitable + 11 in all
- 811 gal + R. 10 in all
- 518 gal + R 8 in all

Expt A 518 t K-12 all exp single transductants
" B 518 t K-12 all exp from single transductants
" C 518 t 902 (mix)
" D 518 t 811 mix
" E 2050 t 811
–
–
–
Exp J
" K
" L
" M 750 t K-12 (-) exp ~~18~~ 17 in all
" N 750 t 902 (-) exp 23 ~~X~~ in all } replaced using streaked gal+ purified to weaker mixes ≈ 750 S ✓ ≈ 902 R ✓ = 220 ?
" O 2175 t K-12 (-) exp 21 in all
" P 518 t 750 (-) exp 3 in all
" Q 518 t K-12 (-) exp 13 in all
" R 518 t 902 (-) exp 23 in all
" S 811 t 902 (-) exp 19 in all
T 2175 t 750 (-) exp ~~8~~ 23
U 2175 t 811 (-) exp 16 in all
V 811 t K-12 (-) exp 20 in all
W 1924 t 902 (-) exp 18 " "
X 1924 t K-12 (-) exp 24 " "

4/9/53

1485T01 - Occurrence of hi-act λ in bursts of single cells mod.

Pre mod.

1. 1485T01 cells count per cc/ml. = 493 /0.1 ml
2. Sp. pop. 750 1
3. No. ppt in mix of 1+2. Expected 494
   Found 494
4. Plaque by 1485T01/1485 0

Post mod.

5. No. 1485T01 cells survs. 123
6. No. " cells giving plaque/1485 140
7. No. " cells giving pop. 750 123
   " Sp. pop 750 1
   Total 124
8. No. ppt mod. 1485T01 on 750 222

No of dying 1485T01 cells giving pop 750 222 - 124 = 98

Fraction of dying 1485T01 cells giving rise to 1 hi act. λ particle 98/140 = 70%

4/10/53 Friday

750 and D1 Occurrence of hi-act-1 in single bursts of D1

C. 10^3-10^4 cells of D1 - sampled before irrad., then irrad ≅ U.V. sampled immediately and after a hour incubation with aerated cult. D1

Dil to 10^3-10^4 → irrad 5 sec, 10 sec. → dil 10 sample → for cell survival / for plagues/1485 / for pap / 750

→ dil 10 with broth → sample immed. → plagues induced pap 750

↓ inc. 2 hours → sample as ↑

Preirrad 0.1 ml

1. Total no. cells = 310
2. plagues/1485 = 1
3. Pap./750 (3) 6

750 spont pap = 1.

Post Irrad.

5 second exposure.

4. Surviving cells 159
5. plagues induced/1485 40 (40 plagues / 151 surviving cells)
6. pap./750 (3) 6
7. plagues after 2hrs/1485 3344
8. pap/750 after 2 hrs. — (3) >47 (many small) upper limit = c. 600.

Plagues assays against 1485 when streaked out - centr of plague shows evidence of (+) ductin

10 sec. exposure.

9. Surviving cells 84
10. plagues induced/1485 40
11. pap./750 (8 - 15
12. plagues after 2hrs/1485 2720
13. pap/750 after 2hrs (20) >53 (many small) upper limit = c. 200

---

582 mix gal+ (also contains) lysate has hi act / 578

1485-tD1 — (-) pap

lysate probably has hi act / 578 but appears quite lysis also.

---

578 gal+ R. #5 and #6 tested in crosses X Y-10 for mimic reversion

578 gt #5 gal+ puts = 420, 650, 512, 416, 466 (all obtained by gal+/ counting either 1/4 or 1/8 plate)

< 0/2304

578 gt #6 gal+ puts: 440, 440, 432, 440, 340, 672 (also obtained by counting 1/4, 1/8 of plate) less (-) puts

< 0/2764

3/10/53
Sat.

Various transductions to secure (-) seg for classification.

		Seg 2 days
217N	1. no add	13
	2. 811λ-4	39
	3. 811λ-7	50

} (-) seg isolations made from both. discarded

750	1. no add.	2
	2. 811λ-4	10
	3. 811λ-5	14
	4. 811λ-6	33
	5. 811λ-7	25

} discard - transduce gal, prototroph and use 750s as crossing tool

518	1. no add.	29
	2. 750λ-2	72

1821 On EMB xyl -

1821 xyl No transduction?	1. no add	3	
	2. 892λ-2 (hi prototroph)	c. 150	✓ These are gal+ xyl- Apparently EMB xyl can select for gal- transduction

3/16/53 Thursday.

750s^r = 2319 test for validity

1. no add		21
2. 750λ-1		25
3. 902λ-3	1/4707	428

4/16/53

750 pr = gal$_1$ prototroph - Isolation of (-) segregants.

1.	no add	1.
2.	750 λ-1	0
3.	K-12 λ 30-1	143
4.	902 λ-3	84
5.	811-6	4

(check on authenticity)

after 2 single colony isolations 2/4 are crossed - probably indicates that stable transductions result 811 → 750

gal$_2$ -

1655 × 2238 (= loc$_1$ gal$_2$) to isolate tp_2^s from (tp^s also if possible)

Control plating	1.	1655	prec +	
	2.	2238	prec -	
		(4)	trial prec	
cross assay	3	30	220	
	4	33	211	
	5	44	433	
		107	864	

$\sqrt{107.0}$ = 0.129

864

1760

1728

3320

+ - 12.4%

Pickings for tp^s, tp_2^+ ~ tp^+

1. 5/34 tp_2^+ all tp^+

2. 30/30 tp^+ ~ tp^+ / λ

750 pr (gal$_1$ prototroph) - Irradiation to obtain tp^s from.

5 pink colonies on EMB but noted. Pick and streak

1871 purified - by transduction of single colony isolate - Stock made of this.

1.	no add	10
2.	K-12 λ (30-1)	84

1485TD1 - 2 additional segregants tested

Seg #9	1.	no add	10
	2.	750 λ-1	4-15 = 60
	3.	902 λ-3	10
	4.	811 λ-7	29
Seg #10	1.	no add	8
	2.	750 λ-1	1/16 = 54 = 864
	3.	902 λ-3	24
	4.	811 λ-7	1/32 = 55 1750

apparently gal$_2$ -

probably gal$_2$ -

Saturday 4/18/53

2175 - attempt to observe "satellite" phenomenon in 2175 found - check to see if other transductions

give 1
1. no acid 17
2. K-141 3:1 248

Examination of some lysates.

518
1. No acid 11
2. 892 Lp2 (sonic) 18
3. 892 M1 (sonic) 11
4. 891-8 11
5. 902-4 1/4 = 118 472
6. 015 1/16 = 52 896
7. 844 1/16 = 44 704

unknown virus exp. 9 →

on the gels secondary smearing?!

W2346.3

(192) A

4/25/53 Saturday. (tested by spotting 0.05ml of 1 prep on 1/2 plate - 1/2 plate serves as control) (1/2 control) Supernat. of segregants

Expt. M 750 prototrophic gal,- transduced by K-12 λ 30-1 - Examination of Segregants (-)

Segregant	No. of Segregant progs. (+/Gal)	No. prog. 750λ-2 pfu	No. prog. 902-4 pfu	Probable locus	To be taken by X	Satellite of Gal, λ
1.	7	0	6	gal,-		0
2.	1	0	10	"	✓	0
3.	4	0	9	"		0
4.	0	0	11	"		0
5.	0	0	11	"		0
6	7	0	7	"		0
7.	0	0	9	"		0
8.	0	0	6	"		0
9.	0	0	8	"		0
10.	0	1	10	"		0
11.	0	0	13	"		0
12.	2	7	9	"		0
13.	0	0	13	"		0
14.	1	0	8	"	✓	0
15.	0	0	10	"	✓	0
16.	0	0	15	"		+
17.	0	0	13	"	✓	+
						2/17

2/24/54

Stocks of this experiment discarded as not worthy of carrying

Examination of lysates M2, M14, M15, M17

	750	518	2175
1. no add	1	40	14
2. M2	3 ok ✓	29	50
3. M14	5 ok ✓	62	70
4. M15	4 ok ✓	15	33
5. M17	5 ok ✓	28	74

Test crosses of M2, M14, M15, M17 × 750 sr - Controls EMBgal 750, M14, M15, M17 ok., M2 no gen R

	(+)	(-)	total	2nd cross	total	3rd cross - Controls ok	total
M2	0(?) ok	125, 89, 115	0+/332	2, 2, 2, 3 = 11	0+/343	21, 16, 14, 13, 19 (83) (4/5)	4/26
						×2196 gal+ : 37, 57, 51, 66, (16/45)	
M14	0(?) ok	121, 126, 136	0+/383	0, 3, 2, 4 = 9	0+/392	24, 38, 37, 36, 27 = 162	0+/554
M15	0(?) ok	210, 130, 171	0+/511	9, 6, 2, 1, = 18	0+/529	×2196 gal+ : 7, 9, 13, 12, 15 contaminated	
M17	0(?) ok	87, 181	0+/268	0, 4, 5, 9 = 21	0+/289	13, 21, 29, 33, 27 = 123	0+/391
				controls ok on 2nd X		×2196 gal+ : 62, 63, 63, 57, 58	

ok = check

SUMMARY 17 segregants.

by transd. test 17/17 gal,-

by donor test 4/4 gal,-

by cross gal, 4/4 gal,- with following mix. prototrophs (332, 383, 511, 268)

source and number

(1913)

4/25/53 Sat. 750 prototroph t 902

Experiment N 750t902 → (-) agr Examination - Plated as 750tK12(-)Xg p. 1924

Segregant	No. spont. /g (½ plate)	No. prot 750t-2 spot	No. prot 902t-4 spot	Probable Nacc.	Scalate	Test cross ± 25% or control
1.	0	0	20	gal1-	0	ok ✓
2.	1.	0	24	1-	0	
3.	4	c. 50	0	2-	0	
4.	3	0	32	1-	0	ok ✓
5.	3	48	0	2-	0	
6.	0	0	14	1-	++	
7.	0	0	19	1-	0	ok ✓
8.	0	0	25	1-	±	
9.	5	45	1	2-	0	OK SEE BELOW
✓ 10.	0 (2)	1 (2) many small	14 17	1-	±	unst. ? ok unstab. + ? + confirmed on ½ duct
✓ 11.	1	0	26	1-	0	ok ✓
12	4	0	26	1-	0	
13.	0 ✗	0 ✗	13 ✗	✗	0	
14.	7 ✗	0 ✗	16 ✗	2< 1-	0	
15.	9	60	0	2-	0	
✓ 16.	4 (12)	53 (98)	4 (27)	2-	±	unst ? ok unstability ? + confirmed each duct SEE BELOW
17.	0	7	22	1-	0	
18.	3	1	13	1-	+	
19.	0	0	33	1-	±	
20.	3	0	25	1-	0	
21.	0	0	12	1-	0	
22.	0	1	14	1-	0	
23.	0	0	23	1-	0	
						750t ↑ = ok
				5 gal2-	21	
				18 gal1-	4±	
					170	

( ) = 4/26/53

5/4/53 → Spgt tests of Segregants (spgt 5/2e about)

Activity of Lysates of These Segregants

Crosses			/750	/518	/2175
See well mixed	gal2- agr X 902S on EMB gal SM				
controls	1. N3 (no + prot. observed)	(-) = 679, 582, 531, 443 = 2135	ok/2135	78	52	0
on EMBgal SM - no growth	2. N5 (")	(-) = 369, 280, 274, 359 = 1283	ok/1283	79	60	1
on EMBgal - all (-)	3. N9 (")	(-) = 363, 477, 419, 540 = 1799	ok/1799	41	46	1
	4. N15 (")	(-) = 454, 529, 479, 466 = 1928	ok/1928	44	43	0
	5. N16 (")	(-) = 638, 530, 535, 655 = 2358	ok/2358	raised vs appear. (well. T)	raised vs appear. (well c. 37 gaps	2
N16 → λ_2^S (5/9/53)		Recovered no (-) = (c.1500) 6 = c.9000		control area 1/67 plate = 18	control area c. 16	control area = 1

5/4 N10 streaked out (i.e. picked) → spgt tested → regal duced → /750l = 0 /811 = 4 /902 = 16 - control area 1

transductions confirmed here by brk. resm.

5/19/53 N16 purified (not transduce 811) 5/8

N16 " " " " /750l = 0 /811 = 3 /902 = 0 ^ control area = 0 Reactels! double(-)?

N16 " " " " /750l = 0 /811 = 0 /902 = 0 double (-)? or tp_2^r?

5/8 Crosses: ? gal- agr (-) X 750S (no prot observed)

				state of cross tst
1.	N1	31, 26, 34, 23 = ot/114	8, 3, 6, 1, = 18	ot/132
2.	N4	21, 28, 11, 28 = ot/88	7, 4, 1, 3 = 15	ot/103
3.	N7	39, 46, 38 35 = ot/178	5, 3, 7, 8 = 23	ot/201
4.	~~N10~~	~~50, 33, 41, 52 = ot/176~~		
5	N11	32, 20, 13, 40 = ot/105	5, 3, 1, 1; 10	
6.	N16	37, 33, 25, 44 = ot/139	3, 2, 2, 2 = 11	ot/150 ot/160

skip

control ok.

Activity of gal- lysates 5/9/53

	/750 spotted	/2175 spotted / solid smear	/811 spotted / solid smear	Does this indicate gal- ? ? or not?	811 1-100/750 spot ÷ bkgrd.	811 1-100/2175 spot / ½ = 92 = 736	811 1-100/811 spot c. 500	811 1-100/gal- spot c. 300
N1	0				—	—	—	—
N4	0	15	15		= bkgrd	½ = 78 : 580	c. 500	c. 300
N7	0	solid smear	solid smear		—	—	—	—
N11	0	11	11		control a little high	control 19	control 27	control 19
	control = 0	control = 1	control = 2					

Expt. N continued.

N16 plate = original plate stock

5/26/53 N16 - single colony isolation - Hershfield referred to as N-16 original or pure stock as

lysates (pure / original)

"original"	restreaked original	ancestor
/150	0 (?)	0
/2175	0	2
/2238	c. 10	c. 10 ← not likely to be same as 2238 lysate
/891	c. 7	c. 7

N16 original	transd	ancestor	← this lysate also fails to transd 1405 → (-)
/750 λ	0	0	1/14/53
/902 λ	0	1	residuum
/2238	0	4	N16 titer now is 1-2.7
/891	0	4	
/K-12 λ	0	4	
/842 gal+	solid smear	4	

N16 "plate" - because s.c. of N16 plate suspension as lysate gave no transd. and was not transd by anything but 842 gal+ (This repeated) - Lysate of plate inoculation was made.

N16 "plate" ½ stock

/891	solid smear ← stock of this showed +, gave tonic growth 0(0)	control: c. 5	hi ael. reconfirmed.
/2175	0	1	

Additional crosses gal$_1^-$ × 2257

N7	1 plate = 1432 × 4 = c. 4200	(-) no (+)	
N4	1 plate = 844 × 4 = c. 3376	(-) no (+)	
N1	1 plate = 2676 × 4 = c. 10,500	(-) no (+)	
N11	1 plate = 1640 × 4 = c. 6560	(-) no (+)	

7/6

Additional crosses gal$_2^-$ × 2251

N3	1 plate = 2660 × 4 = >10640	total > 12,775
N5	1 plate = 994 × 4 = >3976	total 5359
N7	1 plate = 1484 × 4 = >5936	total = >7735
N15	1 plate = 968 4 = >3872	total > 5900

Gal+ reversants obtained of

N1 - 1 on HFT (811)
- 2

N7 - 1 on HFT
- 2
- 3

5/16/53 N16 lysates of 30', 40', 60' exposure dont show a great deal of diff. All HFT

Monday 4/27/53

750 prototrophs x 1436 on EMB gal SM

1.
2.
3.
4. } all plates barren - Control streaks on EMB gal show pure (-)

1924 transduced $\bar{C}$ 01 λ - Observation to see if any plaques present - an indication of the new phage in lysates of 892 gal+ a mix

1. no add 1f
2. 01λ-1 (10^{-4} dil.) 1/4=64. ~~256~~ 238 $\times 10 \times 10^4$. 2.38×10^7 transductants/ml lysate
-1f

2238 x 1655 - gal$_3$ - 4p$_2$ prototrophs.

1. no add 64
2. 750λ-2 199
3. 902λ-4 123
4. 811λ-8 156

2343
750 prototrophic gal$_1$

1. No add 5
2. 811λ-8 8

W 2319
750 S^r =

1. no add 29 a suppressors? - high for gal$_1^-$ - why the change?
2. 811λ-8 128

750

1. no add 3
2. 811λ-8 10

4/24/53 Friday -

Lysate checks: On single plates - 0.05 ml lysate added (spots)

518	1. no add sector	2	
	2. 2238 - 1 spot	4	grow lysate - aeration stopped intermediate - may contain new phage action on 4p+
	3. 2175-4 spot	109	
	4. 750pr "	3	
	5. 750sr "	3	
750.	1. no add sector	0	
	2. 750pr spot	1	
	3. 750sr "	0	
	4. 2175-4 "	148	
	5. 2238 "	2	
2175	1. no add sector	4	
	2. 2238 spot	7	
	3. 2175 "	0	
	4. 750pr	10	
	5. 750sr	16	

~~= W2314~~

Cal 4 - gal4-pr+ x 750sr on EMS gal SM both F+ in cross?

1. Control resp. - no growth EMS gal SM
   - pure (-) on EMB gal

X plate (-)	(+)	total	
1. 5	0	5	
2. 12	0	12	
3. 21	0	21	0/54
4. 16	0	16	
		54	

~~= W2314~~

750pr x 750sr (both probably F+ in cross
EMS gal SM

1. Controls - no growth on both EMS gal SM
   - both (-) on EMB gal

2. X plate.

(+)	(-)	total	
0	42	42	
0	35	35	0/140 ?
0	30	30	
1?	32	33	
		140	

↑
Hm ✓

5/2/53 Saturday

750t 811 - (-) seg 1 Examination

1. no add - 7
2. 750d-2 4
3. 811d-8 10

} no decision possible -

750pr - For the purpose of making transductions of 750 by gal- to observe segregant (-)

1. no add 3
2. 811d-8 12 - pick and look for unstable

811 - For the purpose of making transductions gal- by gal, - to obs. segregant (-)

1. no add 51
2. 750d-2 96 - pick and look for unstable no unstables observed

5/4/53 Monday -

750pr - For the purpose of making transductions of 750 by gal+

Spread plates

1. no add 3
2. 811d-8 3
3. 811d-8 2
4. 811d-8 1

} again / no success

5/2 → 5/7 confusion about gal EMB - due to new mix

2175 transductions test partly modified by part of medium

17/72 gal seg carried out + on und. assuming of these observed are to be gal -

5/7/53 gal₂-

Expt. 0 2175 tk-12 gal₂ (-) segregants.

Segregant	No. spread 902A spot	No. prop. 902A spot	No. prop. 8114-8 spot	Probable locus
01	1	0	11	2-
02	0	0	4	2-
03	contaminated			—
" 4	10 0	1 1	13 12	2-
" 5	6 0	1 0	17 4	gal₂-
" 6	1 0	0 0	12 18	2-
" 7	11 0	0 0	13 14	2-
" 8	8 0	1 1	13 6	2-
" 9	← not spread →			—
" 10	2	1	12	2-
" 11	1	0	13	2-
" 12	0	0	6	2-
" 13	0	0	15	2-
" 14	0	0	4	2-
" 15	0	0	9	2-
" 16	0	0	10	2-
" 17	1	0	9	2-
" 18	0	0	10	2-
" 19	0	0	11	2-
" 20	0	1	7	2-
" 21	0	0	13	2-
" 22	0	1	10	2-
				20/20 2-

Tet Crosses - (-) Seg X (2251) 902A sp on EMS gal SM - control streaks on these EMB gal-OK

	total ptds	tracks (+)	2nd cross: tetr ptds	(+)	Grand ptds	Grand (+)	
03	162, 154, 89, 95, 130 = 630	0/630	604, 608, 567, 796, 814	0	4070	0	
05	139, 167, 162, 161, 128 = 757	0/757	1248, 1064, 914, 1092, 1016	0	5384	0	
011	64, 78, 67, 47, 77 = 336	0/335	237, 228, 400, 416, 456, 8	0	2072	0	now 0/4235
014	278, 201, 242, 318, 284, 73 = 1323	0/1323	1089, 1328, 1382, 644, 1116	0	6988	0	
					22 677		

011 X 2251 - 2nd cross
569, 861, 733 = 0/1563

Segregants	1811	2175
03	c. 15	0
05	c. 50	0
011	c. 50	0
014	c. 30	0
no add	2	0

2/24/54

Strains of this experiment discarded as not worth saving

5/9/53 Sat.

Various 'ductins for the purpose of obtaining seq. for classification

2175

1. mixed 16, 15
2. 811 + 8 32 — 3/24 unstable after 2 streakings
3. 750/1-2 50 — 4/21 unstable after 2

Hfiad. 1 - Anti serum test.

Ductin 1 = 01 - dil 1-100 of 01-1 c. 10^{10}/ml

1-100 = 10^8/ml (Re-test) → phage assay - dil 10^1, 10^2, 10^2 0.1ml (10^6 dil) = 35,24 = 2.95×10^7

+ { 0.5 ml; 0.5 ml 1-100 Anti P1; incubate 37C for 10 minutes } → post-test { phage assay - dil 1/10, 1/5, 10^2 0.1ml (10^5 dil) = 206, 254, = 2.30×10^7

'ductin' assay - 0.1ml → 1/2 = 272 2176

'ductin' assay - 0.1ml → 1/4 = 213 852

2176 · 10 · 10 · 10^2

2.2×10^9

to equate to pre cmc mult. X2 = 1704

Fract phage survivors = $\frac{2.30 \times 10^7}{2.95 \times 10^7}$ = 0.78

Fract phage survivors = $\frac{1704}{2176}$ = 0.78

$\frac{2.2 \times 10^7}{2.95 \times 10^9}$ = $\frac{1}{1.3 \times 10^3}$

Stocks of 2/24/54 this experiment (198) discard.

5/14/53 Source: duction: Pl 191 (w. para thiop at...)

Expt. R- 5/8 t 902 (-) seq. tested by spotting ~ 703λ-4, 841-P.

Seq	Control spot area	902 spot	811 spot	Platable factors	
R 1.	0	0	13	2-	✓
2.	1	7	1	4-	—
3.	0	10	0	4-	—
4.	0	1	9	2-	✓
5.	0	14	0	4-	—
6.	0	4	2	4-? 1pt.	not gal⁻ see below
7.	contaminated c (+)				
8.	1	10	1	4-	—
9.	1	11	1	4-	—
10.	contaminated with (+)				
11.	0	13	0	4-	—
12.	0	17	1	4-	—
13.	2	11	0	4-	—
14.	0	14	0	4-	—
15.	1	20	1	4-	—
16.	2	15	2	4-	—
17.	1	0	14	2-	✓
18.	1	8	2	4-	—
19.	0	8	1	4-	—
20.	2	15	7	4-	
21.	11	12	1	4-	
22.	0	6	0	4-	
23.	0	18	0	4-	

3. 2-
19 4-

Lysates of Gal₂⁻ R1, R4, R17

	Spotted on 811	/2175
R1	c. 70/2	1/0
R4	c. 60/2	0/0
R17	c. 80/2	0/0

Lysate test. (looking for lic ant. & gal⁻ variety) Crosses. Gal₂⁻ X 2257

R2	18/1	0/4
R3	33/8	26/42
R5	83/8	34/42
R8	67/8	34/42
R9	16/1	0/4
R11	8/1	0/4
R12	17/1	2/4
R13	6/1	1/4
R14	14/1	2/4
R16	11/1	0/4
R18	5/1	2/4
R19	2/1	1/4
R 20	3/0	3/4
R21	7/0	1/4
R22	7/0	2/4
R23	5/0	0/4

In these cases no equal → + lysate pop/control pop

others spot test, no pop. sort / control pt.

7/21 R7 was HFT acquired 2275, 7580

lost in 2nd plate

1. controls R1, R6, R17, 2257 on EMB gal⁻ plates. only 2257 & R6 on EMB lactose
R1 = 104, 99, 117, 39, 110, 126 = 0+/595 } 0+/1188
7/6/53 2nd 368, 458, 348, 247
Ignore not included R6 = 147, 175 (225) 127, 108, 109 = 1+/8 91 } NOT gal₂⁻
2nd 152, 138, 318, 444 abs. normal 1436 c 5000
R17 = 83, 41, 76, 78, 113, 85 } 0+/
7/6/53 2nd 221, 107, 231, 152 1187

2. Gal₂⁻ X 1436 4/7 λ^S 8/8 stable on strk
7/6/53 R1 1+/1116, 2+/873, 2+/837, 2+/276 = 7+/3102 = 0.22

6/8 λ^S 8/8 stable on strk
R17 - 6+/1063, 892, 554, 3+/511, 1+/344 = 10+/4364 = 0.228%

3. Gal₂⁻ X 1436 - controls o.k.
R2 - 4 plates one crumbled - 696 total c. 0+/2786
R3-5 " " " = 538 " " 0+/2675
R5- " " " 697 " " 0+/3485
R8- " " " 368 " " 0+/1840 } 0+/5952
7/6/53 " " " 1028 0+/4112

4. Gal₂⁻ X 902 - controls
3+/3 stable on strk R2 885, 583, 1+/740, 1+/410, 1+/345 = 3+/3183 = 0.094%
3+/2 " " R3 800, 679, 1+/563, 1+/625, 804 = 2+/3471 = 0.057%
1+/1 + stable on strk R5 6/849, 2/1513, 4/776, 4/1441, 7/905 = 19+/5432 = 0.35%
1+/1 stable on strk R8, 188, 269, 369, 585, 1+/254 = 1+/1668 = 0.06%
20+/121-611 = 0.21%

(+) recombinants
12/15 λ^S

5/16/53 Saturday.

Effect of anti λ serum on hi-act λ -

antiserum = anti P1
hi-act λ = D1 dil 1-100

plaque assays against 1485
ductive assays against 750

Experimental

antiserum dil 1-10

1-100 dil D1-1

0.2 ml + 0.2 ml → incubate 10 min at 37°

PRE-ASSAYS

1-100 → ductive assay: 123, 139 → ave 131

1-10, 1-5, 1-100 → 0.1 ml → 313, 285 plaque → ave 299 plaque.

surviving plaque titer = 2.99×10^7

surviving ductive titer = $131 \times 10^5 \times 2 = 2.62 \times 10^5$

plaque assay dil 1-100, 1-100, 1-10, → 0.1 ml → 75, 63 plaque → ave 69

titer 1-10 dil = 6.9×10^7 /ml

ductive titer → 0.1 ml : 709, 608 ductive → ave 6.58×10^5 ductive/ml

6.58 duct/ml / 6.9 plaque/ml = 1 ductive / 100 plaque

Plaque survival = $\frac{2.99}{6.90} = 0.433$

ductive survival = $\frac{2.62}{6.58} = 0.398$

---

750 × 902 to assay unit of recombination between parental suspensions of ca on ca 13 gal

Cross plate.

1.	296 (-) : c. 300	1 (+)
2.	c. 300	0
3.	c. 300	2 (+)
4.	"	0
5.	"	0
	c. 1500	2

↖ a minimum no. since there were many small (+) appearing colonies not counted

2/1500 = c. 0.13 %

Recombination between the various gal -

Ref. above		% Recomb	no. particles
	1. gal$_1^-$ × gal$_2^-$	0.13	c 1500
	gal$_1^+$ × gal$_2^+$	0.13	c. 1600
p. 171, 174, 175	2. gal$_4^-$ × gal$_2^-$		
	578 × "	0.38	1289
	811 × "	0.07	1365
	3. gal$_4^-$ × gal$_1^-$	< 1.9	54
	1486 × 750	0.83	4588
	4. gal$_3^-$ × gal$_1^+$		

0.3 - 0.6

5/18/53 Monday.

81 vs lysate of 2277 2278 - cultures of 1485 lysogenic for 892 1

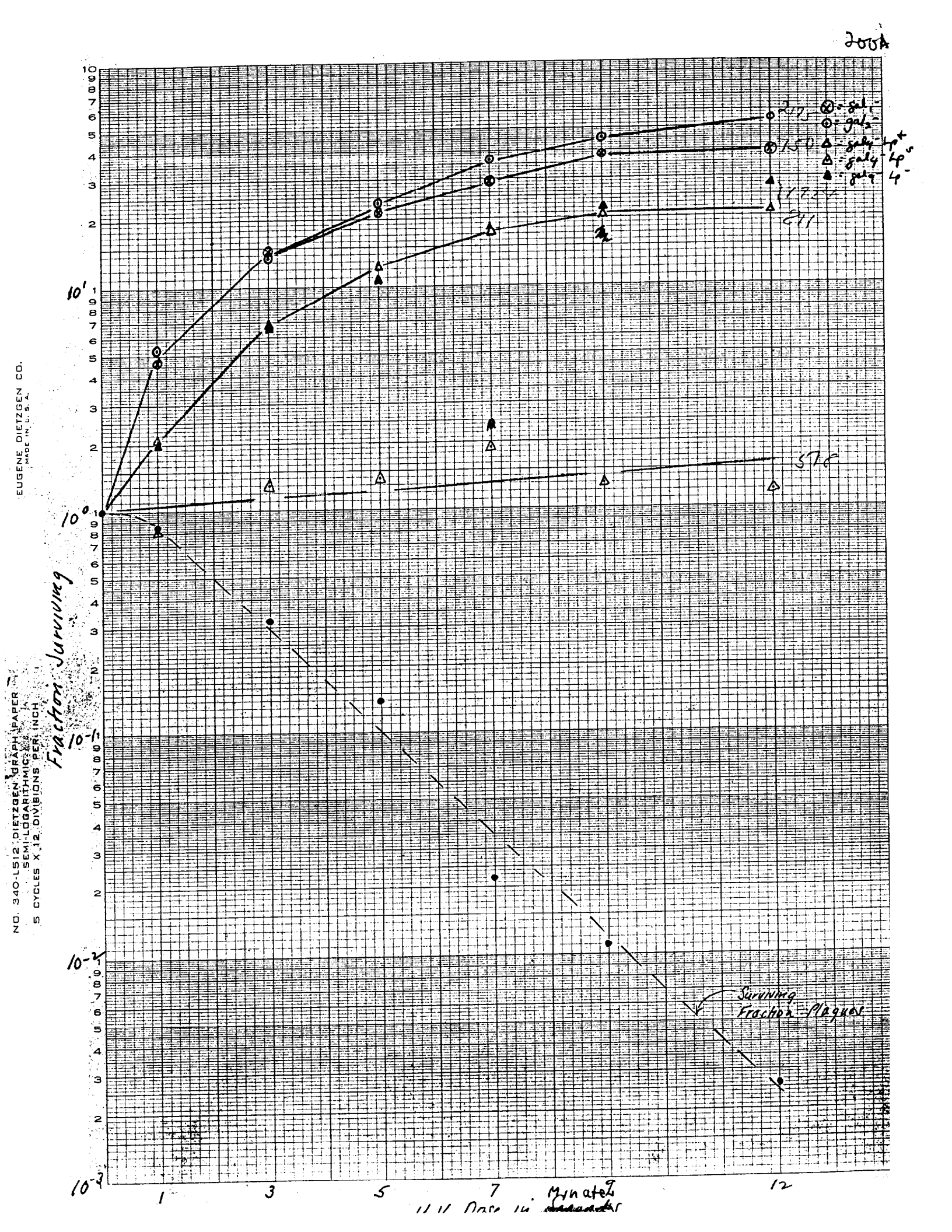
1.	no add	42	
2.	2277	52	plaque $c\ 10^3$
3.	2278	6	many plaques $c\ 10^4$

5/25/53 Monday.

750 X 2238 to obtain double mixed ppt. controls ok.

	(+)	titer
1.	2	c. 400
2.	0	c. 400
3.	0	c. 400
4.	0	c. 400
	2	1600

$\frac{0.00125}{1.}$ = 0.13%



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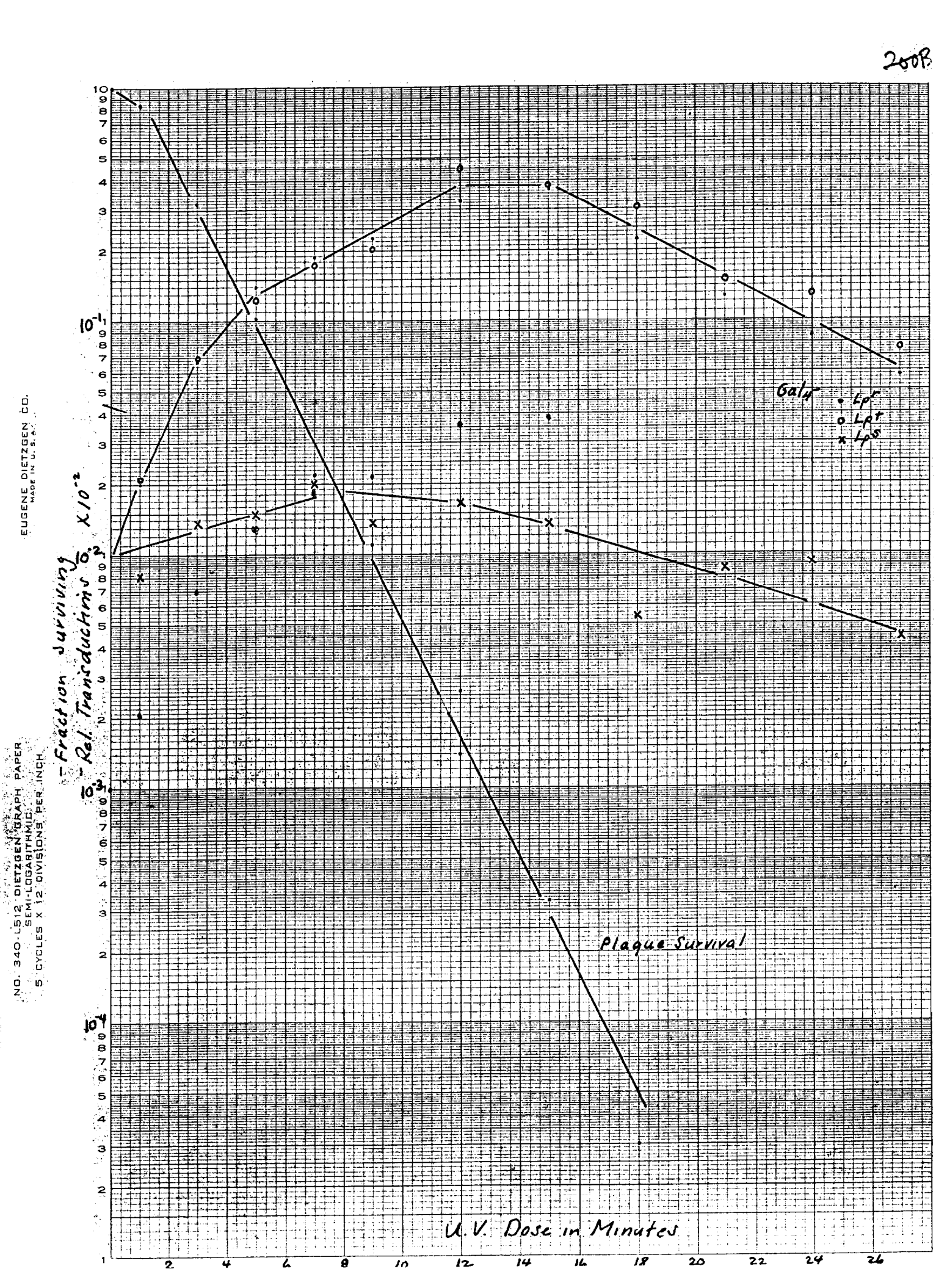
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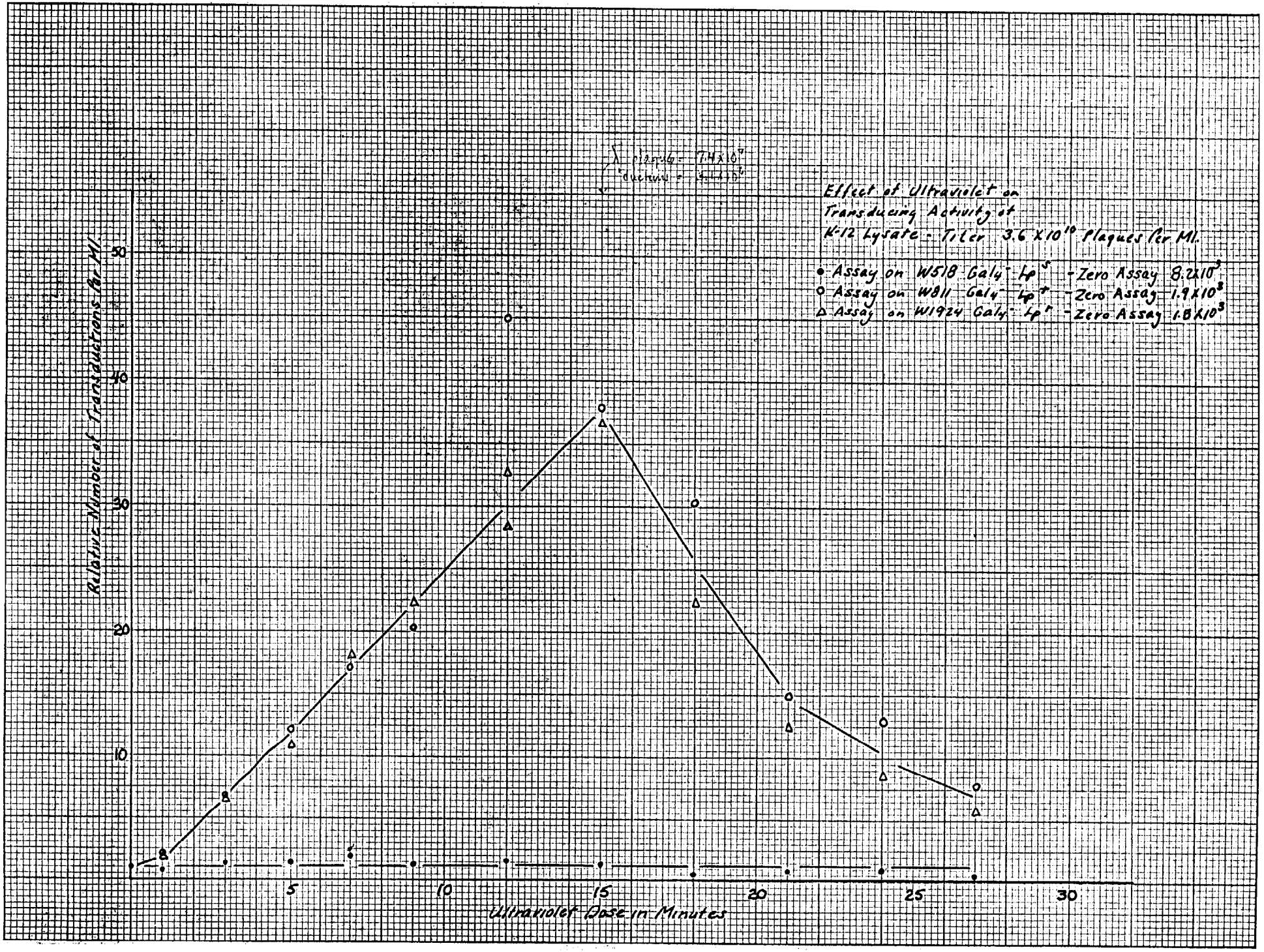

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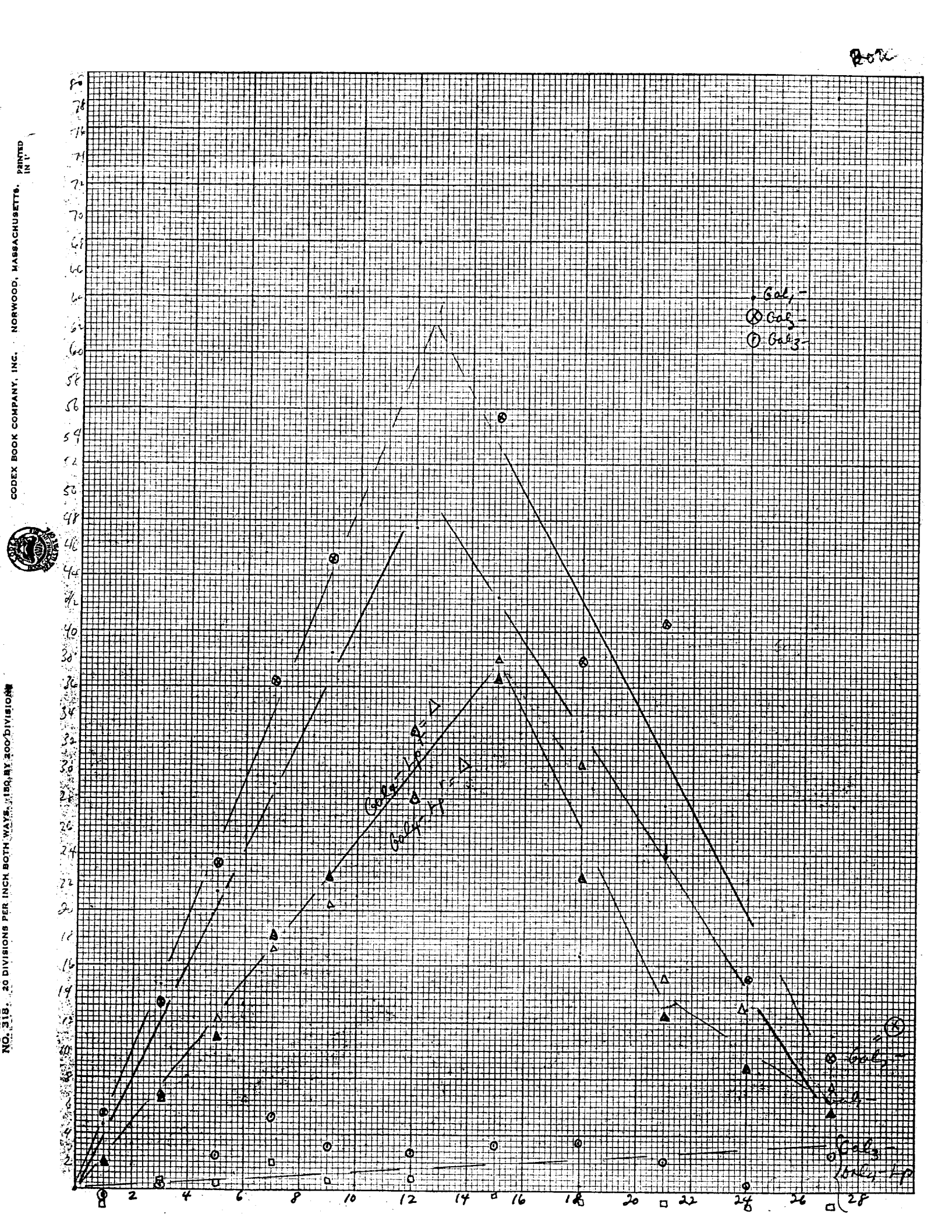

Lysate K12 5/30

Inhibition of pupillar/rel lysate

Irradiation of K-12 λ - Effect on Inhibition

Irradiation	W750 +λ	W750 -bkgrd	W750 ratio	W2175 +λ	W2175 -bkgrd	W2175 ratio	(1-10 die) W2297 +λ	W2297 -bkgrd	W2297 ratio
control-blk	3 (4)	—	—	9 (14)	—	—	6 (12)	—	—
control + 0 dose	89	86	1.0	83	74	1.0	43	370	1.0
1 min	410	407	4.7	401	392	5.3	39	330	0.89
3 "	1257	1254	14.5	1006	997	13.4	49	480	1.3
5 "	2034	2031	21.2	1641	1632	23.4	82	760	2.3
7 "	2512	2509	29.1	2706	2697	36.4	185	1790	5.4
9 "	3306	3303	38.4	3356	3347	45.2	109	1080	3.1
12 "	3420 (1/10 dil = 413)	4100	47.6	4058 (1/10 = 604)	6011	81.4	97	910	2.7
15 "	368	3640	42.3	424	4100	55.5	119	1070	3.3
18 " 1/10	277	2730	32.9	294	2800	37.9	131	1190	3.6
21 " dil.	243	2390	27.8 1/10	313	2990	40.5	82	700	2.1
24 "	116	1120	13.0 dil	115	1100	14.9	58	460	1.4
27 "	55	510	6.0	84	700	9.7	102	900	2.7

(cf W518 Lpt)

Irradiation	+λ	-bkgrd	ratio
control blk	19 (41)	—	—
control + 0 dose	835	816	1.0
1 min	686	667	0.82
3 min	1118	1099	1.35
5 " 1/2	1182	1163	1.4
7 " 1/4	1680	1661	2.04
9 " 1/8	1108	1108	1.36
12 " 1/16 dil = 1060	1370	1150-41=1109 / 750-41=709	1.6 / 1.36
15 " 1/32	115	450-41=409 / 740-41=699	0.84 / 0.86
18 " 1/64	48 (105 plaque)	740-41=699	0.86
21 " 1/128	74 (2.3×10⁴ plaque)	790-41=749	0.92
24 " dil	79 "	400-41=359	0.44
27 " 1/256	40 —		

W811 Lpt

Irradiation	+λ	-bkgrd	ratio
	32 (36)	—	—
	226	194	1.0
	443	411	2.1
	1369	1337	6.9
	2409	2377	12.2
	3384	3352	17.2
	3977 (1/10 = 878)	3945	20.3
	7036	8784	45.0
	776	7400	38.0
	626	5900	30.4
	328	2920	15.0
	290	2540	13.0
	186	1500	7.7

W1924 Lpt

Irradiation	+λ	-bkgrd	ratio
	44 (49)	—	—
	223	179	1.0
	392	358	2.0
	1264	1220	6.7
	1996	1952	10.9
	3296	3252	18.1
1/20 dil = 4056	4056	4012	22.4
1/40 dil = 5144 (49)	5144 (49)	5700	32.4
1/80	330 6600	6551	36.6
	201 4020	3971	22.2
	114 2280	2231	12.5
	79 1580	1531	8.6
	54 1080	1031	5.8

$\frac{1300 \cdot 10 \text{ d/ml}}{1.8 \times 10^3 \lambda/\mu\text{l}} = \frac{1.3 \times 10^4}{1.8 \times 10^3} = \frac{13}{1.8}$

at 12

$\frac{8784 \times 10 \text{ d/ml}}{1.8 \times 10^3 \lambda/\mu\text{l}} = \frac{8.7 \times 10^4}{1.8} = \frac{87}{1.8}$

8784
4036
12760 = 6380
2

226 | 6380 (25) — 502 — 1360

226 | 4036 (17) — 226 — 1770 — 1582

200E

K-12λ 5'30

Irradiation of K-12λ - plaque survival

Dose	dilutions	plaque counts	titer	survival
0	$10^7 \xrightarrow{0.1}$	357, 370	364×10^{10}	1.0
1	$10^7 \rightarrow$	337, 282	3.10×10^{10}	8.5×10^{-1}
3	$10^7 \rightarrow$	122, 114	1.18×10^{10}	3.2×10^{-1}
5	$10^6 \rightarrow$	548, 506	5.27×10^9	1.4×10^{-1}
7	$10^5 \rightarrow$	853, 749	8.00×10^8	2.2×10^{-2}
9	$10^5 \rightarrow$	202, 612	4.07×10^8	1.1×10^{-2}
diluted 1-10	$10^4 \rightarrow$	1033, 897	9.65×10^7	2.6×10^{-3}
	$10^5 \rightarrow$	69, 36	5.3×10^7	1.4×10^{-3}
15	$10^3 \rightarrow$	103, 140	1.22×10^7	3.3×10^{-4}
18	$10^3 \rightarrow$	18, 3	1.1×10^6	3.0×10^{-5}
21	10^2	6, 6	6.0×10^4	1.4×10^{-6}
24	10^2	0		
27	10^2	0		